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COMPOSITIONS AND METHODS FOR THE TREATMENT OF TUMOR

Field of the Invention

The present invention relates to compositions and methods for the diagnosis and treatment of tumor.

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Background of the Invention

Malignant tumors (cancers) are the second leading cause of death in the United States, after heart disease (Boring *et al.*, CA Cancer J. Clin., 43:7 [1993]).

15 Cancer is characterized by an increase in the number of abnormal, or neoplastic cells derived from a normal tissue which proliferate to form a tumor mass, the invasion of adjacent tissues by these neoplastic tumor cells, and the generation of malignant cells which eventually spread via the blood or lymphatic system to regional lymph nodes and to distant sites (metastasis). In a cancerous state, a cell proliferates under conditions in which normal cells would not grow. Cancer manifests itself in a wide variety of forms, characterized by different degrees of invasiveness and aggressiveness.

20 Alteration of gene expression is intimately related to the uncontrolled cell growth and de-differentiation which are a common feature of all cancers. The genomes of certain well studied tumors have been found to show decreased expression of recessive genes, usually referred to as tumor suppression genes, which would normally function to prevent malignant cell growth, and/or overexpression of certain dominant genes, such as oncogenes, that act to promote malignant growth. Each of these genetic changes appears to be responsible for importing some of
25 the traits that, in aggregate, represent the full neoplastic phenotype (Hunter, Cell, 64:1129 [1991] and Bishop, Cell, 64:235-248 [1991]).

A well known mechanism of gene (*e.g.* oncogene) overexpression in cancer cells is gene amplification. This is a process where in the chromosome of the ancestral cell multiple copies of a particular gene are produced. The process involves unscheduled replication of the region of chromosome comprising the gene, followed by
30 recombination of the replicated segments back into the chromosome (Alitalo *et al.*, Adv. Cancer Res., 47:235-281 [1986]). It is believed that the overexpression of the gene parallels gene amplification, *i.e.* is proportionate to the number of copies made.

Proto-oncogenes that encode growth factors and growth factor receptors have been identified to play important roles in the pathogenesis of various human malignancies, including breast cancer. For example, it has been
35 found that the human ErbB2 gene (erbB2, also known as her2, or c-erbB-2), which encodes a 185-kd transmembrane glycoprotein receptor (p185^{HER2}; HER2) related to the epidermal growth factor receptor EGFR, is overexpressed in about 25% to 30% of human breast cancer (Slamon *et al.*, Science, 235:177-182 [1987]; Slamon *et al.*, Science, 244:707-712 [1989]).

It has been reported that gene amplification of a proto-oncogene is an event typically involved in the more malignant forms of cancer, and could act as a predictor of clinical outcome (Schwab *et al.*, Genes Chromosomes Cancer, 1:181-193 [1990]; Alitalo *et al.*, *supra*). Thus, *erbB2* overexpression is commonly regarded as a predictor of a poor prognosis, especially in patients with primary disease that involves axillary lymph nodes (Slamon *et al.*, 5 [1987] and [1989], *supra*; Ravdin and Chamness, Gene, 159:19-27 [1995]; and Hynes and Stern, Biochim Biophys Acta, 1198:165-184 [1994]), and has been linked to sensitivity and/or resistance to hormone therapy and chemotherapeutic regimens, including CMF (cyclophosphamide, methotrexate, and fluoruracil) and anthracyclines (Baselga *et al.*, Oncology, 11 (3 Suppl 1):43-48 [1997]). However, despite the association of *erbB2* overexpression with poor prognosis, the odds of HER2-positive patients responding clinically to treatment with taxanes were greater 10 than three times those of HER2-negative patients (*Ibid*). A recombinant humanized anti-ErbB2 (anti-HER2) monoclonal antibody (a humanized version of the murine anti-ErbB2 antibody 4D5, referred to as rhuMAb HER2 or Herceptin™) has been clinically active in patients with ErbB2-overexpressing metastatic breast cancers that had received extensive prior anticancer therapy. (Baselga *et al.*, J. Clin. Oncol., 14:737-744 [1996]).

In light of the above, there is obvious interest in identifying novel methods and compositions which are 15 useful for diagnosing and treating tumors which are associated with gene amplification.

Summary of the Invention

The present invention concerns compositions and methods for the diagnosis and treatment of neoplastic cell growth and proliferation in mammals, including humans. The present invention is based on the identification of 20 genes that are amplified in the genome of tumor cells. Such gene amplification is expected to be associated with the overexpression of the gene product and contribute to tumorigenesis. Accordingly, the proteins encoded by the amplified genes are believed to be useful targets for the diagnosis and/or treatment (including prevention) of certain cancers, and may act as predictors of the prognosis of tumor treatment.

In one embodiment, the present invention concerns an isolated antibody which binds to a polypeptide 25 designated herein as PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Often, the cell that expresses the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, 30 PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. In one aspect, the isolated antibody specifically binds to a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. In another aspect, the antibody induces the 35 death of a cell which expresses a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Often, the cell that expresses the PRO212, PRO290, PRO341, PRO535, PRO619,

PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is a tumor cell that overexpresses the polypeptide as compared to a normal cell of the same type. In yet another aspect, the antibody is a monoclonal antibody, which preferably has
5 non-human complementarity determining region (CDR) residues and human framework region (FR) residues. The antibody may be labeled and may be immobilized on a solid support. In yet another aspect, the antibody is an antibody fragment, a single-chain antibody, or an anti-idiotypic antibody which binds, preferably specifically, to a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187,
10 PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

In another embodiment, the invention concerns a composition of matter which comprises an antibody which binds, preferably specifically, to a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145
15 or PRO2198 polypeptide in admixture with a pharmaceutically acceptable carrier. In one aspect, the composition of matter comprises a therapeutically effective amount of the antibody. In another aspect, the composition comprises a further active ingredient, which may, for example, be a further antibody or a cytotoxic or chemotherapeutic agent. Preferably, the composition is sterile.

In a further embodiment, the invention concerns an isolated nucleic acid molecule which encodes an anti-
20 PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody, and vectors and recombinant host cells comprising such nucleic acid molecules.

In a still further embodiment, the invention concerns a method for producing an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody, wherein the method
30 comprises culturing a host cell transformed with a nucleic acid molecule which encodes the antibody under conditions sufficient to allow expression of the antibody, and recovering the antibody from the cell culture.

The invention further concerns antagonists of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide that inhibit one or more of the functions or activities of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Agonists of a PRO212, PRO290,

PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide are also contemplated herein.

In a further embodiment, the invention concerns an isolated nucleic acid molecule that hybridizes to the complement of a nucleic acid molecule encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. The isolated nucleic acid molecule is preferably DNA, and hybridization preferably occurs under stringent hybridization and wash conditions. Such nucleic acid molecules can act as antisense molecules of the amplified genes identified herein, which, in turn, can find use in the modulation of the respective amplified genes, or as antisense primers in amplification reactions. Furthermore, such sequences can be used as part of a ribozyme and/or a triple helix sequence which, in turn, may be used in regulation of the amplified genes.

In another embodiment, the invention provides a method for determining the presence of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in a sample suspected of containing a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, wherein the method comprises exposing the sample to an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody and determining binding of the antibody to a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in the sample. In another embodiment, the invention provides a method for determining the presence of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in a cell, wherein the method comprises exposing the cell to an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody and determining binding of the antibody to the cell.

In yet another embodiment, the present invention concerns a method of diagnosing tumor in a mammal, comprising detecting the level of expression of a gene encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, 5 PRO2094, PRO2145 or PRO2198 polypeptide (a) in a test sample of tissue cells obtained from the mammal, and (b) in a control sample of known normal tissue cells of the same cell type, wherein a higher expression level in the test sample as compared to the control sample, is indicative of the presence of tumor in the mammal from which the test tissue cells were obtained.

In another embodiment, the present invention concerns a method of diagnosing tumor in a mammal, 10 comprising (a) contacting an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody with a test sample of tissue cells obtained from the mammal, and (b) 15 detecting the formation of a complex between the anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody and a PRO212, PRO290, PRO341, PRO535, PRO619, 20 PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in the test sample, wherein the formation of a complex is indicative of the presence of a tumor in said mammal. The detection may be qualitative or quantitative, and may be performed in comparison with monitoring the complex formation in a control sample of known normal tissue cells of the same 25 cell type. A larger quantity of complexes formed in the test sample indicates the presence of tumor in the mammal from which the test tissue cells were obtained. The antibody preferably carries a detectable label. Complex formation can be monitored, for example, by light microscopy, flow cytometry, fluorimetry, or other techniques known in the art.

The test sample is usually obtained from an individual suspected to have neoplastic cell growth or 30 proliferation (e.g. cancerous cells).

In another embodiment, the present invention concerns a cancer diagnostic kit comprising an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, 35 anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody and a carrier (e.g., a buffer) in suitable packaging. The kit preferably contains instructions for using the antibody to detect the presence of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184,

PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in a sample suspected of containing the same.

In yet another embodiment, the invention concerns a method for inhibiting the growth of tumor cells comprising exposing tumor cells which express a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide to an effective amount of an agent which inhibits an activity and/or the expression of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, wherein growth of the tumor cells is thereby inhibited. The agent preferably is an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody, a small organic and inorganic molecule, peptide, phosphopeptide, antisense or ribozyme molecule, or a triple helix molecule. In a specific aspect, the agent, *e.g.*, the anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody, induces cell death. In a further aspect, the tumor cells are further exposed to radiation treatment and/or a cytotoxic or chemotherapeutic agent.

In a further embodiment, the invention concerns an article of manufacture, comprising:

a container;

25 a label on the container; and

a composition comprising an active agent contained within the container; wherein the composition is effective for inhibiting the growth of tumor cells and the label on the container indicates that the composition can be used for treating conditions characterized by overexpression of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide as compared to a normal cell of the same tissue type. In particular aspects, the active agent in the composition is an agent which inhibits an activity and/or the expression of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. In preferred aspects, the active agent is an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-

PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody or an antisense oligonucleotide.

The invention also provides a method for identifying a compound that inhibits an activity of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, comprising contacting a candidate compound with a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide under conditions and for a time sufficient to allow these two components to interact and determining whether an activity of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is inhibited. In a specific aspect, either the candidate compound or the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is immobilized on a solid support. In another aspect, the non-immobilized component carries a detectable label. In a preferred aspect, this method comprises the steps of (a) contacting cells and a candidate compound to be screened in the presence of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide under conditions suitable for the induction of a cellular response normally induced by a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide and (b) determining the induction of said cellular response to determine if the test compound is an effective antagonist.

In another embodiment, the invention provides a method for identifying a compound that inhibits the expression of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in cells that express the polypeptide, wherein the method comprises contacting the cells with a candidate compound and determining whether the expression of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is inhibited. In a preferred aspect, this method comprises the steps of (a) contacting cells and a candidate compound to be screened under conditions suitable for allowing expression of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005,

PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide and (b) determining the inhibition of expression of said polypeptide.

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Brief Description of the Figures

Figure 1 shows the nucleotide sequence (SEQ ID NO:1) of a cDNA containing a nucleotide sequence encoding native sequence PRO212, wherein the nucleotide sequence (SEQ ID NO:1) is a clone designated herein as DNA30942-1134. Also presented in bold font and underlined are the positions of the respective start and stop codons.

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Figure 2 shows the amino acid sequence (SEQ ID NO:2) of a native sequence PRO212 polypeptide as derived from the coding sequence of SEQ ID NO:1.

Figures 3A through 3B show the nucleotide sequence (SEQ ID NO:6) of a cDNA containing a nucleotide sequence encoding native sequence PRO290, wherein the nucleotide sequence (SEQ ID NO:6) is a clone designated herein as DNA35680-1212. Also presented in bold font and underlined are the positions of the respective start and stop codons.

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Figure 4 shows the amino acid sequence (SEQ ID NO:7) of a native sequence PRO290 polypeptide as derived from the coding sequence of SEQ ID NO:6.

Figure 5 shows the nucleotide sequence (SEQ ID NO:9) of a cDNA containing a nucleotide sequence encoding native sequence PRO341, wherein the nucleotide sequence (SEQ ID NO:9) is a clone designated herein as DNA26288-1239. Also presented in bold font and underlined are the positions of the respective start and stop codons.

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Figure 6 shows the amino acid sequence (SEQ ID NO:10) of a native sequence PRO341 polypeptide as derived from the coding sequence of SEQ ID NO:9.

Figure 7 shows the nucleotide sequence (SEQ ID NO:13) of a cDNA containing a nucleotide sequence encoding native sequence PRO535, wherein the nucleotide sequence (SEQ ID NO:13) is a clone designated herein as DNA49143-1429. Also presented in bold font and underlined are the positions of the respective start and stop codons.

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Figure 8 shows the amino acid sequence (SEQ ID NO:14) of a native sequence PRO535 polypeptide as derived from the coding sequence of SEQ ID NO:13.

Figure 9 shows the nucleotide sequence (SEQ ID NO:15) of a cDNA containing a nucleotide sequence encoding native sequence PRO619, wherein the nucleotide sequence (SEQ ID NO:15) is a clone designated herein as DNA49821-1562. Also presented in bold font and underlined are the positions of the respective start and stop codons.

30

Figure 10 shows the amino acid sequence (SEQ ID NO:16) of a native sequence PRO619 polypeptide as derived from the coding sequence of SEQ ID NO:15.

35

Figure 11 shows the nucleotide sequence (SEQ ID NO:17) of a cDNA containing a nucleotide sequence encoding native sequence PRO717, wherein the nucleotide sequence (SEQ ID NO:17) is a clone designated herein as DNA50988-1326. Also presented in bold font and underlined are the positions of the respective start and stop

codons.

Figure 12 shows the amino acid sequence (SEQ ID NO:18) of a native sequence PRO717 polypeptide as derived from the coding sequence of SEQ ID NO:17.

Figure 13 shows the nucleotide sequence (SEQ ID NO:22) of a cDNA containing a nucleotide sequence encoding native sequence PRO809, wherein the nucleotide sequence (SEQ ID NO:22) is a clone designated herein as DNA57836-1338. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 14 shows the amino acid sequence (SEQ ID NO:23) of a native sequence PRO809 polypeptide as derived from the coding sequence of SEQ ID NO:22.

Figure 15 shows the nucleotide sequence (SEQ ID NO:24) of a cDNA containing a nucleotide sequence encoding native sequence PRO830, wherein the nucleotide sequence (SEQ ID NO:24) is a clone designated herein as DNA56866-1342. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 16 shows the amino acid sequence (SEQ ID NO:25) of a native sequence PRO830 polypeptide as derived from the coding sequence of SEQ ID NO:24.

Figure 17 shows the nucleotide sequence (SEQ ID NO:26) of a cDNA containing a nucleotide sequence encoding native sequence PRO848, wherein the nucleotide sequence (SEQ ID NO:26) is a clone designated herein as DNA59839-1461. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 18 shows the amino acid sequence (SEQ ID NO:27) of a native sequence PRO848 polypeptide as derived from the coding sequence of SEQ ID NO:26.

Figures 19A through 19B show the nucleotide sequence (SEQ ID NO:28) of a cDNA containing a nucleotide sequence encoding native sequence PRO943, wherein the nucleotide sequence (SEQ ID NO:28) is a clone designated herein as DNA52192-1369. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 20 shows the amino acid sequence (SEQ ID NO:29) of a native sequence PRO943 polypeptide as derived from the coding sequence of SEQ ID NO:28.

Figure 21 shows the nucleotide sequence (SEQ ID NO:33) of a cDNA containing a nucleotide sequence encoding native sequence PRO1005, wherein the nucleotide sequence (SEQ ID NO:33) is a clone designated herein as DNA57708-1411. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 22 shows the amino acid sequence (SEQ ID NO:34) of a native sequence PRO1005 polypeptide as derived from the coding sequence of SEQ ID NO:33.

Figure 23 show the nucleotide sequence (SEQ ID NO:35) of a cDNA containing a nucleotide sequence encoding native sequence PRO1009, wherein the nucleotide sequence (SEQ ID NO:35) is a clone designated herein as DNA57129-1413. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 24 shows the amino acid sequence (SEQ ID NO:36) of a native sequence PRO1009 polypeptide as

derived from the coding sequence of SEQ ID NO:35.

Figure 25 shows the nucleotide sequence (SEQ ID NO:37) of a cDNA containing a nucleotide sequence encoding native sequence PRO1025, wherein the nucleotide sequence (SEQ ID NO:37) is a clone designated herein as DNA59622-1334. Also presented in bold font and underlined are the positions of the respective start and stop
5 codons.

Figure 26 shows the amino acid sequence (SEQ ID NO:38) of a native sequence PRO1025 polypeptide as derived from the coding sequence of SEQ ID NO:37.

Figure 27 shows the nucleotide sequence (SEQ ID NO:39) of a cDNA containing a nucleotide sequence encoding native sequence PRO1030, wherein the nucleotide sequence (SEQ ID NO:39) is a clone designated herein
10 as DNA59485-1336. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 28 shows the amino acid sequence (SEQ ID NO:40) of a native sequence PRO1030 polypeptide as derived from the coding sequence of SEQ ID NO:39.

Figure 29 shows the nucleotide sequence (SEQ ID NO:41) of a cDNA containing a nucleotide sequence encoding native sequence PRO1097, wherein the nucleotide sequence (SEQ ID NO:41) is a clone designated herein
15 as DNA59841-1460. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 30 shows the amino acid sequence (SEQ ID NO:42) of a native sequence PRO1097 polypeptide as derived from the coding sequence of SEQ ID NO:41.

Figure 31 shows the nucleotide sequence (SEQ ID NO:43) of a cDNA containing a nucleotide sequence encoding native sequence PRO1107, wherein the nucleotide sequence (SEQ ID NO:43) is a clone designated herein
20 as DNA59606-1471. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 32 shows the amino acid sequence (SEQ ID NO:44) of a native sequence PRO1107 polypeptide as
25 derived from the coding sequence of SEQ ID NO:43.

Figure 33 shows the nucleotide sequence (SEQ ID NO:45) of a cDNA containing a nucleotide sequence encoding native sequence PRO1111, wherein the nucleotide sequence (SEQ ID NO:45) is a clone designated herein as DNA58721-1475. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 34 shows the amino acid sequence (SEQ ID NO:46) of a native sequence PRO1111 polypeptide as
30 derived from the coding sequence of SEQ ID NO:45.

Figure 35 shows the nucleotide sequence (SEQ ID NO:48) of a cDNA containing a nucleotide sequence encoding native sequence PRO1153, wherein the nucleotide sequence (SEQ ID NO:48) is a clone designated herein as DNA59842-1502. Also presented in bold font and underlined are the positions of the respective start and stop
35 codons.

Figure 36 shows the amino acid sequence (SEQ ID NO:49) of a native sequence PRO1153 polypeptide as derived from the coding sequence of SEQ ID NO:48.

Figure 37 shows the nucleotide sequence (SEQ ID NO:50) of a cDNA containing a nucleotide sequence

encoding native sequence PRO1182, wherein the nucleotide sequence (SEQ ID NO:50) is a clone designated herein as DNA59848-1512. Also presented in bold font and underlined are the positions of the respective start and stop codons.

5 Figure 38 shows the amino acid sequence (SEQ ID NO:51) of a native sequence PRO1182 polypeptide as derived from the coding sequence of SEQ ID NO:50.

Figure 39 shows the nucleotide sequence (SEQ ID NO:52) of a cDNA containing a nucleotide sequence encoding native sequence PRO1184, wherein the nucleotide sequence (SEQ ID NO:52) is a clone designated herein as DNA59220-1514. Also presented in bold font and underlined are the positions of the respective start and stop codons.

10 Figure 40 shows the amino acid sequence (SEQ ID NO:53) of a native sequence PRO1184 polypeptide as derived from the coding sequence of SEQ ID NO:52.

Figure 41 shows the nucleotide sequence (SEQ ID NO:54) of a cDNA containing a nucleotide sequence encoding native sequence PRO1187, wherein the nucleotide sequence (SEQ ID NO:54) is a clone designated herein as DNA62876-1517. Also presented in bold font and underlined are the positions of the respective start and stop
15 codons.

Figure 42 shows the amino acid sequence (SEQ ID NO:55) of a native sequence PRO1187 polypeptide as derived from the coding sequence of SEQ ID NO:54.

Figures 43A through 43B show the nucleotide sequence (SEQ ID NO:56) of a cDNA containing a nucleotide sequence encoding native sequence PRO1281, wherein the nucleotide sequence (SEQ ID NO:56) is a
20 clone designated herein as DNA59820-1549. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 44 shows the amino acid sequence (SEQ ID NO:57) of a native sequence PRO1281 polypeptide as derived from the coding sequence of SEQ ID NO:56.

Figure 45 shows the nucleotide sequence (SEQ ID NO:64) of a cDNA containing the nucleotide sequence
25 encoding native sequence PRO23, wherein the nucleotide sequence (SEQ ID NO:64) is a clone designated herein as DNA36640.

Figure 46 shows the amino acid sequence (SEQ ID NO:65) of a native sequence PRO23 polypeptide as derived from the coding sequence of SEQ ID NO:64. Also presented is a further truncated form of a native sequence PRO23 polypeptide.

30 Figure 47 shows the nucleotide sequence (SEQ ID NO:66) of a cDNA containing the nucleotide sequence encoding native sequence PRO39, wherein the nucleotide sequence (SEQ ID NO:66) is a clone designated as DNA36651.

Figure 48 shows the amino acid sequence (SEQ ID NO:67) of a native sequence PRO39 polypeptide as derived from the coding sequence of SEQ ID NO:66.

35 Figure 49 shows the nucleotide sequence (SEQ ID NO:68) of a cDNA containing the nucleotide sequence encoding native sequence PRO834, wherein the nucleotide sequence (SEQ ID NO:68) is a clone designated herein as DNA56538.

Figure 50 shows the amino acid sequence (SEQ ID NO:69) of a native sequence PRO834 polypeptide as

derived from the coding sequence of SEQ ID NO:68. Also presented is a pro-polypeptide of the PRO834 polypeptide.

Figures 51A through 51B show the nucleotide sequence (SEQ ID NO:70) of a cDNA containing the nucleotide sequence encoding native sequence PRO1317, wherein the nucleotide sequence (SEQ ID NO:70) is a clone designated herein as DNA71166. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 52 shows the amino acid sequence (SEQ ID NO:71) of a native sequence PRO1317 polypeptide as derived from the coding sequence of SEQ ID NO:70.

Figure 53 shows the nucleotide sequence (SEQ ID NO:72) of a cDNA containing the nucleotide sequence encoding native sequence PRO1710, wherein the nucleotide sequence (SEQ ID NO:72) is a clone designated herein as DNA82331. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 54 shows the amino acid sequence (SEQ ID NO:73) of a native sequence PRO1710 polypeptide as derived from the coding sequence of SEQ ID NO:72. Also presented is a pro-polypeptide of the PRO1710 polypeptide.

Figures 55A through 55B show the nucleotide sequence (SEQ ID NO:74) of a cDNA containing the nucleotide sequence encoding native sequence PRO2094, wherein the nucleotide sequence (SEQ ID NO:74) is a clone designated herein as DNA83123. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 56 shows the amino acid sequence (SEQ ID NO:75) of a native sequence PRO2094 polypeptide as derived from the coding sequence of SEQ ID NO:74.

Figure 57 shows the nucleotide sequence (SEQ ID NO:76) of a cDNA containing the nucleotide sequence encoding native sequence PRO2145, wherein the nucleotide sequence (SEQ ID NO:76) is a clone designated herein as DNA88050. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 58 shows the amino acid sequence (SEQ ID NO:77) of a native sequence PRO2145 polypeptide derived from the coding sequence of SEQ ID NO:76.

Figures 59A through 59B show the nucleotide sequence (SEQ ID NO:78) of a cDNA containing the nucleotide sequence encoding native sequence PRO2198, wherein the nucleotide sequence (SEQ ID NO:78) is a clone designated herein as DNA88153. Also presented in bold font and underlined are the positions of the respective start and stop codons.

Figure 60 shows the amino acid sequence (SEQ ID NO:79) of a native sequence PRO2198 polypeptide derived from the coding sequence of SEQ ID NO:78.

Figures 61A through 61D show hypothetical exemplifications for using the below described method to determine % amino acid sequence identity (Figures 61A-B) and % nucleic acid sequence identity (Figures 61C-D) using the ALIGN-2 sequence comparison computer program, wherein "PRO" represents the amino acid sequence of a hypothetical PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide of interest, "Comparison Protein" represents the amino acid sequence of a polypeptide against which

the "PRO" polypeptide of interest is being compared, "PRO-DNA" represents a hypothetical PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding
5 nucleic acid sequence of interest, "Comparison DNA" represents the nucleotide sequence of a nucleic acid molecule against which the "PRO-DNA" nucleic acid molecule of interest is being compared, "X", "Y" and "Z" each represent different hypothetical amino acid residues and "N", "L" and "V" each represent different hypothetical nucleotides.

Figures 62A through 62P provide the complete source code for the ALIGN-2 sequence comparison computer program. This source code may be routinely compiled for use on a UNIX operating system to provide the
10 ALIGN-2 sequence comparison computer program.

Figure 63 is a map of Chromosome 4 showing the mapping region of DNA50988-1326.

Figure 64 is a map of Chromosome 2 showing the mapping region of DNA57708-1411.

Figure 65 is a map of Chromosome 6 showing the mapping region of DNA83123.

Figure 66 is a map of Chromosome 16 showing the mapping region of DNA88153.

15

Detailed Description of the Invention

I. Definitions

The phrases "gene amplification" and "gene duplication" are used interchangeably and refer to a process by which multiple copies of a gene or gene fragment are formed in a particular cell or cell line. The duplicated
20 region (a stretch of amplified DNA) is often referred to as "amplicon." Usually, the amount of the messenger RNA (mRNA) produced, *i.e.*, the level of gene expression, also increases in the proportion of the number of copies made of the particular gene expressed.

"Tumor", as used herein, refers to all neoplastic cell growth and proliferation, whether malignant or benign, and all pre-cancerous and cancerous cells and tissues.

25 The terms "cancer" and "cancerous" refer to or describe the physiological condition in mammals that is typically characterized by unregulated cell growth. Examples of cancer include but are not limited to, carcinoma, lymphoma, blastoma, sarcoma, and leukemia. More particular examples of such cancers include breast cancer, prostate cancer, colon cancer, squamous cell cancer, small-cell lung cancer, non-small cell lung cancer, gastrointestinal cancer, pancreatic cancer, glioblastoma, cervical cancer, ovarian cancer, liver cancer, bladder cancer,
30 hepatoma, colorectal cancer, endometrial carcinoma, salivary gland carcinoma, kidney cancer, liver cancer, vulval cancer, thyroid cancer, hepatic carcinoma and various types of head and neck cancer.

"Treatment" is an intervention performed with the intention of preventing the development or altering the pathology of a disorder. Accordingly, "treatment" refers to both therapeutic treatment and prophylactic or preventative measures. Those in need of treatment include those already with the disorder as well as those in which
35 the disorder is to be prevented. In tumor (*e.g.*, cancer) treatment, a therapeutic agent may directly decrease the pathology of tumor cells, or render the tumor cells more susceptible to treatment by other therapeutic agents, *e.g.*, radiation and/or chemotherapy.

The "pathology" of cancer includes all phenomena that compromise the well-being of the patient. This

includes, without limitation, abnormal or uncontrollable cell growth, metastasis, interference with the normal functioning of neighboring cells, release of cytokines or other secretory products at abnormal levels, suppression or aggravation of inflammatory or immunological response, etc.

"Mammal" for purposes of treatment refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, sports, or pet animals, such as dogs, horses, cats, cattle, pigs, sheep, etc. Preferably, the mammal is human.

"Carriers" as used herein include pharmaceutically acceptable carriers, excipients, or stabilizers which are nontoxic to the cell or mammal being exposed thereto at the dosages and concentrations employed. Often the physiologically acceptable carrier is an aqueous pH buffered solution. Examples of physiologically acceptable carriers include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptide; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEENTM, polyethylene glycol (PEG), and PLURONICSTM.

Administration "in combination with" one or more further therapeutic agents includes simultaneous (concurrent) and consecutive administration in any order.

The term "cytotoxic agent" as used herein refers to a substance that inhibits or prevents the function of cells and/or causes destruction of cells. The term is intended to include radioactive isotopes (e.g., I¹³¹, I¹²⁵, Y⁹⁰ and Re¹⁸⁶), chemotherapeutic agents, and toxins such as enzymatically active toxins of bacterial, fungal, plant or animal origin, or fragments thereof.

A "chemotherapeutic agent" is a chemical compound useful in the treatment of cancer. Examples of chemotherapeutic agents include adriamycin, doxorubicin, epirubicin, 5-fluorouracil, cytosine arabinoside ("Ara-C"), cyclophosphamide, thiopeta, busulfan, cytoxin, taxoids, e.g., paclitaxel (Taxol, Bristol-Myers Squibb Oncology, Princeton, NJ), and doxetaxel (Taxotere, Rhône-Poulenc Rorer, Antony, France), taxotere, methotrexate, cisplatin, melphalan, vinblastine, bleomycin, etoposide, ifosfamide, mitomycin C, mitoxantrone, vincristine, vinorelbine, carboplatin, teniposide, daunomycin, carminomycin, aminopterin, dactinomycin, mitomycins, esperamicins (see U.S. Pat. No. 4,675,187), 5-FU, 6-thioguanine, 6-mercaptopurine, actinomycin D, VP-16, chlorambucil, melphalan, and other related nitrogen mustards. Also included in this definition are hormonal agents that act to regulate or inhibit hormone action on tumors such as tamoxifen and onapristone.

A "growth inhibitory agent" when used herein refers to a compound or composition which inhibits growth of a cell, especially cancer cell overexpressing any of the genes identified herein, either *in vitro* or *in vivo*. Thus, the growth inhibitory agent is one which significantly reduces the percentage of cells overexpressing such genes in S phase. Examples of growth inhibitory agents include agents that block cell cycle progression (at a place other than S phase), such as agents that induce G1 arrest and M-phase arrest. Classical M-phase blockers include the vincas (vincristine and vinblastine), taxol, and topo II inhibitors such as doxorubicin, epirubicin, daunorubicin, etoposide, and bleomycin. Those agents that arrest G1 also spill over into S-phase arrest, for example, DNA alkylating agents such as tamoxifen, prednisone, dacarbazine, mechlorethamine, cisplatin, methotrexate, 5-

fluorouracil, and ara-C. Further information can be found in *The Molecular Basis of Cancer*, Mendelsohn and Israel, eds., Chapter 1, entitled "Cell cycle regulation, oncogens, and antineoplastic drugs" by Murakami *et al.*, (WB Saunders: Philadelphia, 1995), especially p. 13.

"Doxorubicin" is an anthracycline antibiotic. The full chemical name of doxorubicin is (8S-cis)-10-[(3-amino-2,3,6-trideoxy- α -L-lyxo-hexapyranosyl)oxy]-7,8,9,10-tetrahydro-6,8,11-trihydroxy-8-(hydroxyacetyl)-1-methoxy-5,12-naphthacenedione.

The term "cytokine" is a generic term for proteins released by one cell population which act on another cell as intercellular mediators. Examples of such cytokines are lymphokines, monokines, and traditional polypeptide hormones. Included among the cytokines are growth hormone such as human growth hormone, N-methionyl human growth hormone, and bovine growth hormone; parathyroid hormone; thyroxine; insulin; proinsulin; relaxin; prolrelaxin; glycoprotein hormones such as follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH), and luteinizing hormone (LH); hepatic growth factor; fibroblast growth factor; prolactin; placental lactogen; tumor necrosis factor- α and - β ; mullerian-inhibiting substance; mouse gonadotropin-associated peptide; inhibin; activin; vascular endothelial growth factor; integrin; thrombopoietin (TPO); nerve growth factors such as NGF- β ; platelet-growth factor; transforming growth factors (TGFs) such as TGF- α and TGF- β ; insulin-like growth factor-I and -II; erythropoietin (EPO); osteoinductive factors; interferons such as interferon - α , - β , and - γ ; colony stimulating factors (CSFs) such as macrophage-CSF (M-CSF); granulocyte-macrophage-CSF (GM-CSF); and granulocyte-CSF (G-CSF); interleukins (ILs) such as IL-1, IL-1a, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-11, IL-12; a tumor necrosis factor such as TNF- α or TNF- β ; and other polypeptide factors including LIF and kit ligand (KL). As used herein, the term cytokine includes proteins from natural sources or from recombinant cell culture and biologically active equivalents of the native sequence cytokines.

The term "prodrug" as used in this application refers to a precursor or derivative form of a pharmaceutically active substance that is less cytotoxic to tumor cells compared to the parent drug and is capable of being enzymatically activated or converted into the more active parent form. See, *e.g.*, Wilman, "Prodrugs in Cancer Chemotherapy", *Biochemical Society Transactions*, 14:375-382, 615th Meeting, Belfast (1986), and Stella *et al.*, "Prodrugs: A Chemical Approach to Targeted Drug Delivery", *Directed Drug Delivery*, Borchardt *et al.*, (ed.), pp. 147-267, Humana Press (1985). The prodrugs of this invention include, but are not limited to, phosphate-containing prodrugs, thiophosphate-containing prodrugs, sulfate-containing prodrugs, peptide-containing prodrugs, D-amino acid-modified prodrugs, glycosylated prodrugs, β -lactam-containing prodrugs, optionally substituted phenoxyacetamide-containing prodrugs or optionally substituted phenylacetamide-containing prodrugs, 5-fluorocytosine and other 5-fluorouridine prodrugs which can be converted into the more active cytotoxic free drug. Examples of cytotoxic drugs that can be derivatized into a prodrugs form for use in this invention include, but are not limited to, those chemotherapeutic agents described above.

An "effective amount" of a polypeptide disclosed herein or an antagonist thereof, in reference to inhibition of neoplastic cell growth, tumor growth or cancer cell growth, is an amount capable of inhibiting, to some extent, the growth of target cells. The term includes an amount capable of invoking a growth inhibitory, cytostatic and/or cytotoxic effect and/or apoptosis of the target cells. An "effective amount" of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030,

PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide or an antagonist thereof for purposes of inhibiting neoplastic cell growth, tumor growth or cancer cell growth, may be determined empirically and in a routine manner.

5 A "therapeutically effective amount", in reference to the treatment of tumor, refers to an amount capable of invoking one or more of the following effects: (1) inhibition, to some extent, of tumor growth, including, slowing down and complete growth arrest; (2) reduction in the number of tumor cells; (3) reduction in tumor size; (4) inhibition (*i.e.*, reduction, slowing down or complete stopping) of tumor cell infiltration into peripheral organs; (5) inhibition (*i.e.*, reduction, slowing down or complete stopping) of metastasis; (6) enhancement of anti-tumor immune
10 response, which may, but does not have to, result in the regression or rejection of the tumor; and/or (7) relief, to some extent, of one or more symptoms associated with the disorder. A "therapeutically effective amount" of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide or an
15 antagonist thereof for purposes of treatment of tumor may be determined empirically and in a routine manner.

A "growth inhibitory amount" of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 antagonist is an amount capable of inhibiting the growth of a cell, especially tumor, *e.g.*,
20 cancer cell, either *in vitro* or *in vivo*. A "growth inhibitory amount" of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 antagonist for purposes of inhibiting neoplastic cell growth may be determined empirically and in a routine manner.

25 A "cytotoxic amount" of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 antagonist is an amount capable of causing the destruction of a cell, especially tumor, *e.g.*, cancer cell, either *in vitro* or *in vivo*. A "cytotoxic amount" of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717,
30 PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 antagonist for purposes of inhibiting neoplastic cell growth may be determined empirically and in a routine manner.

The terms "PRO212", "PRO290", "PRO341", "PRO535", "PRO619", "PRO717", "PRO809", "PRO830",
35 "PRO848", "PRO943", "PRO1005", "PRO1009", "PRO1025", "PRO1030", "PRO1097", "PRO1107", "PRO1111", "PRO1153", "PRO1182", "PRO1184", "PRO1187", "PRO1281", "PRO23", "PRO39", "PRO834", "PRO1317", "PRO1710", "PRO2094", "PRO2145", "PRO2198", "PRO212 polypeptide", "PRO290 polypeptide", "PRO341 polypeptide", "PRO535 polypeptide", "PRO619 polypeptide", "PRO717 polypeptide", "PRO809 polypeptide",

"PRO830 polypeptide", "PRO848 polypeptide", "PRO943 polypeptide", "PRO1005 polypeptide", "PRO1009 polypeptide", "PRO1025 polypeptide", "PRO1030 polypeptide", "PRO1097 polypeptide", "PRO1107 polypeptide", "PRO1111 polypeptide", "PRO1153 polypeptide", "PRO1182 polypeptide", "PRO1184 polypeptide", "PRO1187 polypeptide", "PRO1281 polypeptide", "PRO23 polypeptide", "PRO39 polypeptide", "PRO834 polypeptide",
5 "PRO1317 polypeptide", "PRO1710 polypeptide", "PRO2094 polypeptide", "PRO2145 polypeptide" or "PRO2198 polypeptide" when used herein encompass native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptides and PRO212, PRO290, PRO341, PRO535, PRO619, PRO717,
10 PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide variants (which are further defined herein). The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834,
15 PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide may be isolated from a variety of sources, such as from human tissue types or from another source, or prepared by recombinant and/or synthetic methods.

A "native sequence" PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145
20 or PRO2198 polypeptide comprises a polypeptide having the same amino acid sequence as a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide derived from nature. Such native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943,
25 PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide can be isolated from nature or can be produced by recombinant and/or synthetic means. The term "native sequence" PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184,
30 PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide specifically encompasses naturally-occurring truncated or secreted forms (e.g., an extracellular domain sequence), naturally-occurring variant forms (e.g., alternatively spliced forms) and naturally-occurring allelic variants of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187,
35 PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. In certain embodiments of the invention, the native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094,

PRO2145 or PRO2198 polypeptide is a mature or full-length native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide comprising the amino acid sequence of Figure 2 (SEQ ID NO:2), Figure 4 (SEQ ID NO:7), Figure 6 (SEQ ID NO:10), Figure 8 (SEQ ID NO:14), Figure 10 (SEQ ID NO:16), Figure 12 (SEQ ID NO:18), Figure 14 (SEQ ID NO:23), Figure 16 (SEQ ID NO:25), Figure 18 (SEQ ID NO:27), Figure 20 (SEQ ID NO:29), Figure 22 (SEQ ID NO:34), Figure 24 (SEQ ID NO:36), Figure 26 (SEQ ID NO:38), Figure 28 (SEQ ID NO:40), Figure 30 (SEQ ID NO:42), Figure 32 (SEQ ID NO:44), Figure 34 (SEQ ID NO:46), Figure 36 (SEQ ID NO:49), Figure 38 (SEQ ID NO:51), Figure 40 (SEQ ID NO:53), Figure 42 (SEQ ID NO:55), Figure 44 (SEQ ID NO:57), Figure 46 (SEQ ID NO:65), Figure 48 (SEQ ID NO:67), Figure 50 (SEQ ID NO:69), Figure 52 (SEQ ID NO:71), Figure 54 (SEQ ID NO:73), Figure 56 (SEQ ID NO:75), Figure 58 (SEQ ID NO:77) or Figure 60 (SEQ ID NO:79) respectively. Fragments of the respective native polypeptides herein include, but are not limited, to polypeptide variants from which the native N-terminal signal sequence has been fully or partially deleted or replaced by another sequence, and extracellular domains of the respective native sequences, regardless whether such truncated (secreted) forms occur in nature. Fragments are preferably sufficient in length for the production of an antibody specifically binding the corresponding native "PRO" polypeptide.

"PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polypeptide" means an active PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide as defined below having at least about 80% amino acid sequence identity with the amino acid sequence shown in Figure 2 (SEQ ID NO:2), Figure 4 (SEQ ID NO:7), Figure 6 (SEQ ID NO:10), Figure 8 (SEQ ID NO:14), Figure 10 (SEQ ID NO:16), Figure 12 (SEQ ID NO:18), Figure 14 (SEQ ID NO:23), Figure 16 (SEQ ID NO:25), Figure 18 (SEQ ID NO:27), Figure 20 (SEQ ID NO:29), Figure 22 (SEQ ID NO:34), Figure 24 (SEQ ID NO:36), Figure 26 (SEQ ID NO:38), Figure 28 (SEQ ID NO:40), Figure 30 (SEQ ID NO:42), Figure 32 (SEQ ID NO:44), Figure 34 (SEQ ID NO:46), Figure 36 (SEQ ID NO:49), Figure 38 (SEQ ID NO:51), Figure 40 (SEQ ID NO:53), Figure 42 (SEQ ID NO:55), Figure 44 (SEQ ID NO:57), Figure 46 (SEQ ID NO:65), Figure 48 (SEQ ID NO:67), Figure 50 (SEQ ID NO:69), Figure 52 (SEQ ID NO:71), Figure 54 (SEQ ID NO:73), Figure 56 (SEQ ID NO:75), Figure 58 (SEQ ID NO:77) or Figure 60 (SEQ ID NO:79) respectively, or a specifically derived fragment thereof. Such PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polypeptides include, for instance, PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptides wherein one or more amino acid residues are added, or deleted, at the N- and/or

C-terminus, as well as within one or more internal domains, of the sequence of Figure 2 (SEQ ID NO:2), Figure 4 (SEQ ID NO:7), Figure 6 (SEQ ID NO:10), Figure 8 (SEQ ID NO:14), Figure 10 (SEQ ID NO:16), Figure 12 (SEQ ID NO:18), Figure 14 (SEQ ID NO:23), Figure 16 (SEQ ID NO:25), Figure 18 (SEQ ID NO:27), Figure 20 (SEQ ID NO:29), Figure 22 (SEQ ID NO:34), Figure 24 (SEQ ID NO:36), Figure 26 (SEQ ID NO:38), Figure 28 (SEQ ID NO:40), Figure 30 (SEQ ID NO:42), Figure 32 (SEQ ID NO:44), Figure 34 (SEQ ID NO:46), Figure 36 (SEQ ID NO:49), Figure 38 (SEQ ID NO:51), Figure 40 (SEQ ID NO:53), Figure 42 (SEQ ID NO:55), Figure 44 (SEQ ID NO:57), Figure 46 (SEQ ID NO:65), Figure 48 (SEQ ID NO:67), Figure 50 (SEQ ID NO:69), Figure 52 (SEQ ID NO:71), Figure 54 (SEQ ID NO:73), Figure 56 (SEQ ID NO:75), Figure 58 (SEQ ID NO:77) or Figure 60 (SEQ ID NO:79) respectively. Ordinarily, a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polypeptide will have at least about 80% amino acid sequence identity, more preferably at least about 81% amino acid sequence identity, more preferably at least about 82% amino acid sequence identity, more preferably at least about 83% amino acid sequence identity, more preferably at least about 84% amino acid sequence identity, more preferably at least about 85% amino acid sequence identity, more preferably at least about 86% amino acid sequence identity, more preferably at least about 87% amino acid sequence identity, more preferably at least about 88% amino acid sequence identity, more preferably at least about 89% amino acid sequence identity, more preferably at least about 90% amino acid sequence identity, more preferably at least about 91% amino acid sequence identity, more preferably at least about 92% amino acid sequence identity, more preferably at least about 93% amino acid sequence identity, more preferably at least about 94% amino acid sequence identity, more preferably at least about 95% amino acid sequence identity, more preferably at least about 96% amino acid sequence identity, more preferably at least about 97% amino acid sequence identity, more preferably at least about 98% amino acid sequence identity and yet more preferably at least about 99% amino acid sequence identity with the amino acid sequence of Figure 2 (SEQ ID NO:2), Figure 4 (SEQ ID NO:7), Figure 6 (SEQ ID NO:10), Figure 8 (SEQ ID NO:14), Figure 10 (SEQ ID NO:16), Figure 12 (SEQ ID NO:18), Figure 14 (SEQ ID NO:23), Figure 16 (SEQ ID NO:25), Figure 18 (SEQ ID NO:27), Figure 20 (SEQ ID NO:29), Figure 22 (SEQ ID NO:34), Figure 24 (SEQ ID NO:36), Figure 26 (SEQ ID NO:38), Figure 28 (SEQ ID NO:40), Figure 30 (SEQ ID NO:42), Figure 32 (SEQ ID NO:44), Figure 34 (SEQ ID NO:46), Figure 36 (SEQ ID NO:49), Figure 38 (SEQ ID NO:51), Figure 40 (SEQ ID NO:53), Figure 42 (SEQ ID NO:55), Figure 44 (SEQ ID NO:57), Figure 46 (SEQ ID NO:65), Figure 48 (SEQ ID NO:67), Figure 50 (SEQ ID NO:69), Figure 52 (SEQ ID NO:71), Figure 54 (SEQ ID NO:73), Figure 56 (SEQ ID NO:75), Figure 58 (SEQ ID NO:77) or Figure 60 (SEQ ID NO:79) respectively, or a derived polypeptide fragment thereof. Variants do not encompass the native PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide sequence. Ordinarily, PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polypeptides are at least about 10 amino acids in length, often at least

about 20 amino acids in length, more often at least about 30 amino acids in length, more often at least about 40 amino acids in length, more often at least about 50 amino acids in length, more often at least about 60 amino acids in length, more often at least about 70 amino acids in length, more often at least about 80 amino acids in length, more often at least about 90 amino acids in length, more often about 100 amino acids in length, more often at least about 150 amino acids in length, more often at least about 200 amino acids in length, more often at least about 300 amino acids in length, or more.

"Percent (%) amino acid sequence identity" with respect to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide sequences identified herein is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN, ALIGN-2 or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full-length of the sequences being compared. For purposes herein, however, % amino acid sequence identity values are obtained as described below by using the sequence comparison computer program ALIGN-2, wherein the complete source code for the ALIGN-2 program is provided in Figures 62A-P. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc., and the source code shown in Figures 62A-P has been filed with user documentation in the U.S. Copyright Office, Washington D.C., 20559, where it is registered under U.S. Copyright Registration No. TXU510087. The ALIGN-2 program is publicly available through Genentech, Inc., South San Francisco, California or may be compiled from the source code provided in Figures 62A-P. The ALIGN-2 program should be compiled for use on a UNIX operating system, preferably digital UNIX V4.0D. All sequence comparison parameters are set by the ALIGN-2 program and do not vary.

For purposes herein, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) is calculated as follows:

100 times the fraction X/Y

where X is the number of amino acid residues scored as identical matches by the sequence alignment program ALIGN-2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It

will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A. As examples of % amino acid sequence identity calculations, Figures 61A-B demonstrate how to calculate the % amino acid sequence identity of the amino acid sequence designated "Comparison Protein" to the amino acid sequence designated "PRO".

Unless specifically stated otherwise, all % amino acid sequence identity values used herein are obtained as described above using the ALIGN-2 sequence comparison computer program. However, % amino acid sequence identity may also be determined using the sequence comparison program NCBI-BLAST2 (Altschul *et al.*, Nucleic Acids Res., 25:3389-3402 (1997)). The NCBI-BLAST2 sequence comparison program may be downloaded from <http://www.ncbi.nlm.nih.gov>. NCBI-BLAST2 uses several search parameters, wherein all of those search parameters are set to default values including, for example, unmask = yes, strand = all, expected occurrences = 10, minimum low complexity length = 15/5, multi-pass e-value = 0.01, constant for multi-pass = 25, dropoff for final gapped alignment = 25 and scoring matrix = BLOSUM62.

In situations where NCBI-BLAST2 is employed for amino acid sequence comparisons, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) is calculated as follows:

$$100 \text{ times the fraction } X/Y$$

where X is the number of amino acid residues scored as identical matches by the sequence alignment program NCBI-BLAST2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A.

"PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polynucleotide" or "PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant nucleic acid sequence" means a nucleic acid molecule which encodes an active PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide as defined below and which has at least about 80% nucleic acid sequence identity with the nucleotide acid sequence of the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding

nucleic acid sequence shown in Figure 1 (SEQ ID NO:1), Figures 3A-B (SEQ ID NO:6), Figure 5 (SEQ ID NO:9), Figure 7 (SEQ ID NO:13), Figure 9 (SEQ ID NO:15), Figure 11 (SEQ ID NO:17), Figure 13 (SEQ ID NO:22), Figure 15 (SEQ ID NO:24), Figure 17 (SEQ ID NO:26), Figures 19A-B (SEQ ID NO:28), Figure 21 (SEQ ID NO:33), Figures 23 (SEQ ID NO:35), Figure 25 (SEQ ID NO:37), Figure 27 (SEQ ID NO:39), Figure 29 (SEQ ID NO:41), Figure 31 (SEQ ID NO:43), Figure 33 (SEQ ID NO:45), Figure 35 (SEQ ID NO:48), Figure 37 (SEQ ID NO:50), Figure 39 (SEQ ID NO:52), Figure 41 (SEQ ID NO:54), Figures 43A-B (SEQ ID NO:56), Figure 45 (SEQ ID NO:64), Figure 47 (SEQ ID NO:66), Figure 49 (SEQ ID NO:68), Figures 51A-B (SEQ ID NO:70), Figure 53 (SEQ ID NO:72), Figures 55A-B (SEQ ID NO:74), Figure 57 (SEQ ID NO:76) and Figures 59A-B (SEQ ID NO:78) respectively, or a specifically derived fragment thereof. Ordinarily, a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polynucleotide will have at least about 80% nucleic acid sequence identity, more preferably at least about 81% nucleic acid sequence identity, more preferably at least about 82% nucleic acid sequence identity, more preferably at least about 83% nucleic acid sequence identity, more preferably at least about 84% nucleic acid sequence identity, more preferably at least about 85% nucleic acid sequence identity, more preferably at least about 86% nucleic acid sequence identity, more preferably at least about 87% nucleic acid sequence identity, more preferably at least about 88% nucleic acid sequence identity, more preferably at least about 89% nucleic acid sequence identity, more preferably at least about 90% nucleic acid sequence identity, more preferably at least about 91% nucleic acid sequence identity, more preferably at least about 92% nucleic acid sequence identity, more preferably at least about 93% nucleic acid sequence identity, more preferably at least about 94% nucleic acid sequence identity, more preferably at least about 95% nucleic acid sequence identity, more preferably at least about 96% nucleic acid sequence identity, more preferably at least about 97% nucleic acid sequence identity, more preferably at least about 98% nucleic acid sequence identity and yet more preferably at least about 99% nucleic acid sequence identity with the nucleic acid sequence of Figure 1 (SEQ ID NO:1), Figures 3A-B (SEQ ID NO:6), Figure 5 (SEQ ID NO:9), Figure 7 (SEQ ID NO:13), Figure 9 (SEQ ID NO:15), Figure 11 (SEQ ID NO:17), Figure 13 (SEQ ID NO:22), Figure 15 (SEQ ID NO:24), Figure 17 (SEQ ID NO:26), Figures 19A-B (SEQ ID NO:28), Figure 21 (SEQ ID NO:33), Figures 23 (SEQ ID NO:35), Figure 25 (SEQ ID NO:37), Figure 27 (SEQ ID NO:39), Figure 29 (SEQ ID NO:41), Figure 31 (SEQ ID NO:43), Figure 33 (SEQ ID NO:45), Figure 35 (SEQ ID NO:48), Figure 37 (SEQ ID NO:50), Figure 39 (SEQ ID NO:52), Figure 41 (SEQ ID NO:54), Figures 43A-B (SEQ ID NO:56), Figure 45 (SEQ ID NO:64), Figure 47 (SEQ ID NO:66), Figure 49 (SEQ ID NO:68), Figures 51A-B (SEQ ID NO:70), Figure 53 (SEQ ID NO:72), Figures 55A-B (SEQ ID NO:74), Figure 57 (SEQ ID NO:76) or Figures 59A-B (SEQ ID NO:78) respectively, or a derived fragment thereof. Variants do not encompass the native nucleotide sequence. In this regard, due to the degeneracy of the genetic code, one of ordinary skill in the art will immediately recognize that a large number of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polynucleotides having at least about 80% nucleic acid sequence identity to the nucleotide sequence of Figure 1 (SEQ ID NO:1), Figures 3A-B (SEQ ID NO:6), Figure 5

(SEQ ID NO:9), Figure 7 (SEQ ID NO:13), Figure 9 (SEQ ID NO:15), Figure 11 (SEQ ID NO:17), Figure 13 (SEQ ID NO:22), Figure 15 (SEQ ID NO:24), Figure 17 (SEQ ID NO:26), Figures 19A-B (SEQ ID NO:28), Figure 21 (SEQ ID NO:33), Figures 23 (SEQ ID NO:35), Figure 25 (SEQ ID NO:37), Figure 27 (SEQ ID NO:39), Figure 29 (SEQ ID NO:41), Figure 31 (SEQ ID NO:43), Figure 33 (SEQ ID NO:45), Figure 35 (SEQ ID NO:48), Figure 37 (SEQ ID NO:50), Figure 39 (SEQ ID NO:52), Figure 41 (SEQ ID NO:54), Figures 43A-B (SEQ ID NO:56), Figure 45 (SEQ ID NO:64), Figure 47 (SEQ ID NO:66), Figure 49 (SEQ ID NO:68), Figures 51A-B (SEQ ID NO:70), Figure 53 (SEQ ID NO:72), Figures 55A-B (SEQ ID NO:74), Figure 57 (SEQ ID NO:76) or Figures 59A-B (SEQ ID NO:78) respectively, will encode a polypeptide having an amino acid sequence which is identical to the amino acid sequence shown in Figure 2 (SEQ ID NO:2), Figure 4 (SEQ ID NO:7), Figure 6 (SEQ ID NO:10), Figure 8 (SEQ ID NO:14), Figure 10 (SEQ ID NO:16), Figure 12 (SEQ ID NO:18), Figure 14 (SEQ ID NO:23), Figure 16 (SEQ ID NO:25), Figure 18 (SEQ ID NO:27), Figure 20 (SEQ ID NO:29), Figure 22 (SEQ ID NO:34), Figure 24 (SEQ ID NO:36), Figure 26 (SEQ ID NO:38), Figure 28 (SEQ ID NO:40), Figure 30 (SEQ ID NO:42), Figure 32 (SEQ ID NO:44), Figure 34 (SEQ ID NO:46), Figure 36 (SEQ ID NO:49), Figure 38, (SEQ ID NO:51), Figure 40 (SEQ ID NO:53), Figure 42 (SEQ ID NO:55), Figure 44 (SEQ ID NO:57), Figure 46 (SEQ ID NO:65), Figure 48 (SEQ ID NO:67), Figure 50 (SEQ ID NO:69), Figure 52 (SEQ ID NO:71), Figure 54 (SEQ ID NO:73), Figure 56 (SEQ ID NO:75), Figure 58 (SEQ ID NO:77) or Figure 60 (SEQ ID NO:79) respectively or the amino acid sequence encoded by the DNA30942-1134, DNA35680-1212, DNA26288-1239, DNA49143-1429, DNA49821-1562, DNA50988-1326, DNA57836-1338, DNA56866-1342, DNA59839-1461, DNA52192-1369, DNA57708-1411, DNA57129-1413, DNA59622-1334, DNA59485-1336, DNA59841-1460, DNA59606-1471, DNA58721-1475, DNA59842-1502, DNA59848-1512, DNA59220-1514, DNA62876-1517 or DNA59820-1549 clone respectively, (the former DNA clones are shown in Figures 1, 3A-B, 5, 7, 9, 11, 13, 15, 17, 19A-B, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41 and 43A-B respectively, and have been deposited with the ATCC), or the amino acid sequence encoded by the known clones DNA36640, DNA36651, DNA56538, DNA71166, DNA82331, DNA83123, DNA88050 or DNA88153 respectively (the known DNA clones are shown in Figures 45, 47, 49, 51A-B, 53, 55A-B, 57, and 59A-B respectively).

Ordinarily, PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polynucleotides are at least about 30 nucleotides in length, often at least about 60 nucleotides in length, more often at least about 90 nucleotides in length, more often at least about 120 nucleotides in length, more often at least about 150 nucleotides in length, more often at least about 180 nucleotides in length, more often at least about 210 nucleotides in length, more often at least about 240 nucleotides in length, more often at least about 270 nucleotides in length, more often at least about 300 nucleotides in length, more often at least about 450 nucleotides in length, more often at least about 600 nucleotides in length, more often at least about 900 nucleotides in length, or more.

"Percent (%) nucleic acid sequence identity" with respect to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317,

PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide-encoding nucleic acid sequences identified herein is defined as the percentage of nucleotides in a candidate sequence that are identical with the nucleotides in a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide-encoding nucleic acid sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity. Alignment for purposes of determining percent nucleic acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN, ALIGN-2 or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full-length of the sequences being compared. For purposes herein, however, % nucleic acid sequence identity values are obtained as described below by using the sequence comparison computer program ALIGN-2, wherein the complete source code for the ALIGN-2 program is provided in Figures 62A-P. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc., and the source code shown in Figures 62A-P has been filed with user documentation in the U.S. Copyright Office, Washington D.C., 20559, where it is registered under U.S. Copyright Registration No. TXU510087. The ALIGN-2 program is publicly available through Genentech, Inc., South San Francisco, California or may be compiled from the source code provided in Figures 62A-P. The ALIGN-2 program should be compiled for use on a UNIX operating system, preferably digital UNIX V4.0D. All sequence comparison parameters are set by the ALIGN-2 program and do not vary.

For purposes herein, the % nucleic acid sequence identity of a given nucleic acid sequence C to, with, or against a given nucleic acid sequence D (which can alternatively be phrased as a given nucleic acid sequence C that has or comprises a certain % nucleic acid sequence identity to, with, or against a given nucleic acid sequence D) is calculated as follows:

$$100 \text{ times the fraction } W/Z$$

where W is the number of nucleotides scored as identical matches by the sequence alignment program ALIGN-2 in that program's alignment of C and D, and where Z is the total number of nucleotides in D. It will be appreciated that where the length of nucleic acid sequence C is not equal to the length of nucleic acid sequence D, the % nucleic acid sequence identity of C to D will not equal the % nucleic acid sequence identity of D to C. As examples of % nucleic acid sequence identity calculations, Figures 61C-D demonstrate how to calculate the % nucleic acid sequence identity of the nucleic acid sequence designated "Comparison DNA" to the nucleic acid sequence designated "PRO-DNA".

Unless specifically stated otherwise, all % nucleic acid sequence identity values used herein are obtained as described above using the ALIGN-2 sequence comparison computer program. However, % nucleic acid sequence identity may also be determined using the sequence comparison program NCBI-BLAST2 (Altschul *et al.*, Nucleic Acids Res., 25:3389-3402 (1997)). The NCBI-BLAST2 sequence comparison program may be downloaded from <http://www.ncbi.nlm.nih.gov>. NCBI-BLAST2 uses several search parameters, wherein all of those search parameters

are set to default values including, for example, unmask = yes, strand = all, expected occurrences = 10, minimum low complexity length = 15/5, multi-pass e-value = 0.01, constant for multi-pass = 25, dropoff for final gapped alignment = 25 and scoring matrix = BLOSUM62.

In situations where NCBI-BLAST2 is employed for sequence comparisons, the % nucleic acid sequence identity of a given nucleic acid sequence C to, with, or against a given nucleic acid sequence D (which can alternatively be phrased as a given nucleic acid sequence C that has or comprises a certain % nucleic acid sequence identity to, with, or against a given nucleic acid sequence D) is calculated as follows:

$$100 \text{ times the fraction } W/Z$$

10

where W is the number of nucleotides scored as identical matches by the sequence alignment program NCBI-BLAST2 in that program's alignment of C and D, and where Z is the total number of nucleotides in D. It will be appreciated that where the length of nucleic acid sequence C is not equal to the length of nucleic acid sequence D, the % nucleic acid sequence identity of C to D will not equal the % nucleic acid sequence identity of D to C.

15 In other embodiments, PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polynucleotides are nucleic acid molecules that encode an active PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide and which are capable of hybridizing, preferably under stringent hybridization and wash conditions, to nucleotide sequences encoding the full-length PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide shown in Figure 2 (SEQ ID NO:2), Figure 4 (SEQ ID NO:7), Figure 6 (SEQ ID NO:10), Figure 8 (SEQ ID NO:14), Figure 10 (SEQ ID NO:16), Figure 12 (SEQ ID NO:18), Figure 14 (SEQ ID NO:23), Figure 16 (SEQ ID NO:25), Figure 18 (SEQ ID NO:27), Figure 20 (SEQ ID NO:29), Figure 22 (SEQ ID NO:34), Figure 24 (SEQ ID NO:36), Figure 26 (SEQ ID NO:38), Figure 28 (SEQ ID NO:40), Figure 30 (SEQ ID NO:42), Figure 32 (SEQ ID NO:44), Figure 34 (SEQ ID NO:46), Figure 36 (SEQ ID NO:49), Figure 38 (SEQ ID NO:51), Figure 40 (SEQ ID NO:53), Figure 42 (SEQ ID NO:55), Figure 44 (SEQ ID NO:57), Figure 46 (SEQ ID NO:65), Figure 48 (SEQ ID NO:67), Figure 50 (SEQ ID NO:69), Figure 52 (SEQ ID NO:71), Figure 54 (SEQ ID NO:73), Figure 56 (SEQ ID NO:75), Figure 58 (SEQ ID NO:77) or Figure 60 (SEQ ID NO:79) respectively. PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polypeptides may be those that are encoded by a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39,

PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant polynucleotide.

The term "positives", in the context of the amino acid sequence identity comparisons performed as described above, includes amino acid residues in the sequences compared that are not only identical, but also those that have similar properties. Amino acid residues that score a positive value to an amino acid residue of interest are those that are either identical to the amino acid residue of interest or are a preferred substitution (as defined in Table 1 below) of the amino acid residue of interest.

For purposes herein, the % value of positives of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % positives to, with, or against a given amino acid sequence B) is calculated as follows:

10

$$100 \text{ times the fraction } X/Y$$

where X is the number of amino acid residues scoring a positive value as defined above by the sequence alignment program ALIGN-2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % positives of A to B will not equal the % positives of B to A.

"Isolated," when used to describe the various polypeptides disclosed herein, means polypeptide that has been identified and separated and/or recovered from a component of its natural environment. Preferably, the isolated polypeptide is free of association with all components with which it is naturally associated. Contaminant components of its natural environment are materials that would typically interfere with diagnostic or therapeutic uses for the polypeptide, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous solutes. In preferred embodiments, the polypeptide will be purified (1) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (2) to homogeneity by SDS-PAGE under non-reducing or reducing conditions using Coomassie blue or, preferably, silver stain. Isolated polypeptide includes polypeptide *in situ* within recombinant cells, since at least one component of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 natural environment will not be present. Ordinarily, however, isolated polypeptide will be prepared by at least one purification step.

An "isolated" nucleic acid molecule encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is a nucleic acid molecule that is identified and separated from at least one contaminant nucleic acid molecule with which it is ordinarily associated in the natural source of the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding nucleic acid. Preferably, the isolated nucleic acid is free of association with all components with which

it is naturally associated. An isolated PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding nucleic acid molecule is other than in the form or setting in which it is found in nature. Isolated nucleic acid molecules therefore are distinguished from the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding nucleic acid molecule as it exists in natural cells. However, an isolated nucleic acid molecule encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide includes PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-nucleic acid molecules contained in cells that ordinarily express PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 where, for example, the nucleic acid molecule is in a chromosomal location different from that of natural cells.

The term "control sequences" refers to DNA sequences necessary for the expression of an operably linked coding sequence in a particular host organism. The control sequences that are suitable for prokaryotes, for example, include a promoter, optionally an operator sequence, and a ribosome binding site. Eukaryotic cells are known to utilize promoters, polyadenylation signals, and enhancers.

Nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading phase. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

The term "antibody" is used in the broadest sense and specifically covers, for example, single anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 monoclonal antibodies

(including agonist, antagonist, and neutralizing antibodies), anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO845, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody compositions with polyepitopic specificity, single chain anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibodies, and fragments of anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibodies (see below). The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, *i.e.*, the individual antibodies comprising the population are identical except for possible naturally-occurring mutations that may be present in minor amounts.

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while shorter probes need lower temperatures. Hybridization generally depends on the ability of denatured DNA to reanneal when complementary strands are present in an environment below their melting temperature. The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature which can be used. As a result, it follows that higher relative temperatures would tend to make the reaction conditions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel *et al.*, Current Protocols in Molecular Biology, Wiley Interscience Publishers, (1995).

"Stringent conditions" or "high stringency conditions", as defined herein, may be identified by those that: (1) employ low ionic strength and high temperature for washing, for example 0.015 M sodium chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42°C; or (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium citrate) and 50% formamide at 55°C, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55°C.

"Moderately stringent conditions" may be identified as described by Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Press, 1989, and include the use of washing solution and hybridization conditions (*e.g.*, temperature, ionic strength and % SDS) less stringent than those described above.

An example of moderately stringent conditions is overnight incubation at 37°C in a solution comprising: 20% formamide, 5 x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5 x Denhardt's solution, 10% dextran sulfate, and 20 mg/mL denatured sheared salmon sperm DNA, followed by washing the filters in 1 x SSC at about 35-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc.

5 as necessary to accommodate factors such as probe length and the like.

The term "epitope tagged" when used herein refers to a chimeric polypeptide comprising a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide fused to a "tag polypeptide". The tag polypeptide has enough residues to provide an epitope against which an antibody can be made, yet is short enough such that it does not interfere with activity of the polypeptide to which it is fused. The tag polypeptide preferably also is fairly unique so that the antibody does not substantially cross-react with other epitopes. Suitable tag polypeptides generally have at least six amino acid residues and usually between about 8 and 50 amino acid residues (preferably, between about 10 and 20 amino acid residues).

15 "Active" or "activity" for the purposes herein refers to form(s) of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptides which retain a biological and/or an immunological activity/property of a native or naturally-occurring PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, wherein "biological" activity refers to a function (either inhibitory or stimulatory) caused by a native or naturally-occurring PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide other than the ability to induce the production of an antibody against an antigenic epitope possessed by a native or naturally-occurring PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide and an "immunological" activity refers to the ability to induce the production of an antibody against an antigenic epitope possessed by a native or naturally- occurring PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

35 "Biological activity" in the context of an antibody or another molecule that can be identified by the screening assays disclosed herein (*e.g.*, an organic or inorganic small molecule, peptide, etc.) is used to refer to the ability of such molecules to bind or complex with the polypeptides encoded by the amplified genes identified herein, or otherwise interfere with the interaction of the encoded polypeptides with other cellular proteins. A preferred

biological activity is growth inhibition of a target tumor cell. Another preferred biological activity is cytotoxic activity resulting in the death of the target tumor cell.

The phrase "immunological property" means immunological cross-reactivity with at least one epitope of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

"Immunological cross-reactivity" as used herein means that the candidate polypeptide is capable of competitively inhibiting the qualitative biological activity of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide having this activity with polyclonal antisera raised against the known active PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Such antisera are prepared in conventional fashion by injecting goats or rabbits, for example, subcutaneously with the known active analogue in complete Freund's adjuvant, followed by booster intraperitoneal or subcutaneous injection in incomplete Freund's. The immunological cross-reactivity preferably is "specific", which means that the binding affinity of the immunologically cross-reactive molecule (*e.g.*, antibody) identified, to the corresponding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is significantly higher (preferably at least about 2-times, more preferably at least about 4-times, even more preferably at least about 8-times, most preferably at least about 10-times higher) than the binding affinity of that molecule to any other known native polypeptide.

The term "antagonist" is used in the broadest sense, and includes any molecule that partially or fully blocks, inhibits, or neutralizes a biological activity of a native PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide disclosed herein. In a similar manner, the term "agonist" is used in the broadest sense and includes any molecule that mimics a biological activity of a native PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide disclosed herein. Suitable agonist or antagonist molecules specifically include agonist or antagonist antibodies or antibody fragments, fragments or amino acid sequence variants of native PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptides, peptides, small organic molecules, etc. Methods for identifying agonists or antagonists of a PRO212,

PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide with a candidate agonist or antagonist molecule and measuring a detectable change in one or more biological activities normally associated with the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

A "small molecule" is defined herein to have a molecular weight below about 500 Daltons.

"Antibodies" (Abs) and "immunoglobulins" (Igs) are glycoproteins having the same structural characteristics. While antibodies exhibit binding specificity to a specific antigen, immunoglobulins include both antibodies and other antibody-like molecules which lack antigen specificity. Polypeptides of the latter kind are, for example, produced at low levels by the lymph system and at increased levels by myelomas. The term "antibody" is used in the broadest sense and specifically covers, without limitation, intact monoclonal antibodies, polyclonal antibodies, multispecific antibodies (*e.g.*, bispecific antibodies) formed from at least two intact antibodies, and antibody fragments so long as they exhibit the desired biological activity.

"Native antibodies" and "native immunoglobulins" are usually heterotetrameric glycoproteins of about 150,000 Dalton, composed of two identical light (L) chains and two identical heavy (H) chains. Each light chain is linked to a heavy chain by one covalent disulfide bond, while the number of disulfide linkages varies among the heavy chains of different immunoglobulin isotypes. Each heavy and light chain also has regularly spaced intrachain disulfide bridges. Each heavy chain has at one end a variable domain (V_H) followed by a number of constant domains. Each light chain has a variable domain at one end (V_L) and a constant domain at its other end; the constant domain of the light chain is aligned with the first constant domain of the heavy chain, and the light-chain variable domain is aligned with the variable domain of the heavy chain. Particular amino acid residues are believed to form an interface between the light- and heavy-chain variable domains.

The term "variable" refers to the fact that certain portions of the variable domains differ extensively in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen. However, the variability is not evenly distributed throughout the variable domains of antibodies. It is concentrated in three segments called complementarity-determining regions (CDRs) or hypervariable regions both in the light-chain and the heavy-chain variable domains. The more highly conserved portions of variable domains are called the framework (FR). The variable domains of native heavy and light chains each comprise four FR regions, largely adopting a β -sheet configuration, connected by three CDRs, which form loops connecting, and in some cases forming part of, the β -sheet structure. The CDRs in each chain are held together in close proximity by the FR regions and, with the CDRs from the other chain, contribute to the formation of the antigen-binding site of antibodies (see Kabat *et al.*, NIH Publ. No.91-3242, Vol. I, pages 647-669 (1991)). The constant domains are not involved directly in binding an antibody to an antigen, but exhibit various effector functions, such as participation of the antibody in antibody-dependent cellular toxicity.

The term "hypervariable region" when used herein refers to the amino acid residues of an antibody which

are responsible for antigen-binding. The hypervariable region comprises amino acid residues from a "complementarity determining region" to "CDR" (*i.e.*, residues 24-34 (L1), 50-56 (L2) and 89-97 (L3) in the light chain variable domain and 31-35 (H1), 50-65 (H2) and 95-102 (H3) in the heavy chain variable domain; Kabat *et al.*, Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institute of Health, Bethesda, MD. [1991]) and/or those residues from a "hypervariable loop" (*i.e.*, residues 26-32 (L1), 50-52 (L2) and 91-96 (L3) in the light chain variable domain and 26-32 (H1), 53-55 (H2) and 96-101 (H3) in the heavy chain variable domain; Clothia and Lesk, J. Mol. Biol., 196:901-917 [1987]). "Framework" or "FR" residues are those variable domain residues other than the hypervariable region residues as herein defined.

"Antibody fragments" comprise a portion of an intact antibody, preferably the antigen binding or variable region of the intact antibody. Examples of antibody fragments include Fab, Fab', F(ab')₂, and Fv fragments; diabodies; linear antibodies (Zapata *et al.*, Protein Eng., 8(10):1057-1062 [1995]); single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site, and a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an F(ab')₂ fragment that has two antigen-combining sites and is still capable of cross-linking antigen.

"Fv" is the minimum antibody fragment which contains a complete antigen-recognition and -binding site. This region consists of a dimer of one heavy- and one light-chain variable domain in tight, non-covalent association. It is in this configuration that the three CDRs of each variable domain interact to define an antigen-binding site on the surface of the V_H-V_L dimer. Collectively, the six CDRs confer antigen-binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

The Fab fragment also contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab fragments differ from Fab' fragments by the addition of a few residues at the carboxy terminus of the heavy chain CH1 domain including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. F(ab')₂ antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

The "light chains" of antibodies (immunoglobulins) from any vertebrate species can be assigned to one of two clearly distinct types, called kappa (κ) and lambda (λ), based on the amino acid sequences of their constant domains.

Depending on the amino acid sequence of the constant domain of their heavy chains, immunoglobulins can be assigned to different classes. There are five major classes of immunoglobulins: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into subclasses (isotypes), *e.g.*, IgG1, IgG2, IgG3, IgG4, IgA, and IgA2. The heavy-chain constant domains that correspond to the different classes of immunoglobulins are called α, δ, ε, γ, and μ, respectively. The subunit structures and three-dimensional configurations of different classes of immunoglobulins are well known.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of

substantially homogeneous antibodies, *i.e.*, the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations which typically include different antibodies directed against different determinants (epitopes), each
5 monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they are synthesized by the hybridoma culture, uncontaminated by other immunoglobulins. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the
10 present invention may be made by the hybridoma method first described by Kohler *et al.*, Nature, 256:495 [1975], or may be made by recombinant DNA methods (see, *e.g.*, U.S. Patent No. 4,816,567). The "monoclonal antibodies" may also be isolated from phage antibody libraries using the techniques described in Clackson *et al.*, Nature, 352:624-628 [1991] and Marks *et al.*, J. Mol. Biol., 222:581-597 (1991), for example.

The monoclonal antibodies herein specifically include "chimeric" antibodies (immunoglobulins) in which
15 a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (U.S. Patent No. 4,816,567; Morrison *et al.*, Proc. Natl. Acad. Sci. USA, 81:6851-6855
20 [1984]).

"Humanized" forms of non-human (*e.g.*, murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', F(ab')₂ or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which residues from a CDR of the
25 recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity, and capacity. In some instances, Fv FR residues of the human immunoglobulin are replaced by corresponding non-human residues. Furthermore, humanized antibodies may comprise residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. These modifications are made to further refine and maximize antibody performance. In general, the
30 humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. For further details, see Jones *et al.*, Nature, 321:522-525 (1986); Reichmann *et al.*, Nature, 332:323-329 [1988]; and
35 Presta, Curr. Op. Struct. Biol., 2:593-596 (1992). The humanized antibody includes a PRIMATIZED™ antibody wherein the antigen-binding region of the antibody is derived from an antibody produced by immunizing macaque monkeys with the antigen of interest.

"Single-chain Fv" or "sFv" antibody fragments comprise the V_H and V_L domains of antibody, wherein these

domains are present in a single polypeptide chain. Preferably, the Fv polypeptide further comprises a polypeptide linker between the V_H and V_L domains which enables the sFv to form the desired structure for antigen binding. For a review of sFv see Pluckthun in The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenberg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994).

5 The term "diabodies" refers to small antibody fragments with two antigen-binding sites, which fragments comprise a heavy-chain variable domain (V_H) connected to a light-chain variable domain (V_L) in the same polypeptide chain (V_H - V_L). By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites. Diabodies are described more fully in, for example, EP 404,097; WO 93/11161; and Hollinger *et al.*,
10 Proc. Natl. Acad. Sci. USA, 90:6444-6448 (1993).

An "isolated" antibody is one which has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or nonproteinaceous solutes. In preferred embodiments, the antibody will be purified (1) to greater than 95% by
15 weight of antibody as determined by the Lowry method, and most preferably more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (3) to homogeneity by SDS-PAGE under reducing or nonreducing conditions using Coomassie blue or, preferably, silver stain. Isolated antibody includes the antibody *in situ* within recombinant cells since at least one component of the antibody's natural environment will not be present. Ordinarily, however, isolated antibody
20 will be prepared by at least one purification step.

The word "label" when used herein refers to a detectable compound or composition which is conjugated directly or indirectly to the antibody so as to generate a "labeled" antibody. The label may be detectable by itself (*e.g.*, radioisotope labels or fluorescent labels) or, in the case of an enzymatic label, may catalyze chemical alteration of a substrate compound or composition which is detectable. Radionuclides that can serve as detectable labels
25 include, for example, I-131, I-123, I-125, Y-90, Re-188, Re-186, At-211, Cu-67, Bi-212, and Pd-109.

By "solid phase" is meant a non-aqueous matrix to which the antibody of the present invention can adhere. Examples of solid phases encompassed herein include those formed partially or entirely of glass (*e.g.*, controlled pore glass), polysaccharides (*e.g.*, agarose), polyacrylamides, polystyrene, polyvinyl alcohol and silicones. In certain embodiments, depending on the context, the solid phase can comprise the well of an assay plate; in others it is a
30 purification column (*e.g.*, an affinity chromatography column). This term also includes a discontinuous solid phase of discrete particles, such as those described in U.S. Patent No. 4,275,149.

A "liposome" is a small vesicle composed of various types of lipids, phospholipids and/or surfactant which is useful for delivery of a drug (such as a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111,
35 PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide or antibody thereto and, optionally, a chemotherapeutic agent) to a mammal. The components of the liposome are commonly arranged in a bilayer formation, similar to the lipid arrangement of biological membranes.

As used herein, the term "immunoadhesin" designates antibody-like molecules which combine the binding specificity of a heterologous protein (an "adhesin") with the effector functions of immunoglobulin constant domains. Structurally, the immunoadhesins comprise a fusion of an amino acid sequence with the desired binding specificity which is other than the antigen recognition and binding site of an antibody (*i.e.*, is "heterologous"), and an immunoglobulin constant domain sequence. The adhesin part of an immunoadhesin molecule typically is a contiguous amino acid sequence comprising at least the binding site of a receptor or a ligand. The immunoglobulin constant domain sequence in the immunoadhesin may be obtained from any immunoglobulin, such as IgG-1, IgG-2, IgG-3, or IgG-4 subtypes, IgA (including IgA-1 and IgA-2), IgE, IgD or IgM.

II. Compositions and Methods of the Invention

I. Full-length PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198. In particular, cDNA encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198 polypeptides has been identified and isolated, as disclosed in further detail in the Examples below. It is noted that proteins produced in separate expression rounds may be given different PRO numbers but the UNQ number is unique for any given DNA and the encoded protein, and will not be changed. However, for sake of simplicity, in the present specification the proteins encoded by the herein disclosed nucleic acid sequences as well as all further native homologues and variants included in the foregoing definition of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198 will be referred to as "PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198", regardless of their origin or mode of preparation.

As disclosed in the Examples below, cDNA clones have been deposited with the ATCC with the exception of known clones DNA36640, DNA36651, DNA56538, DNA71166, DNA82331, DNA83123, DNA88050 and DNA88153, respectively. The actual nucleotide sequence of the clones can readily be determined by the skilled artisan by sequencing of the deposited clone using routine methods in the art. The predicted amino acid sequences can be determined from the nucleotide sequences using routine skill. For the PRO212, PRO290, PRO341, PRO535,

PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptides and encoding nucleic acid described herein, Applicants have identified what are believed to be the reading frames best identifiable with the sequence information available at the time.

B. PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198 Variants

In addition to the full-length native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptides described herein, it is contemplated that PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198 variants can be prepared. PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198 variants can be prepared by introducing appropriate nucleotide changes into the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 DNA, and/or by synthesis of the desired PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Those skilled in the art will appreciate that amino acid changes may alter post-translational processes of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198, such as changing the number or position of glycosylation sites or altering the membrane anchoring characteristics.

Variations in the native full-length sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 or in various domains of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 described herein, can be made, for example, using any of the techniques and guidelines for

conservative and non-conservative mutations set forth, for instance, in U.S. Patent No. 5,364,934. Variations may be a substitution, deletion or insertion of one or more codons encoding the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 that results in a change in the amino acid sequence of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 as compared with the native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. Optionally the variation is by substitution of at least one amino acid with any other amino acid in one or more of the domains of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. Guidance in determining which amino acid residue may be inserted, substituted or deleted without adversely affecting the desired activity may be found by comparing the sequence of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 with that of homologous known protein molecules and minimizing the number of amino acid sequence changes made in regions of high homology. Amino acid substitutions can be the result of replacing one amino acid with another amino acid having similar structural and/or chemical properties, such as the replacement of a leucine with a serine, *i.e.*, conservative amino acid replacements. Insertions or deletions may optionally be in the range of about 1 to 5 amino acids. The variation allowed may be determined by systematically making insertions, deletions or substitutions of amino acids in the sequence and testing the resulting variants for activity exhibited by the full-length or mature native sequence.

PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145, and PRO2198 polypeptide fragments are provided herein. Such fragments may be truncated at the N-terminus or C-terminus, or may lack internal residues, for example, when compared with a full length native protein. Certain fragments lack amino acid residues that are not essential for a desired biological activity of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 fragments

may be prepared by any of a number of conventional techniques. Desired peptide fragments may be chemically synthesized. An alternative approach involves generating PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 fragments by enzymatic digestion, *e.g.*, by treating the protein with an enzyme known to cleave proteins at sites defined by particular amino acid residues, or by digesting the DNA with suitable restriction enzymes and isolating the desired fragment. Yet another suitable technique involves isolating and amplifying a DNA fragment encoding a desired polypeptide fragment, by polymerase chain reaction (PCR). Oligonucleotides that define the desired termini of the DNA fragment are employed at the 5' and 3' primers in the PCR. Preferably, PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide fragments share at least one biological and/or immunological activity with the native PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

In particular embodiments, conservative substitutions of interest are shown in Table 1 under the heading of preferred substitutions. If such substitutions result in a change in biological activity, then more substantial changes, denominated exemplary substitutions in Table 1, or as further described below in reference to amino acid classes, are introduced and the products screened.

Table 1

	<u>Original Residue</u>	<u>Exemplary Substitutions</u>	<u>Preferred Substitutions</u>
5	Ala (A)	val; leu; ile	val
	Arg (R)	lys; gln; asn	lys
	Asn (N)	gln; his; lys; arg	gln
	Asp (D)	glu	glu
10	Cys (C)	ser	ser
	Gln (Q)	asn	asn
	Glu (E)	asp	asp
	Gly (G)	pro; ala	ala
	His (H)	asn; gln; lys; arg	arg
15	Ile (I)	leu; val; met; ala; phe; norleucine	leu
	Leu (L)	norleucine; ile; val; met; ala; phe	ile
	Lys (K)	arg; gln; asn	arg
20	Met (M)	leu; phe; ile	leu
	Phe (F)	leu; val; ile; ala; tyr	leu
	Pro (P)	ala	ala
	Ser (S)	thr	thr
	Thr (T)	ser	ser
25	Trp (W)	tyr; phe	tyr
	Tyr (Y)	trp; phe; thr; ser	phe
	Val (V)	ile; leu; met; phe; ala; norleucine	leu

30 Substantial modifications in function or immunological identity of the polypeptide are accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues are divided into groups based on common side-chain properties:

- 35 (1) hydrophobic: norleucine, met, ala, val, leu, ile;
 (2) neutral hydrophilic: cys, ser, thr;
 (3) acidic: asp, glu;
 (4) basic: asn, gln, his, lys, arg;
 (5) residues that influence chain orientation: gly, pro; and
 40 (6) aromatic: trp, tyr, phe.

Non-conservative substitutions will entail exchanging a member of one of these classes for another class. Such substituted residues also may be introduced into the conservative substitution sites or, more preferably, into the remaining (non-conserved) sites.

The variations can be made using methods known in the art such as oligonucleotide-mediated (site-directed)
 45 mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis [Carter *et al.*, Nucl. Acids Res., 13:4331 (1986); Zoller *et al.*, Nucl. Acids Res., 10:6487 (1987)], cassette mutagenesis [Wells *et al.*, Gene, 34:315 (1985)], restriction selection mutagenesis [Wells *et al.*, Philos. Trans. R. Soc. London SerA, 317:415 (1986)] or

other known techniques can be performed on the cloned DNA to produce the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 variant DNA.

5 Scanning amino acid analysis can also be employed to identify one or more amino acids along a contiguous sequence. Among the preferred scanning amino acids are relatively small, neutral amino acids. Such amino acids include alanine, glycine, serine, and cysteine. Alanine is typically a preferred scanning amino acid among this group because it eliminates the side-chain beyond the beta-carbon and is less likely to alter the main-chain conformation of the variant [Cunningham and Wells, Science, 244: 1081-1085 (1989)]. Alanine is also typically preferred because
10 it is the most common amino acid. Further, it is frequently found in both buried and exposed positions [Creighton, The Proteins, (W.H. Freeman & Co., N.Y.); Chothia, J. Mol. Biol., 150:1 (1976)]. If alanine substitution does not yield adequate amounts of variant, an isoteric amino acid can be used.

C. Modifications of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198

Covalent modifications of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 and PRO2198 are included within the scope of this invention. One type of covalent modification includes reacting targeted amino acid residues of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C- terminal residues of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. Derivatization with bifunctional agents is useful, for instance, for crosslinking PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 to a water-insoluble support matrix or surface for use in the method for purifying anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibodies, and vice-versa. Commonly used crosslinking agents include, e.g., 1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde, N-hydroxysuccinimide esters, for example, esters with 4-

azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis(succinimidylpropionate), bifunctional maleimides such as bis-N-maleimido-1,8-octane and agents such as methyl-3-[(p-azidophenyl)dithio]propioimide.

Other modifications include deamidation of glutamyl and asparaginyl residues to the corresponding
5 glutamyl and aspartyl residues, respectively, hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α -amino groups of lysine, arginine, and histidine side chains [T.E. Creighton, Proteins: Structure and Molecular Properties, W.H. Freeman & Co., San Francisco, pp. 79-86 (1983)], acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

Another type of covalent modification of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717,
10 PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide included within the scope of this invention comprises altering the native glycosylation pattern of the polypeptide. "Altering the native glycosylation pattern" is intended for purposes herein to mean deleting one or more carbohydrate moieties found in native sequence PRO212, PRO290, PRO341, PRO535,
15 PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 (either by removing the underlying glycosylation site or by deleting the glycosylation by chemical and/or enzymatic means), and/or adding one or more glycosylation sites that are not present in the native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830,
20 PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. In addition, the phrase includes qualitative changes in the glycosylation of the native proteins, involving a change in the nature and proportions of the various carbohydrate moieties present.

Addition of glycosylation sites to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809,
25 PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide may be accomplished by altering the amino acid sequence. The alteration may be made, for example, by the addition of, or substitution by, one or more serine or threonine residues to the native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943,
30 PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 (for O-linked glycosylation sites). The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145
35 or PRO2198 amino acid sequence may optionally be altered through changes at the DNA level, particularly by mutating the DNA encoding the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145

or PRO2198 polypeptide at preselected bases such that codons are generated that will translate into the desired amino acids.

Another means of increasing the number of carbohydrate moieties on the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is by chemical or enzymatic coupling of glycosides to the polypeptide. Such methods are described in the art, *e.g.*, in WO 87/05330 published 11 September 1987, and in Aplin and Wriston, CRC Crit. Rev. Biochem., pp. 259-306 (1981).

Removal of carbohydrate moieties present on the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide may be accomplished chemically or enzymatically or by mutational substitution of codons encoding for amino acid residues that serve as targets for glycosylation. Chemical deglycosylation techniques are known in the art and described, for instance, by Hakimuddin, *et al.*, Arch. Biochem. Biophys., 259:52 (1987) and by Edge *et al.*, Anal. Biochem., 118:131 (1981). Enzymatic cleavage of carbohydrate moieties on polypeptides can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura *et al.*, Meth. Enzymol., 138:350 (1987).

Another type of covalent modification of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 comprises linking the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide to one of a variety of nonproteinaceous polymers, *e.g.*, polyethylene glycol (PEG), polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.

The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 of the present invention may also be modified in a way to form a chimeric molecule comprising PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 fused to another, heterologous polypeptide or amino acid sequence.

In one embodiment, such a chimeric molecule comprises a fusion of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 with a tag polypeptide which provides an epitope to which

an anti-tag antibody can selectively bind. The epitope tag is generally placed at the amino- or carboxyl-terminus of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. The presence of

5 such epitope-tagged forms of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can be detected using an antibody against the tag polypeptide. Also, provision of the epitope tag enables the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943,

10 PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 to be readily purified by affinity purification using an anti-tag antibody or another type of affinity matrix that binds to the epitope tag. Various tag polypeptides and their respective antibodies are well known in the art. Examples include poly-histidine (poly-his) or poly-histidine-glycine (poly-his-gly) tags; the flu HA tag polypeptide and its antibody

15 12CA5 [Field *et al.*, Mol. Cell. Biol., 8:2159-2165 (1988)]; the c-myc tag and the 8F9, 3C7, 6E10, G4, B7 and 9E10 antibodies thereto [Evan *et al.*, Molecular and Cellular Biology, 5:3610-3616 (1985)]; and the Herpes Simplex virus glycoprotein D (gD) tag and its antibody [Paborsky *et al.*, Protein Engineering, 3(6):547-553 (1990)]. Other tag polypeptides include the Flag-peptide [Hopp *et al.*, BioTechnology, 6:1204-1210 (1988)]; the KT3 epitope peptide [Martin *et al.*, Science, 255:192-194 (1992)]; an α -tubulin epitope peptide [Skinner *et al.*, J. Biol. Chem.,

20 266:15163-15166 (1991)]; and the T7 gene 10 protein peptide tag [Lutz-Freyermuth *et al.*, Proc. Natl. Acad. Sci. USA, 87:6393-6397 (1990)].

In an alternative embodiment, the chimeric molecule may comprise a fusion of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39,

25 PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the chimeric molecule (also referred to as an "immunoadhesin"), such a fusion could be to the Fc region of an IgG molecule. The Ig fusions preferably include the substitution of a soluble (transmembrane domain deleted or inactivated) form of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111,

30 PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in place of at least one variable region within an Ig molecule. In a particularly preferred embodiment, the immunoglobulin fusion includes the hinge, CH2 and CH3, or the hinge, CH1, CH2 and CH3 regions of an IgG1 molecule. For the production of immunoglobulin fusions see also US Patent No. 5,428,130 issued June 27, 1995.

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D. Preparation of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198

5 The description below relates primarily to production of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 by culturing cells transformed or transfected with a vector containing PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 nucleic acid. It is, of course, contemplated that alternative methods, which are well known in the art, may be employed to prepare PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. For instance, the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 sequence, or portions thereof, may be produced by direct peptide synthesis using solid-phase techniques [see, e.g., Stewart *et al.*, Solid-Phase Peptide Synthesis, W.H. Freeman Co., San Francisco, CA (1969); Merrifield, J. Am. Chem. Soc., 85:2149-2154 (1963)]. *In vitro* protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be accomplished, for instance, using an Applied Biosystems Peptide Synthesizer (Foster City, CA) using manufacturer's instructions. Various portions of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 may be chemically synthesized separately and combined using chemical or enzymatic methods to produce the full-length PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198.

a. Isolation of DNA Encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 Polypeptide

DNA encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or

PRO2198 may be obtained from a cDNA library prepared from tissue believed to possess the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 mRNA and to express it at a detectable level.

5 Accordingly, human PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 DNA can be conveniently obtained from a cDNA library prepared from human tissue, such as described in the Examples. PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943,
10 PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 encoding gene may also be obtained from a genomic library or by oligonucleotide synthesis.

Libraries can be screened with probes (such as antibodies to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097,
15 PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, or oligonucleotides of at least about 20-80 bases) designed to identify the gene of interest or the protein encoded by it. Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook *et al.*, Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to
20 isolate the gene encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 is to use PCR methodology [Sambrook *et al.*, *supra*; Dieffenbach *et al.*, PCR Primer: A Laboratory Manual (Cold Spring Harbor Laboratory Press, 1995)].

25 The Examples below describe techniques for screening a cDNA library. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized. The oligonucleotide is preferably labeled such that it can be detected upon hybridization to DNA in the library being screened. Methods of labeling are well known in the art, and include the use of radiolabels like ³²P-labeled ATP, biotinylation or enzyme labeling. Hybridization conditions, including moderate stringency and high stringency, are
30 provided in Sambrook *et al.*, *supra*.

Sequences identified in such library screening methods can be compared and aligned to other known sequences deposited and available in public databases such as GenBank or other private sequence databases. Sequence identity (at either the amino acid or nucleotide level) within defined regions of the molecule or across the full-length sequence can be determined using methods known in the art and as described herein..

35 Nucleic acid having protein coding sequence may be obtained by screening selected cDNA or genomic libraries using the deduced amino acid sequence disclosed herein for the first time, and, if necessary, using conventional primer extension procedures as described in Sambrook *et al.*, *supra*, to detect precursors and processing intermediates of mRNA that may not have been reverse-transcribed into cDNA.

b. Selection and Transformation of Host Cells

Host cells are transfected or transformed with expression or cloning vectors described herein for PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281,
5 PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in Mammalian Cell Biotechnology: a
10 Practical Approach, M. Butler, ed. (IRL Press, 1991) and Sambrook *et al.*, *supra*.

Methods of eukaryotic cell transfection and prokaryotic cell transformation are known to the ordinarily skilled artisan, for example, CaCl_2 , CaPO_4 , liposome-mediated and electroporation. Depending on the host cell used, transformation is performed using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described in Sambrook *et al.*, *supra*, or electroporation is generally used for prokaryotes.
15 Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw *et al.*, Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) can be employed. General aspects of mammalian cell host system transfections have been described in U.S. Patent No. 4,399,216. Transformations into yeast are typically carried out according to the method of Van Solingen *et al.*, J.
20 Bact., 130:946 (1977) and Hsiao *et al.*, Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, *e.g.*, polybrene, polyornithine, may also be used. For various techniques for transforming mammalian cells, see Keown *et al.*, Methods in Enzymology, 185:527-537 (1990) and Mansour *et al.*, Nature, 336:348-352 (1988).

25 Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as *E. coli*. Various *E. coli* strains are publicly available, such as *E. coli* K12 strain MM294 (ATCC 31,446); *E. coli* X1776 (ATCC 31,537); *E. coli* strain W3110 (ATCC 27,325) and *E. Coli* strain K5 772 (ATCC 53,635). Other suitable prokaryotic host cells include
30 Enterobacteriaceae such as *Escherichia*, *e.g.*, *E. coli*, *Enterobacter*, *Erwinia*, *Klebsiella*, *Proteus*, *Salmonella*, *e.g.*, *Salmonella typhimurium*, *Serratia*, *e.g.*, *Serratia marcescans*, and *Shigella*, as well as *Bacilli* such as *B. subtilis* and *B. licheniformis* (*e.g.*, *B. licheniformis* 41P disclosed in DD 266,710 published 12 April 1989), *Pseudomonas* such as *P. aeruginosa*, and *Streptomyces*. These examples are illustrative rather than limiting. Strain W3110 is one particularly preferred host or parent host because it is a common host strain for recombinant DNA product
35 fermentations. Preferably, the host cell secretes minimal amounts of proteolytic enzymes. For example, strain W3110 may be modified to effect a genetic mutation in the genes encoding proteins endogenous to the host, with examples of such hosts including *E. coli* W3110 strain 1A2, which has the complete genotype *tonA*; *E. coli* W3110 strain 9E4, which has the complete genotype *tonA ptr3*; *E. coli* W3110 strain 27C7 (ATCC 55,244), which has the

complete genotype *tonA ptr3 phoA E15 (argF-lac)169 degP ompT kan^r*; *E. coli* W3110 strain 37D6, which has the complete genotype *tonA ptr3 phoA E15 (argF-lac)169 degP ompT rbs7 ilvG kan^r*; *E. coli* W3110 strain 40B4, which is strain 37D6 with a non-kanamycin resistant *degP* deletion mutation; and an *E. coli* strain having mutant periplasmic protease disclosed in U.S. Patent No. 4,946,783 issued 7 August 1990. Alternatively, *in vitro* methods of cloning, *e.g.*, PCR or other nucleic acid polymerase reactions, are suitable.

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding vectors. *Saccharomyces cerevisiae* is a commonly used lower eukaryotic host microorganism. Others include *Schizosaccharomyces pombe* (Beach and Nurse, Nature, 290: 140 [1981]; EP 139,383 published 2 May 1985); *Kluyveromyces* hosts (U.S. Patent No. 4,943,529; Fleer *et al.*, Bio/Technology, 9: 968-975 (1991)) such as, *e.g.*, *K. lactis* (MW98-8C, CBS683, CBS4574; Louvencourt *et al.*, J. Bacteriol., 737 [1983]), *K. fragilis* (ATCC 12,424), *K. bulgaricus* (ATCC 16,045), *K. wickerhamii* (ATCC 24,178), *K. waltii* (ATCC 56,500), *K. drosophilae* (ATCC 36,906; Vanden Berg *et al.*, Bio/Technology, 8:135 (1990)), *K. thermotolerans*, and *K. marxianus*; *Yarrowia* (EP 402,226); *Pichia pastoris* (EP 183,070; Sreekrishna *et al.*, J. Basic Microbiol., 28:265-278 [1988]); *Candida*; *Trichoderma reesei* (EP 244,234); *Neurospora crassa* (Case *et al.*, Proc. Natl. Acad. Sci. USA, 76:5259-5263 [1979]); *Schwanniomyces* such as *Schwanniomyces occidentalis* (EP 394,538 published 31 October 1990); and filamentous fungi such as, *e.g.*, *Neurospora*, *Penicillium*, *Tolypocladium* (WO 91/00357 published 10 January 1991), and *Aspergillus* hosts such as *A. nidulans* (Ballance *et al.*, Biochem. Biophys. Res. Commun., 112:284-289 [1983]; Tilburn *et al.*, Gene, 26:205-221 [1983]; Yelton *et al.*, Proc. Natl. Acad. Sci. USA, 81:1470-1474 [1984]) and *A. niger* (Kelly and Hynes, EMBO J., 4:475-479 [1985]). Methylotrophic yeasts are suitable herein and include, but are not limited to, yeast capable of growth on methanol selected from the genera consisting of *Hansenula*, *Candida*, *Kloeckera*, *Pichia*, *Saccharomyces*, *Torulopsis*, and *Rhodotorula*. A list of specific species that are exemplary of this class of yeasts may be found in C. Anthony, The Biochemistry of Methylotrophs, 269 (1982).

Suitable host cells for the expression of glycosylated PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 are derived from multicellular organisms. Examples of invertebrate cells include insect cells such as *Drosophila* S2 and *Spodoptera* Sf9, as well as plant cells. Examples of useful mammalian host cell lines include Chinese hamster ovary (CHO) and COS cells. More specific examples include monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham *et al.*, J. Gen. Virol., 36:59 (1977)); Chinese hamster ovary cells/DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 (1980)); mouse sertoli cells (TM4, Mather, Biol. Reprod., 23:243-251 (1980)); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); and mouse mammary tumor (MMT 060562, ATCC CCL51). The selection of the appropriate host cell is deemed to be within the skill in the art.

c. Selection and Use of a Replicable Vector

The nucleic acid (*e.g.*, cDNA or genomic DNA) encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 may be inserted into a replicable vector for cloning (amplification of the DNA) or for expression. Various vectors are publicly available. The vector may, for example, be in the form of a plasmid, cosmid, viral particle, or phage. The appropriate nucleic acid sequence may be inserted into the vector by a variety of procedures. In general, DNA is inserted into an appropriate restriction endonuclease site(s) using techniques known in the art. Vector components generally include, but are not limited to, one or more of a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence. Construction of suitable vectors containing one or more of these components employs standard ligation techniques which are known to the skilled artisan.

The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 may be produced recombinantly not only directly, but also as a fusion polypeptide with a heterologous polypeptide, which may be a signal sequence or other polypeptide having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the vector, or it may be a part of the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding DNA that is inserted into the vector. The signal sequence may be a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, lpp, or heat-stable enterotoxin II leaders. For yeast secretion the signal sequence may be, *e.g.*, the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* α -factor leaders, the latter described in U.S. Patent No. 5,010,182), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression, mammalian signal sequences may be used to direct secretion of the protein, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders.

Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2 μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells.

Expression and cloning vectors will typically contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, *e.g.*, ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, *e.g.*, the gene encoding D-alanine racemase for *Bacilli*.

An example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding nucleic acid, such as DHFR or thymidine kinase. An appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and propagated as described by Urlaub *et al.*, Proc. Natl. Acad. Sci. USA, 77:4216 (1980). A suitable selection gene for use in yeast is the *trp1* gene present in the yeast plasmid YRp7 [Stinchcomb *et al.*, Nature, 282:39 (1979); Kingsman *et al.*, Gene, 7:141 (1979); Tschemper *et al.*, Gene, 10:157 (1980)]. The *trp1* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076 or PEP4-1 [Jones, Genetics, 85:12 (1977)].

Expression and cloning vectors usually contain a promoter operably linked to the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding nucleic acid sequence to direct mRNA synthesis. Promoters recognized by a variety of potential host cells are well known. Promoters suitable for use with prokaryotic hosts include the β -lactamase and lactose promoter systems [Chang *et al.*, Nature, 275:615 (1978); Goeddel *et al.*, Nature, 281:544 (1979)], alkaline phosphatase, a tryptophan (*trp*) promoter system [Goeddel, Nucleic Acids Res., 8:4057 (1980); EP 36,776], and hybrid promoters such as the tac promoter [deBoer *et al.*, Proc. Natl. Acad. Sci. USA, 80:21-25 (1983)]. Promoters for use in bacterial systems also will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198.

Examples of suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase [Hitzeman *et al.*, J. Biol. Chem., 255:2073 (1980)] or other glycolytic enzymes [Hess *et al.*, J. Adv. Enzyme Reg., 7:149 (1968); Holland, Biochemistry, 17:4900 (1978)], such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in EP 73,657.

PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198

transcription from vectors in mammalian host cells is controlled, for example, by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), from heterologous mammalian promoters, *e.g.*, the actin promoter or an immunoglobulin promoter, and from heat-shock promoters, provided such promoters are compatible with the host cell systems.

Transcription of a DNA encoding the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 by higher eukaryotes may be increased by inserting an enhancer sequence into the vector.

Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. The enhancer may be spliced into the vector at a position 5' or 3' to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 coding sequence, but is preferably located at a site 5' from the promoter.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3', untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198.

Still other methods, vectors, and host cells suitable for adaptation to the synthesis of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 in recombinant vertebrate cell culture are described in Gething *et al.*, Nature, 293:620-625 (1981); Mantei *et al.*, Nature, 281:40-46 (1979); EP 117,060; and EP 117,058.

d. Detecting Gene Amplification/Expression

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA [Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 (1980)], dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies may be employed that can recognize specific

duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of cells or tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. Antibodies useful for immunohistochemical staining and/or assay of sample fluids may be either monoclonal or polyclonal, and may be prepared in any mammal. Conveniently, the antibodies may be prepared against a native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide or against a synthetic peptide based on the DNA sequences provided herein or against an exogenous sequence fused to PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 DNA and encoding a specific antibody epitope.

e. Purification of Polypeptide

Forms of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 may be recovered from culture medium or from host cell lysates. If membrane-bound, it can be released from the membrane using a suitable detergent solution (*e.g.*, Triton-X 100) or by enzymatic cleavage. Cells employed in expression of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can be disrupted by various physical or chemical means, such as freeze-thaw cycling, sonication, mechanical disruption, or cell lysing agents.

It may be desired to purify PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 from recombinant cell proteins or polypeptides. The following procedures are exemplary of suitable purification procedures: by fractionation on an ion-exchange column; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation-exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel filtration using, for example, Sephadex G-75; protein A Sepharose columns to remove contaminants such as IgG; and metal chelating columns to bind epitope-tagged forms of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. Various methods of protein purification may

be employed and such methods are known in the art and described for example in Deutscher, Methods in Enzymology, 182 (1990); Scopes, Protein Purification: Principles and Practice, Springer-Verlag, New York (1982). The purification step(s) selected will depend, for example, on the nature of the production process used and the particular PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 produced.

E. Amplification of Genes Encoding the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 Polypeptides in Tumor Tissues and Cell Lines

The present invention is based on the identification and characterization of genes that are amplified in certain cancer cells.

The genome of prokaryotic and eukaryotic organisms is subjected to two seemingly conflicting requirements. One is the preservation and propagation of DNA as the genetic information in its original form, to guarantee stable inheritance through multiple generations. On the other hand, cells or organisms must be able to adapt to lasting environmental changes. The adaptive mechanisms can include qualitative or quantitative modifications of the genetic material. Qualitative modifications include DNA mutations, in which coding sequences are altered resulting in a structurally and/or functionally different protein. Gene amplification is a quantitative modification, whereby the actual number of complete coding sequence, *i.e.*, a gene, increases, leading to an increased number of available templates for transcription, an increased number of translatable transcripts, and, ultimately, to an increased abundance of the protein encoded by the amplified gene.

The phenomenon of gene amplification and its underlying mechanisms have been investigated *in vitro* in several prokaryotic and eukaryotic culture systems. The best-characterized example of gene amplification involves the culture of eukaryotic cells in medium containing variable concentrations of the cytotoxic drug methotrexate (MTX). MTX is a folic acid analogue and interferes with DNA synthesis by blocking the enzyme dihydrofolate reductase (DHFR). During the initial exposure to low concentrations of MTX most cells (>99.9%) will die. A small number of cells survive, and are capable of growing in increasing concentrations of MTX by producing large amounts of DHFR-RNA and protein. The basis of this overproduction is the amplification of the single DHFR gene. The additional copies of the gene are found as extrachromosomal copies in the form of small, supernumerary chromosomes (double minutes) or as integrated chromosomal copies.

Gene amplification is most commonly encountered in the development of resistance to cytotoxic drugs (antibiotics for bacteria and chemotherapeutic agents for eukaryotic cells) and neoplastic transformation. Transformation of a eukaryotic cell as a spontaneous event or due to a viral or chemical/environmental insult is typically associated with changes in the genetic material of that cell. One of the most common genetic changes observed in human malignancies are mutations of the p53 protein. p53 controls the transition of cells from the stationary (G1) to the replicative (S) phase and prevents this transition in the presence of DNA damage. In other words, one of the main consequences of disabling p53 mutations is the accumulation and propagation of DNA

damage, *i.e.*, genetic changes. Common types of genetic changes in neoplastic cells are, in addition to point mutations, amplifications and gross, structural alterations, such as translocations.

The amplification of DNA sequences may indicate specific functional requirement as illustrated in the DHFR experimental system. Therefore, the amplification of certain oncogenes in malignancies points toward a causative role of these genes in the process of malignant transformation and maintenance of the transformed phenotype. This hypothesis has gained support in recent studies. For example, the *bcl-2* protein was found to be amplified in certain types of non-Hodgkin's lymphoma. This protein inhibits apoptosis and leads to the progressive accumulation of neoplastic cells. Members of the gene family of growth factor receptors have been found to be amplified in various types of cancers suggesting that overexpression of these receptors may make neoplastic cells less susceptible to limiting amounts of available growth factor. Examples include the amplification of the androgen receptor in recurrent prostate cancer during androgen deprivation therapy and the amplification of the growth factor receptor homologue ERB2 in breast cancer. Lastly, genes involved in intracellular signaling and control of cell cycle progression can undergo amplification during malignant transformation. This is illustrated by the amplification of the *bcl-1* and *ras* genes in various epithelial and lymphoid neoplasms.

These earlier studies illustrate the feasibility of identifying amplified DNA sequences in neoplasms, because this approach can identify genes important for malignant transformation. The case of ERB2 also demonstrates the feasibility from a therapeutic standpoint, since transforming proteins may represent novel and specific targets for tumor therapy.

Several different techniques can be used to demonstrate amplified genomic sequences. Classical cytogenetic analysis of chromosome spreads prepared from cancer cells is adequate to identify gross structural alterations, such as translocations, deletions and inversions. Amplified genomic regions can only be visualized, if they involve large regions with high copy numbers or are present as extrachromosomal material. While cytogenetics was the first technique to demonstrate the consistent association of specific chromosomal changes with particular neoplasms, it is inadequate for the identification and isolation of manageable DNA sequences. The more recently developed technique of comparative genomic hybridization (CGH) has illustrated the widespread phenomenon of genomic amplification in neoplasms. Tumor and normal DNA are hybridized simultaneously onto metaphases of normal cells and the entire genome can be screened by image analysis for DNA sequences that are present in the tumor at an increased frequency. (WO 93/18,186; Gray *et al.*, Radiation Res., 137:275-289 [1994]). As a screening method, this type of analysis has revealed a large number of recurring amplicons (a stretch of amplified DNA) in a variety of human neoplasms. Although CGH is more sensitive than classical cytogenetic analysis in identifying amplified stretches of DNA, it does not allow a rapid identification and isolation of coding sequences within the amplicon by standard molecular genetic techniques.

The most sensitive methods to detect gene amplification are polymerase chain reaction (PCR)-based assays. These assays utilize very small amount of tumor DNA as starting material, are exquisitely sensitive, provide DNA that is amenable to further analysis, such as sequencing and are suitable for high-volume throughput analysis.

The above-mentioned assays are not mutually exclusive, but are frequently used in combination to identify amplifications in neoplasms. While cytogenetic analysis and CGH represent screening methods to survey the entire genome for amplified regions, PCR-based assays are most suitable for the final identification of coding sequences,

i.e., genes in amplified regions.

According to the present invention, such genes have been identified by quantitative PCR (S. Gelmini *et al.*, Clin. Chem., 43:752 [1997]), by comparing DNA from a variety of primary tumors, including breast, lung, colon, prostate, brain, liver, kidney, pancreas, spleen, thymus, testis, ovary, uterus, etc. tumor, or tumor cell lines, with
5 pooled DNA from healthy donors. Quantitative PCR was performed using a TaqMan instrument (ABI). Gene-specific primers and fluorogenic probes were designed based upon the coding sequences of the DNAs.

Human lung carcinoma cell lines include A549 (SRCC768), Calu-1 (SRCC769), Calu-6 (SRCC770), H157 (SRCC771), H441 (SRCC772), H460 (SRCC773), H522 (SRCC832), H810 (SRCC833), SKMES-1 (SRCC774) and SW900 (SRCC775), all available from ATCC. Primary human lung tumor cells usually derive from
10 adenocarcinomas, squamous cell carcinomas, large cell carcinomas, non-small cell carcinomas, small cell carcinomas, and broncho alveolar carcinomas, and include, for example, SRCC724 (adenocarcinoma, abbreviated as "AdenoCa")(LT1), SRCC725 (squamous cell carcinoma, abbreviated as "SqCCa")(LT1a), SRCC726 (adenocarcinoma)(LT2), SRCC727 (adenocarcinoma)(LT3), SRCC728 (adenocarcinoma)(LT4), SRCC729 (squamous cell carcinoma)(LT6), SRCC730 (adeno/squamous cell carcinoma)(LT7), SRCC825
15 (adenocarcinoma)(LT8), SRCC731 (adenocarcinoma)(LT9), SRCC732 (squamous cell carcinoma)(LT10), SRCC733 (squamous cell carcinoma)(LT11), SRCC734 (adenocarcinoma)(LT12), SRCC735 (adeno/squamous cell carcinoma)(LT13), SRCC736 (squamous cell carcinoma)(LT15), SRCC737 (squamous cell carcinoma)(LT16), SRCC738 (squamous cell carcinoma)(LT17), SRCC739 (squamous cell carcinoma)(LT18), SRCC740 (squamous cell carcinoma)(LT19), SRCC741 (lung cell carcinoma, abbreviated as "LCCa")(LT21), SRCC811
20 (adenocarcinoma)(LT22).

Colon cancer cell lines include, for example, ATCC cell lines SW480 (adenocarcinoma, SRCC776), SW620 (lymph node metastasis of colon adenocarcinoma, SRCC777), Colo320 (carcinoma, SRCC778), Colo205 (carcinoma, SRCC828), HCC2998 (carcinoma, SRCC830), HT29 (adenocarcinoma, SRCC779), HM7 (carcinoma, SRCC780), KM12 (carcinoma, SRCC831), CaWiDr (adenocarcinoma, SRCC781), HCT15 (carcinoma, SRCC829),
25 HCT116 (carcinoma, SRCC782), SKCO1 (adenocarcinoma, SRCC783), SW403 (adenocarcinoma, SRCC784), LS174T (carcinoma, SRCC785), and HM7 (a high mucin producing variant of ATCC colon adenocarcinoma cell line LS 174T, SRCC780, obtained from Dr. Robert Warren, UCSF). Primary colon tumors include colon adenoocarcinomas designated CT1 (SRCC751), CT2 (SRCC742), CT3 (SRCC743), CT4 (SRCC752), CT5 (SRCC753), CT6 (SRCC754), CT7 (SRCC755), CT8 (SRCC744), CT9 (SRCC756), CT10 (SRCC745), CT11
30 (SRCC757), CT12 (SRCC746), CT14 (SRCC747), CT15 (SRCC748), CT16 (SRCC749), CT17 (SRCC750), CT18 (SRCC758).

Human breast carcinoma cell lines include, for example, HBL100 (SRCC759), MB435s (SRCC760), T47D (SRCC761), MB468(SRCC762), MB175 (SRCC763), MB361 (SRCC764), BT20 (SRCC765), MCF7 (SRCC766), SKBR3 (SRCC767).

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F. Tissue Distribution

The results of the gene amplification assays herein can be verified by further studies, such as, by determining mRNA expression in various human tissues.

As noted before, gene amplification and/or gene expression in various tissues may be measured by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes.

Gene expression in various tissues, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. Antibodies useful for immunohistochemical staining and/or assay of sample fluids may be either monoclonal or polyclonal, and may be prepared in any mammal. Conveniently, the antibodies may be prepared against a native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide or against a synthetic peptide based on the DNA sequences provided herein or against exogenous sequence fused to sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 DNA and encoding a specific antibody epitope. General techniques for generating antibodies, and special protocols for Northern blotting and *in situ* hybridization are provided hereinbelow.

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G. Chromosome Mapping

If the amplification of a given gene is functionally relevant, then that gene should be amplified more than neighboring genomic regions which are not important for tumor survival. To test this, the gene can be mapped to a particular chromosome, *e.g.*, by radiation-hybrid analysis. The amplification level is then determined at the location identified, and at neighboring genomic region. Selective or preferential amplification at the genomic region to which the gene has been mapped is consistent with the possibility that the gene amplification observed promotes tumor growth or survival. Chromosome mapping includes both framework and epicenter mapping. For further details see *e.g.*, Stewart *et al.*, Genome Research, 7:422-433 (1997).

H. Antibody Binding Studies

The results of the gene amplification study can be further verified by antibody binding studies, in which the ability of anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145, or anti-PRO2198 to inhibit the expression of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094,

PRO2145 or PRO2198 polypeptides on tumor (cancer) cells is tested. Exemplary antibodies include polyclonal, monoclonal, humanized, bispecific, and heteroconjugate antibodies, the preparation of which will be described hereinbelow.

Antibody binding studies may be carried out in any known assay method, such as competitive binding assays, direct and indirect sandwich assays, and immunoprecipitation assays. Zola, Monoclonal Antibodies: A Manual of Techniques, pp.147-158 (CRC Press, Inc., 1987).

Competitive binding assays rely on the ability of a labeled standard to compete with the test sample analyte for binding with a limited amount of antibody. The amount of target protein (encoded by a gene amplified in a tumor cell) in the test sample is inversely proportional to the amount of standard that becomes bound to the antibodies. To facilitate determining the amount of standard that becomes bound, the antibodies preferably are insolubilized before or after the competition, so that the standard and analyte that are bound to the antibodies may conveniently be separated from the standard and analyte which remain unbound.

Sandwich assays involve the use of two antibodies, each capable of binding to a different immunogenic portion, or epitope, of the protein to be detected. In a sandwich assay, the test sample analyte is bound by a first antibody which is immobilized on a solid support, and thereafter a second antibody binds to the analyte, thus forming an insoluble three-part complex. See, *e.g.*, U.S. Patent No. 4,376,110. The second antibody may itself be labeled with a detectable moiety (direct sandwich assays) or may be measured using an anti-immunoglobulin antibody that is labeled with a detectable moiety (indirect sandwich assay). For example, one type of sandwich assay is an ELISA assay, in which case the detectable moiety is an enzyme.

For immunohistochemistry, the tumor sample may be fresh or frozen or may be embedded in paraffin and fixed with a preservative such as formalin, for example.

I. Cell-Based Tumor Assays

Cell-based assays and animal models for tumors (*e.g.*, cancers) can be used to verify the findings of the gene amplification assay, and further understand the relationship between the genes identified herein and the development and pathogenesis of neoplastic cell growth. The role of gene products identified herein in the development and pathology of tumor or cancer can be tested by using primary tumor cells or cells lines that have been identified to amplify the genes herein. Such cells include, for example, the breast, colon and lung cancer cells and cell lines listed above.

In a different approach, cells of a cell type known to be involved in a particular tumor are transfected with the cDNAs herein, and the ability of these cDNAs to induce excessive growth is analyzed. Suitable cells include, for example, stable tumor cells lines such as, the B104-1-1 cell line (stable NIH-3T3 cell line transfected with the *neu* protooncogene) and *ras*-transfected NIH-3T3 cells, which can be transfected with the desired gene, and monitored for tumorigenic growth. Such transfected cell lines can then be used to test the ability of poly- or monoclonal antibodies or antibody compositions to inhibit tumorigenic cell growth by exerting cytostatic or cytotoxic activity on the growth of the transformed cells, or by mediating antibody-dependent cellular cytotoxicity (ADCC). Cells transfected with the coding sequences of the genes identified herein can further be used to identify drug candidates for the treatment of cancer.

In addition, primary cultures derived from tumors in transgenic animals (as described below) can be used in the cell-based assays herein, although stable cell lines are preferred. Techniques to derive continuous cell lines from transgenic animals are well known in the art (see, *e.g.*, Small *et al.*, Mol. Cell. Biol., 5:642-648 [1985]).

5 J. Animal Models

A variety of well known animal models can be used to further understand the role of the genes identified herein in the development and pathogenesis of tumors, and to test the efficacy of candidate therapeutic agents, including antibodies, and other antagonists of the native polypeptides, including small molecule antagonists. The *in vivo* nature of such models makes them particularly predictive of responses in human patients. Animal models of tumors and cancers (*e.g.*, breast cancer, colon cancer, prostate cancer, lung cancer, etc.) include both non-recombinant and recombinant (transgenic) animals. Non-recombinant animal models include, for example, rodent, *e.g.*, murine models. Such models can be generated by introducing tumor cells into syngeneic mice using standard techniques, *e.g.*, subcutaneous injection, tail vein injection, spleen implantation, intraperitoneal implantation, implantation under the renal capsule, or orthopin implantation, *e.g.*, colon cancer cells implanted in colonic tissue. 15 (See, *e.g.*, PCT publication No. WO 97/33551, published September 18, 1997).

Probably the most often used animal species in oncological studies are immunodeficient mice and, in particular, nude mice. The observation that the nude mouse with hypo/aplasia could successfully act as a host for human tumor xenografts has lead to its widespread use for this purpose. The autosomal recessive *nu* gene has been introduced into a very large number of distinct congenic strains of nude mouse, including, for example, ASW, A/He, 20 AKR, BALB/c, B10.LP, C17, C3H, C57BL, C57, CBA, DBA, DDD, I/st, NC, NFR, NFS, NFS/N, NZB, NZC, NZW, P, RIII and SJL. In addition, a wide variety of other animals with inherited immunological defects other than the nude mouse have been bred and used as recipients of tumor xenografts. For further details see, *e.g.*, The Nude Mouse in Oncology Research, E. Boven and B. Winograd, eds., CRC Press, Inc., 1991.

The cells introduced into such animals can be derived from known tumor/cancer cell lines, such as, any of 25 the above-listed tumor cell lines, and, for example, the B104-1-1 cell line (stable NIH-3T3 cell line transfected with the *neu* protooncogene); *ras*-transfected NIH-3T3 cells; Caco-2 (ATCC HTB-37); a moderately well-differentiated grade II human colon adenocarcinoma cell line, HT-29 (ATCC HTB-38), or from tumors and cancers. Samples of tumor or cancer cells can be obtained from patients undergoing surgery, using standard conditions, involving freezing and storing in liquid nitrogen (Karmali *et al.*, Br. J. Cancer, 48:689-696 [1983]).

30 Tumor cells can be introduced into animals, such as nude mice, by a variety of procedures. The subcutaneous (s.c.) space in mice is very suitable for tumor implantation. Tumors can be transplanted s.c. as solid blocks, as needle biopsies by use of a trochar, or as cell suspensions. For solid block or trochar implantation, tumor tissue fragments of suitable size are introduced into the s.c. space. Cell suspensions are freshly prepared from primary tumors or stable tumor cell lines, and injected subcutaneously. Tumor cells can also be injected as 35 subdermal implants. In this location, the inoculum is deposited between the lower part of the dermal connective tissue and the s.c. tissue. Boven and Winograd (1991), *supra*.

Animal models of breast cancer can be generated, for example, by implanting rat neuroblastoma cells (from which the *neu* oncogen was initially isolated), or *neu*-transformed NIH-3T3 cells into nude mice, essentially as

described by Drebin *et al.*, PNAS USA, 83:9129-9133 (1986).

Similarly, animal models of colon cancer can be generated by passaging colon cancer cells in animals, *e.g.*, nude mice, leading to the appearance of tumors in these animals. An orthotopic transplant model of human colon cancer in nude mice has been described, for example, by Wang *et al.*, Cancer Research, 54:4726-4728 (1994) and
5 Too *et al.*, Cancer Research, 55:681-684 (1995). This model is based on the so-called "METAMOUSE" sold by AntiCancer, Inc. (San Diego, California).

Tumors that arise in animals can be removed and cultured *in vitro*. Cells from the *in vitro* cultures can then be passaged to animals. Such tumors can serve as targets for further testing or drug screening. Alternatively, the tumors resulting from the passage can be isolated and RNA from pre-passage cells and cells isolated after one or
10 more rounds of passage analyzed for differential expression of genes of interest. Such passaging techniques can be performed with any known tumor or cancer cell lines.

For example, Meth A, CMS4, CMS5, CMS21, and WEHI-164 are chemically induced fibrosarcomas of BALB/c female mice (DeLeo *et al.*, J. Exp. Med., 146:720 [1977]), which provide a highly controllable model system for studying the anti-tumor activities of various agents (Palladino *et al.*, J. Immunol., 138:4023-4032 [1987]).
15 Briefly, tumor cells are propagated *in vitro* in cell culture. Prior to injection into the animals, the cell lines are washed and suspended in buffer, at a cell density of about 10×10^6 to 10×10^7 cells/ml. The animals are then infected subcutaneously with 10 to 100 μ l of the cell suspension, allowing one to three weeks for a tumor to appear.

In addition, the Lewis lung (3LL) carcinoma of mice, which is one of the most thoroughly studied experimental tumors, can be used as an investigational tumor model. Efficacy in this tumor model has been
20 correlated with beneficial effects in the treatment of human patients diagnosed with small cell carcinoma of the lung (SCCL). This tumor can be introduced in normal mice upon injection of tumor fragments from an affected mouse or of cells maintained in culture (Zupi *et al.*, Br. J. Cancer, 41:suppl. 4:309 [1980]), and evidence indicates that tumors can be started from injection of even a single cell and that a very high proportion of infected tumor cells survive. For further information about this tumor model see Zacharski, Haemostasis, 16:300-320 [1986]).

One way of evaluating the efficacy of a test compound in an animal model is implanted tumor is to measure
25 the size of the tumor before and after treatment. Traditionally, the size of implanted tumors has been measured with a slide caliper in two or three dimensions. The measure limited to two dimensions does not accurately reflect the size of the tumor, therefore, it is usually converted into the corresponding volume by using a mathematical formula. However, the measurement of tumor size is very inaccurate. The therapeutic effects of a drug candidate can be better
30 described as treatment-induced growth delay and specific growth delay. Another important variable in the description of tumor growth is the tumor volume doubling time. Computer programs for the calculation and description of tumor growth are also available, such as the program reported by Rygaard and Spang-Thomsen, Proc. 6th Int. Workshop on Immune-Deficient Animals, Wu and Sheng eds., Basel, 1989, 301. It is noted, however, that necrosis and inflammatory responses following treatment may actually result in an increase in tumor size, at least
35 initially. Therefore, these changes need to be carefully monitored, by a combination of a morphometric method and flow cytometric analysis.

Recombinant (transgenic) animal models can be engineered by introducing the coding portion of the genes identified herein into the genome of animals of interest, using standard techniques for producing transgenic animals.

Animals that can serve as a target for transgenic manipulation include, without limitation, mice, rats, rabbits, guinea pigs, sheep, goats, pigs, and non-human primates, *e.g.*, baboons, chimpanzees and monkeys. Techniques known in the art to introduce a transgene into such animals include pronucleic microinjection (Hoppe and Wanger, U.S. Patent No. 4,873,191); retrovirus-mediated gene transfer into germ lines (*e.g.*, Van der Putten *et al.*, Proc. Natl. Acad. Sci. USA, 82:6148-615 [1985]); gene targeting in embryonic stem cells (Thompson *et al.*, Cell, 56:313-321 [1989]); electroporation of embryos (Lo, Mol. Cell Biol., 3:1803-1814 [1983]); sperm-mediated gene transfer (Lavitrano *et al.*, Cell, 57:717-73 [1989]). For review, see, for example, U.S. Patent No. 4,736,866.

For the purpose of the present invention, transgenic animals include those that carry the transgene only in part of their cells ("mosaic animals"). The transgene can be integrated either as a single transgene, or in concatamers, *e.g.*, head-to-head or head-to-tail tandems. Selective introduction of a transgene into a particular cell type is also possible by following, for example, the technique of Laskó *et al.*, Proc. Natl. Acad. Sci. USA, 89:6232-636 (1992).

The expression of the transgene in transgenic animals can be monitored by standard techniques. For example, Southern blot analysis or PCR amplification can be used to verify the integration of the transgene. The level of mRNA expression can then be analyzed using techniques such as *in situ* hybridization, Northern blot analysis, PCR, or immunocytochemistry. The animals are further examined for signs of tumor or cancer development.

Alternatively, "knock out" animals can be constructed which have a defective or altered gene encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide identified herein, as a result of homologous recombination between the endogenous gene encoding the polypeptide and altered genomic DNA encoding the same polypeptide introduced into an embryonic cell of the animal. For example, cDNA encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide can be used to clone genomic DNA encoding that polypeptide in accordance with established techniques. A portion of the genomic DNA encoding a particular PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide can be deleted or replaced with another gene, such as a gene encoding a selectable marker which can be used to monitor integration. Typically, several kilobases of unaltered flanking DNA (both at the 5' and 3' ends) are included in the vector [see *e.g.*, Thomas and Capecchi, Cell, 51:503 (1987) for a description of homologous recombination vectors]. The vector is introduced into an embryonic stem cell line (*e.g.*, by electroporation) and cells in which the introduced DNA has homologously recombined with the endogenous DNA are selected [see *e.g.*, Li *et al.*, Cell, 69:915 (1992)]. The selected cells are then injected into a blastocyst of an animal (*e.g.*, a mouse or rat) to form aggregation chimeras [see *e.g.*, Bradley, in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E. J. Robertson, ed. (IRL, Oxford, 1987), pp. 113-152]. A chimeric embryo can

then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term to create a "knock out" animal. Progeny harboring the homologously recombined DNA in their germ cells can be identified by standard techniques and used to breed animals in which all cells of the animal contain the homologously recombined DNA. Knockout animals can be characterized for instance, by their ability to defend against certain pathological conditions and by their development of pathological conditions due to absence of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

The efficacy of antibodies specifically binding the polypeptides identified herein and other drug candidates, can be tested also in the treatment of spontaneous animal tumors. A suitable target for such studies is the feline oral squamous cell carcinoma (SCC). Feline oral SCC is a highly invasive, malignant tumor that is the most common oral malignancy of cats, accounting for over 60% of the oral tumors reported in this species. It rarely metastasizes to distant sites, although this low incidence of metastasis may merely be a reflection of the short survival times for cats with this tumor. These tumors are usually not amenable to surgery, primarily because of the anatomy of the feline oral cavity. At present, there is no effective treatment for this tumor. Prior to entry into the study, each cat undergoes complete clinical examination, biopsy, and is scanned by computed tomography (CT). Cats diagnosed with sublingual oral squamous cell tumors are excluded from the study. The tongue can become paralyzed as a result of such tumor, and even if the treatment kills the tumor, the animals may not be able to feed themselves. Each cat is treated repeatedly, over a longer period of time. Photographs of the tumors will be taken daily during the treatment period, and at each subsequent recheck. After treatment, each cat undergoes another CT scan. CT scans and thoracic radiograms are evaluated every 8 weeks thereafter. The data are evaluated for differences in survival, response and toxicity as compared to control groups. Positive response may require evidence of tumor regression, preferably with improvement of quality of life and/or increased life span.

In addition, other spontaneous animal tumors, such as fibrosarcoma, adenocarcinoma, lymphoma, chondroma, leiomyosarcoma of dogs, cats, and baboons can also be tested. Of these mammary adenocarcinoma in dogs and cats is a preferred model as its appearance and behavior are very similar to those in humans. However, the use of this model is limited by the rare occurrence of this type of tumor in animals.

K. Screening Assays for Drug Candidates

Screening assays for drug candidates are designed to identify compounds that bind or complex with the polypeptides encoded by the genes identified herein, or otherwise interfere with the interaction of the encoded polypeptides with other cellular proteins. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates. Small molecules contemplated include synthetic organic or inorganic compounds, including peptides, preferably soluble peptides, (poly)peptide-immunoglobulin fusions, and, in particular, antibodies including, without limitation, poly- and monoclonal antibodies and antibody fragments, single-chain antibodies, anti-idiotypic antibodies, and chimeric or humanized versions of such antibodies or fragments, as well as human antibodies and antibody fragments. The assays can be performed in a variety of formats, including protein-protein binding assays,

biochemical screening assays, immunoassays and cell based assays, which are well characterized in the art.

All assays are common in that they call for contacting the drug candidate with a polypeptide encoded by a nucleic acid identified herein under conditions and for a time sufficient to allow these two components to interact.

In binding assays, the interaction is binding and the complex formed can be isolated or detected in the reaction mixture. In a particular embodiment, the polypeptide encoded by the gene identified herein or the drug candidate is immobilized on a solid phase, *e.g.*, on a microtiter plate, by covalent or non-covalent attachments. Non-covalent attachment generally is accomplished by coating the solid surface with a solution of the polypeptide and drying. Alternatively, an immobilized antibody, *e.g.*, a monoclonal antibody, specific for the polypeptide to be immobilized can be used to anchor it to a solid surface. The assay is performed by adding the non-immobilized component, which may be labeled by a detectable label, to the immobilized component, *e.g.*, the coated surface containing the anchored component. When the reaction is complete, the non-reacted components are removed, *e.g.*, by washing, and complexes anchored on the solid surface are detected. When the originally non-immobilized component carries a detectable label, the detection of label immobilized on the surface indicates that complexing occurred. Where the originally non-immobilized component does not carry a label, complexing can be detected, for example, by using a labeled antibody specifically binding the immobilized complex.

If the candidate compound interacts with but does not bind to a particular PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide encoded by a gene identified herein, its interaction with that polypeptide can be assayed by methods well known for detecting protein-protein interactions. Such assays include traditional approaches, such as, cross-linking, co-immunoprecipitation, and co-purification through gradients or chromatographic columns. In addition, protein-protein interactions can be monitored by using a yeast-based genetic system described by Fields and co-workers [Fields and Song, *Nature*, 340:245-246 (1989); Chien *et al.*, *Proc. Natl. Acad. Sci. USA*, 88: 9578-9582 (1991)] as disclosed by Chevray and Nathans, *Proc. Natl. Acad. Sci. USA*, 89:5789-5793 (1991)]. Many transcriptional activators, such as yeast GAL4, consist of two physically discrete modular domains, one acting as the DNA-binding domain, while the other one functioning as the transcription activation domain. The yeast expression system described in the foregoing publications (generally referred to as the "two-hybrid system") takes advantage of this property, and employs two hybrid proteins, one in which the target protein is fused to the DNA-binding domain of GAL4, and another, in which candidate activating proteins are fused to the activation domain. The expression of a GAL1-*lacZ* reporter gene under control of a GAL4-activated promoter depends on reconstitution of GAL4 activity via protein-protein interaction. Colonies containing interacting polypeptides are detected with a chromogenic substrate for β -galactosidase. A complete kit (MATCHMAKER™) for identifying protein-protein interactions between two specific proteins using the two-hybrid technique is commercially available from Clontech. This system can also be extended to map protein domains involved in specific protein interactions as well as to pinpoint amino acid residues that are crucial for these interactions.

Compounds that interfere with the interaction of a PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-,

PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding gene identified herein and other intra- or extracellular components can be tested as follows: usually a reaction mixture is prepared containing the product of the amplified gene and the intra- or extracellular component under conditions and for a time allowing for the interaction and binding of the two products. To test the ability of a test compound to inhibit binding, the reaction is run in the absence and in the presence of the test compound. In addition, a placebo may be added to a third reaction mixture, to serve as positive control. The binding (complex formation) between the test compound and the intra- or extracellular component present in the mixture is monitored as described hereinabove. The formation of a complex in the control reaction(s) but not in the reaction mixture containing the test compound indicates that the test compound interferes with the interaction of the test compound and its reaction partner.

To assay for antagonists, the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide may be added to a cell along with the compound to be screened for a particular activity and the ability of the compound to inhibit the activity of interest in the presence of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide indicates that the compound is an antagonist to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Alternatively, antagonists may be detected by combining the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide and a potential antagonist with membrane-bound PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide receptors or recombinant receptors under appropriate conditions for a competitive inhibition assay. The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide can be labeled, such as by radioactivity, such that the number of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide molecules bound to the receptor can be used to determine the effectiveness of the potential antagonist. The gene encoding the receptor can be identified by numerous methods known to those of skill in the art, for example, ligand panning and FACS sorting. Coligan *et al.*, Current Protocols in Immun., 1(2): Chapter 5 (1991).

Preferably, expression cloning is employed wherein polyadenylated RNA is prepared from a cell responsive to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide and a cDNA library created from this RNA is divided into pools and used to transfect COS cells or other cells that are not responsive to the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Transfected cells that are grown on glass slides are exposed to labeled PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide can be labeled by a variety of means including iodination or inclusion of a recognition site for a site-specific protein kinase. Following fixation and incubation, the slides are subjected to autoradiographic analysis. Positive pools are identified and sub-pools are prepared and re-transfected using an interactive sub-pooling and re-screening process, eventually yielding a single clone that encodes the putative receptor.

As an alternative approach for receptor identification, labeled PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide can be photoaffinity-linked with cell membrane or extract preparations that express the receptor molecule. Cross-linked material is resolved by PAGE and exposed to X-ray film. The labeled complex containing the receptor can be excised, resolved into peptide fragments, and subjected to protein micro-sequencing. The amino acid sequence obtained from micro-sequencing would be used to design a set of degenerate oligonucleotide probes to screen a cDNA library to identify the gene encoding the putative receptor.

In another assay for antagonists, mammalian cells or a membrane preparation expressing the receptor would be incubated with labeled PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in the presence of the candidate compound. The ability of the compound to enhance or block this interaction could then be measured.

More specific examples of potential antagonists include an oligonucleotide that binds to the fusions of immunoglobulin with PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or

PRO2198 polypeptide, and, in particular, antibodies including, without limitation, poly- and monoclonal antibodies and antibody fragments, single-chain antibodies, anti-idiotypic antibodies, and chimeric or humanized versions of such antibodies or fragments, as well as human antibodies and antibody fragments. Alternatively, a potential antagonist may be a closely related protein, for example, a mutated form of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide that recognizes the receptor but imparts no effect, thereby competitively inhibiting the action of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

Another potential PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide antagonist is an antisense RNA or DNA construct prepared using antisense technology, where, *e.g.*, an antisense RNA or DNA molecule acts to block directly the translation of mRNA by hybridizing to targeted mRNA and preventing protein translation. Antisense technology can be used to control gene expression through triple-helix formation or antisense DNA or RNA, both of which methods are based on binding of a polynucleotide to DNA or RNA. For example, the 5' coding portion of the polynucleotide sequence, which encodes the mature PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptides herein, is used to design an antisense RNA oligonucleotide of from about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene involved in transcription (triple helix - see Lee *et al.*, Nucl. Acids Res., 6:3073 (1979); Cooney *et al.*, Science, 241: 456 (1988); Dervan *et al.*, Science, 251:1360 (1991)), thereby preventing transcription and the production of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. The antisense RNA oligonucleotide hybridizes to the mRNA *in vivo* and blocks translation of the mRNA molecule into the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide (antisense - Okano, Neurochem., 56:560 (1991); Oligodeoxynucleotides as Antisense Inhibitors of Gene Expression (CRC Press: Boca Raton, FL, 1988). The oligonucleotides described above can also be delivered to cells such that the antisense RNA or DNA may be expressed *in vivo* to inhibit production of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094,

PRO2145 or PRO2198 polypeptide. When antisense DNA is used, oligodeoxyribonucleotides derived from the translation-initiation site, *e.g.*, between about -10 and +10 positions of the target gene nucleotide sequence, are preferred.

Potential antagonists include small molecules that bind to the active site, the receptor binding site, or growth factor or other relevant binding site of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, thereby blocking the normal biological activity of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. Examples of small molecules include, but are not limited to, small peptides or peptide-like molecules, preferably soluble peptides, and synthetic non-peptidyl organic or inorganic compounds.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. Ribozymes act by sequence-specific hybridization to the complementary target RNA, followed by endonucleolytic cleavage. Specific ribozyme cleavage sites within a potential RNA target can be identified by known techniques. For further details see, *e.g.*, Rossi, Current Biology, 4:469-471 (1994), and PCT publication No. WO 97/33551 (published September 18, 1997).

Nucleic acid molecules in triple-helix formation used to inhibit transcription should be single-stranded and composed of deoxynucleotides. The base composition of these oligonucleotides is designed such that it promotes triple-helix formation via Hoogsteen base-pairing rules, which generally require sizeable stretches of purines or pyrimidines on one strand of a duplex. For further details see, *e.g.*, PCT publication No. WO 97/33551, *supra*.

These small molecules can be identified by any one or more of the screening assays discussed hereinabove and/or by any other screening techniques well known for those skilled in the art.

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L. Compositions and Methods for the Treatment of Tumors

The compositions useful in the treatment of tumors associated with the amplification of the genes identified herein include, without limitation, antibodies, small organic and inorganic molecules, peptides, phosphopeptides, antisense and ribozyme molecules, triple helix molecules, etc. that inhibit the expression and/or activity of the target gene product.

For example, antisense RNA and RNA molecule act to directly block the translation of mRNA by hybridizing to targeted mRNA and preventing protein translation. When antisense DNA is used, oligodeoxyribonucleotides derived from the translation initiation site, *e.g.*, between about -10 and +10 positions of the target gene nucleotide sequence, are preferred.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. Ribozymes act by sequence-specific hybridization to the complementary target RNA, followed by endonucleolytic cleavage. Specific ribozyme cleavage sites within a potential RNA target can be identified by known techniques. For further details see, *e.g.*, Rossi, Current Biology, 4:469-471 (1994), and PCT publication No. WO 97/33551 (published

September 18, 1997).

Nucleic acid molecules in triple helix formation used to inhibit transcription should be single-stranded and composed of deoxynucleotides. The base composition of these oligonucleotides is designed such that it promotes triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of purines or pyrimidines on one strand of a duplex. For further details see, *e.g.*, PCT publication No. WO 97/33551, *supra*.

These molecules can be identified by any or any combination of the screening assays discussed hereinabove and/or by any other screening techniques well known for those skilled in the art.

M. Antibodies

Some of the most promising drug candidates according to the present invention are antibodies and antibody fragments which may inhibit the production or the gene product of the amplified genes identified herein and/or reduce the activity of the gene products.

1. Polyclonal Antibodies

Methods of preparing polyclonal antibodies are known to the skilled artisan. Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal injections. The immunizing agent may include the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide or a fusion protein thereof. It may be useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized. Examples of such immunogenic proteins include but are not limited to keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, and soybean trypsin inhibitor. Examples of adjuvants which may be employed include Freund's complete adjuvant and MPL-TDM adjuvant (monophosphoryl Lipid A, synthetic trehalose dicorynomycolate). The immunization protocol may be selected by one skilled in the art without undue experimentation.

2. Monoclonal Antibodies

The anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibodies may, alternatively, be monoclonal antibodies. Monoclonal antibodies may be prepared using hybridoma methods, such as those described by Kohler and Milstein, *Nature*, 256:495 (1975). In a hybridoma method, a mouse, hamster, or other appropriate host animal, is typically immunized with an immunizing agent to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the immunizing agent. Alternatively, the lymphocytes may be immunized *in vitro*.

The immunizing agent will typically include the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717,

PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, including fragments, or a fusion protein of such protein or a fragment thereof. Generally, either peripheral blood lymphocytes ("PBLs") are used if cells of human origin are desired, or spleen cells
5 or lymph node cells are used if non-human mammalian sources are desired. The lymphocytes are then fused with an immortalized cell line using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell [Goding, Monoclonal Antibodies: Principles and Practice, Academic Press, (1986) pp. 59-103]. Immortalized cell lines are usually transformed mammalian cells, particularly myeloma cells of rodent, bovine and human origin. Usually, rat or mouse myeloma cell lines are employed. The hybridoma cells may be cultured in a suitable culture
10 medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, immortalized cells. For example, if the parental cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine ("HAT medium"), which substances prevent the growth of HGPRT-deficient cells.

Preferred immortalized cell lines are those that fuse efficiently, support stable high level expression of
15 antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. More preferred immortalized cell lines are murine myeloma lines, which can be obtained, for instance, from the Salk Institute Cell Distribution Center, San Diego, California and the American Type Culture Collection (ATCC), Manassas, Virginia. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies [Kozbor, J. Immunol., 133:3001 (1984); Brodeur *et al.*, Monoclonal
20 Antibody Production Techniques and Applications, Marcel Dekker, Inc., New York, (1987) pp. 51-63].

The culture medium in which the hybridoma cells are cultured can then be assayed for the presence of monoclonal antibodies directed against PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094,
25 PRO2145 or PRO2198. Preferably, the binding specificity of monoclonal antibodies produced by the hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA). Such techniques and assays are known in the art. The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson and Pollard, Anal. Biochem., 107:220 (1980).

30 After the desired hybridoma cells are identified, the clones may be subcloned by limiting dilution procedures and grown by standard methods [Goding, *supra*]. Suitable culture media for this purpose include, for example, Dulbecco's Modified Eagle's Medium and RPMI-1640 medium. Alternatively, the hybridoma cells may be grown *in vivo* as ascites in a mammal.

The monoclonal antibodies secreted by the subclones may be isolated or purified from the culture medium
35 or ascites fluid by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

The monoclonal antibodies may also be made by recombinant DNA methods, such as those described in U.S. Patent No. 4,816,567. DNA encoding the monoclonal antibodies of the invention can be readily isolated and

sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells of the invention serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. The DNA also may be modified, for example, by substituting the coding sequence for human heavy and light chain constant domains in place of the homologous murine sequences [U.S. Patent No. 4,816,567; Morrison *et al.*, *supra*] or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide. Such a non-immunoglobulin polypeptide can be substituted for the constant domains of an antibody of the invention, or can be substituted for the variable domains of one antigen-combining site of an antibody of the invention to create a chimeric bivalent antibody.

The antibodies may be monovalent antibodies. Methods for preparing monovalent antibodies are well known in the art. For example, one method involves recombinant expression of immunoglobulin light chain and modified heavy chain. The heavy chain is truncated generally at any point in the Fc region so as to prevent heavy chain crosslinking. Alternatively, the relevant cysteine residues are substituted with another amino acid residue or are deleted so as to prevent crosslinking.

In vitro methods are also suitable for preparing monovalent antibodies. Digestion of antibodies to produce fragments thereof, particularly, Fab fragments, can be accomplished using routine techniques known in the art.

3. Human and Humanized Antibodies

The anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibodies may further comprise humanized antibodies or human antibodies. Humanized forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', F(ab')₂ or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. Humanized antibodies include human immunoglobulins (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, Fv framework residues of the human immunoglobulin are replaced by corresponding non-human residues. Humanized antibodies may also comprise residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin [Jones *et al.*, *Nature*, 321:522-525 (1986); Riechmann *et al.*, *Nature*, 332:323-329 (1988); and

Presta, Curr. Op. Struct. Biol., 2:593-596 (1992)].

Methods for humanizing non-human antibodies are well known in the art. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source which is non-human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain.

5 Humanization can be essentially performed following the method of Winter and co-workers [Jones *et al.*, Nature, 321:522-525 (1986); Riechmann *et al.*, Nature, 332:323-327 (1988); Verhoeven *et al.*, Science, 239:1534-1536 (1988)], by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Patent No. 4,816,567), wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from
10 a non-human species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

Human antibodies can also be produced using various techniques known in the art, including phage display libraries [Hoogenboom and Winter, J. Mol. Biol., 227:381 (1991); Marks *et al.*, J. Mol. Biol., 222:581 (1991)]. The techniques of Cole *et al.*, and Boerner *et al.*, are also available for the preparation of human monoclonal
15 antibodies (Cole *et al.*, Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985) and Boerner *et al.*, J. Immunol., 147(1):86-95 (1991)]. Similarly, human antibodies can be made by introducing of human immunoglobulin loci into transgenic animals, *e.g.*, mice in which the endogenous immunoglobulin genes have been partially or completely inactivated. Upon challenge, human antibody production is observed, which closely resembles that seen in humans in all respects, including gene rearrangement, assembly, and antibody repertoire. This
20 approach is described, for example, in U.S. Patent Nos. 5,545,807; 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,661,016, and in the following scientific publications: Marks *et al.*, Bio/Technology, 10:779-783 (1992); Lonberg *et al.*, Nature, 368:856-859 (1994); Morrison, Nature, 368:812-13 (1994); Fishwild *et al.*, Nature Biotechnology, 14:845-51 (1996); Neuberger, Nature Biotechnology, 14:826 (1996); Lonberg and Huszar, Intern. Rev. Immunol., 13:65-93 (1995).

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4. Antibody Dependent Enzyme Mediated Prodrug Therapy (ADEPT)

The antibodies of the present invention may also be used in ADEPT by conjugating the antibody to a prodrug-activating enzyme which converts a prodrug (*e.g.*, a peptidyl chemotherapeutic agent, see WO 81/01145) to an active anti-cancer drug. See, for example, WO 88/07378 and U. S. Patent No. 4,975,278.

30 The enzyme component of the immunoconjugate useful for ADEPT includes any enzyme capable of acting on a prodrug in such as way so as to convert it into its more active, cytotoxic form.

Enzymes that are useful in the method of this invention include, but are not limited to, glycosidase, glucose oxidase, human lysosyme, human glucuronidase, alkaline phosphatase useful for converting phosphate-containing prodrugs into free drugs; arylsulfatase useful for converting sulfate-containing prodrugs into free drugs; cytosine
35 deaminase useful for converting non-toxic 5-fluorocytosine into the anti-cancer drug 5-fluorouracil; proteases, such as serratia protease, thermolysin, subtilisin, carboxypeptidases (*e.g.*, carboxypeptidase G2 and carboxypeptidase A) and cathepsins (such as cathepsins B and L), that are useful for converting peptide-containing prodrugs into free drugs; D-alanylcarboxypeptidases, useful for converting prodrugs that contain D-amino acid substituents;

carbohydrate-cleaving enzymes such as β -galactosidase and neuraminidase useful for converting glycosylated prodrugs into free drugs; β -lactamase useful for converting drugs derivatized with β -lactams into free drugs; and penicillin amidases, such as penicillin Vamidase or penicillin G amidase, useful for converting drugs derivatized at their amine nitrogens with phenoxyacetyl or phenylacetyl groups, respectively, into free drugs. Alternatively, antibodies with enzymatic activity, also known in the art as "abzymes" can be used to convert the prodrugs of the invention into free active drugs (see, *e.g.*, Massey, Nature, 328:457-458 (1987)). Antibody-abzyme conjugates can be prepared as described herein for delivery of the abzyme to a tumor cell population.

The enzymes of this invention can be covalently bound to the anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibodies by techniques well known in the art such as the use of the heterobifunctional cross-linking agents discussed above. Alternatively, fusion proteins comprising at least the antigen binding region of the antibody of the invention linked to at least a functionally active portion of an enzyme of the invention can be constructed using recombinant DNA techniques well known in the art (see, *e.g.*, Neuberger *et al.*, Nature, 312:604-608 (1984)).

5. Bispecific Antibodies

Bispecific antibodies are monoclonal, preferably human or humanized, antibodies that have binding specificities for at least two different antigens. In the present case, one of the binding specificities is for the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 the other one is for any other antigen, and preferably for a cell-surface protein or receptor or receptor subunit.

Methods for making bispecific antibodies are known in the art. Traditionally, the recombinant production of bispecific antibodies is based on the co-expression of two immunoglobulin heavy-chain/light-chain pairs, where the two heavy chains have different specificities (Milstein and Cuello, Nature, 305:537-539 [1983]). Because of the random assortment of immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of ten different antibody molecules, of which only one has the correct bispecific structure. The purification of the correct molecule is usually accomplished by affinity chromatography steps. Similar procedures are disclosed in WO 93/08829, published 13 May 1993, and in Traunecker *et al.*, EMBO J., 10:3655-3659 (1991).

Antibody variable domains with the desired binding specificities (antibody-antigen combining sites) can be fused to immunoglobulin constant domain sequences. The fusion preferably is with an immunoglobulin heavy-chain constant domain, comprising at least part of the hinge, CH2, and CH3 regions. It is preferred to have the first heavy-chain constant region (CH1) containing the site necessary for light-chain binding present in at least one of the fusions. DNAs encoding the immunoglobulin heavy-chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host organism. For further details of generating bispecific antibodies see, for example, Suresh *et al.*, Methods in Enzymology, 121:210 (1986).

According to another approach described in WO 96/27011, the interface between a pair of antibody molecules can be engineered to maximize the percentage of heterodimers which are recovered from recombinant cell culture. The preferred interface comprises at least a part of the CH3 region of an antibody constant domain. In this method, one or more small amino acid side chains from the interface of the first antibody molecule are replaced with larger side chains (e.g., tyrosine or tryptophan). Compensatory "cavities" of identical or similar size to the large side chain(s) are created on the interface of the second antibody molecule by replacing large amino acid side chains with smaller ones (e.g., alanine or threonine). This provides a mechanism for increasing the yield of the heterodimer over other unwanted end-products such as homodimers.

Bispecific antibodies can be prepared as full length antibodies or antibody fragments (e.g., F(ab')₂ bispecific antibodies). Techniques for generating bispecific antibodies from antibody fragments have been described in the literature. For example, bispecific antibodies can be prepared using chemical linkage. Brennan *et al.*, Science, 229:81 (1985) describe a procedure wherein intact antibodies are proteolytically cleaved to generate F(ab')₂ fragments. These fragments are reduced in the presence of the dithiol complexing agent sodium arsenite to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The Fab' fragments generated are then converted to thionitrobenzoate (TNB) derivatives. One of the Fab'-TNB derivatives is then reconverted to the Fab'-thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other Fab'-TNB derivative to form the bispecific antibody. The bispecific antibodies produced can be used as agents for the selective immobilization of enzymes.

Fab' fragments may be directly recovered from *E. coli* and chemically coupled to form bispecific antibodies. Shalaby *et al.*, J. Exp. Med., 175:217-225 (1992) describe the production of a fully humanized bispecific antibody F(ab')₂ molecule. Each Fab' fragment was separately secreted from *E. coli* and subjected to directed chemical coupling *in vitro* to form the bispecific antibody. The bispecific antibody thus formed was able to bind to cells overexpressing the ErbB2 receptor and normal human T cells, as well as trigger the lytic activity of human cytotoxic lymphocytes against human breast tumor targets.

Various technique for making and isolating bispecific antibody fragments directly from recombinant cell culture have also been described. For example, bispecific antibodies have been produced using leucine zippers. Kostelny *et al.*, J. Immunol., 148(5):1547-1553 (1992). The leucine zipper peptides from the Fos and Jun proteins were linked to the Fab' portions of two different antibodies by gene fusion. The antibody homodimers were reduced at the hinge region to form monomers and then re-oxidized to form the antibody heterodimers. This method can also be utilized for the production of antibody homodimers. The "diabody" technology described by Hollinger *et al.*, Proc. Natl. Acad. Sci. USA, 90:6444-6448 (1993) has provided an alternative mechanism for making bispecific antibody fragments. The fragments comprise a heavy-chain variable domain (V_H) connected to a light-chain variable domain (V_L) by a linker which is too short to allow pairing between the two domains on the same chain. Accordingly, the V_H and V_L domains of one fragment are forced to pair with the complementary V_L and V_H domains of another fragment, thereby forming two antigen-binding sites. Another strategy for making bispecific antibody fragments by the use of single-chain Fv (sFv) dimers has also been reported. See, Gruber *et al.*, J. Immunol., 152:5368 (1994).

Antibodies with more than two valencies are contemplated. For example, trispecific antibodies can be

prepared. Tutt *et al.*, J. Immunol., 147:60 (1991).

Exemplary bispecific antibodies may bind to two different epitopes on a given polypeptide herein. Alternatively, an anti-polypeptide arm may be combined with an arm which binds to a triggering molecule on a leukocyte such as a T-cell receptor molecule (*e.g.*, CD2, CD3, CD28, or B7), or Fc receptors for IgG (FcγR), such as FcγRI (CD64), FcγRII (CD32) and FcγRIII (CD16) so as to focus cellular defense mechanisms to the cell expressing the particular polypeptide. Bispecific antibodies may also be used to localize cytotoxic agents to cells which express a particular polypeptide. These antibodies possess a polypeptide-binding arm and an arm which binds a cytotoxic agent or a radionuclide chelator, such as EÖTUBE, DPTA, DOTA, or TETA. Another bispecific antibody of interest binds the polypeptide and further binds tissue factor (TF).

6. Heteroconjugate Antibodies

Heteroconjugate antibodies are composed of two covalently joined antibodies. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells [U.S. Patent No. 4,676,980], and for treatment of HIV infection [WO 91/00360; WO 92/200373; EP 03089]. It is contemplated that the antibodies may be prepared *in vitro* using known methods in synthetic protein chemistry, including those involving crosslinking agents. For example, immunotoxins may be constructed using a disulfide exchange reaction or by forming a thioether bond. Examples of suitable reagents for this purpose include iminothiolate and methyl-4-mercaptobutyrimidate and those disclosed, for example, in U.S. Patent No. 4,676,980.

7. Effector function engineering

It may be desirable to modify the antibody of the invention with respect to effector function, so as to enhance the effectiveness of the antibody in treating cancer, for example. For example, cysteine residue(s) may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). See Caron *et al.*, J. Exp Med., 176:1191-1195 (1992) and Shopes, J. Immunol., 148:2918-2922 (1992). Homodimeric antibodies with enhanced anti-tumor activity may also be prepared using heterobifunctional cross-linkers as described in Wolff *et al.*, Cancer Research, 53:2560-2565 (1993). Alternatively, an antibody can be engineered which has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. See Stevenson *et al.*, Anti-Cancer Drug Design, 3:219-230 (1989).

8. Immunoconjugates

The invention also pertains to immunoconjugates comprising an antibody conjugated to a cytotoxic agent such as a chemotherapeutic agent, toxin (*e.g.*, an enzymatically active toxin of bacterial, fungal, plant or animal origin, or fragments thereof, or a small molecule toxin), or a radioactive isotope (*i.e.*, a radioconjugate).

Chemotherapeutic agents useful in the generation of such immunoconjugates have been described above. Enzymatically active protein toxins and fragments thereof which can be used include diphtheria A chain, nonbinding active fragments of diphtheria toxin, cholera toxin, botulinus toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, *Aleurites fordii* proteins, dianthin

proteins, *Phytolacca americana* proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcumin, crocin, saponaria officinalis inhibitor, gelonin, saporin, mitogellin, restrictocin, phenomycin, enomycin and the tricothecenes. Small molecule toxins include, for example, calicheamicins, maytansinoids, palytoxin and CC1065. A variety of radionuclides are available for the production of radioconjugated antibodies. Examples include ^{212}Bi , ^{131}I , ^{131}In , ^{90}Y and ^{186}Re .

Conjugates of the antibody and cytotoxic agent are made using a variety of bifunctional protein coupling agents such as N-succinimidyl-3-(2-pyridyldithiol) propionate (SPDP), iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutaraldehyde), bis-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), bis-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as tolyene 2,6-diisocyanate), and bis-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). For example, a ricin immunotoxin can be prepared as described in Vitetta *et al.*, Science, 238:1098 (1987). Carbon-14-labeled 1-isothiocyanatobenzyl-3-methyldiethylene triaminepentaacetic acid (MX-DTPA) is an exemplary chelating agent for conjugation of radionucleotide to the antibody. See WO94/11026.

In another embodiment, the antibody may be conjugated to a "receptor" (such streptavidin) for utilization in tumor pretargeting wherein the antibody-receptor conjugate is administered to the patient, followed by removal of unbound conjugate from the circulation using a clearing agent and then administration of a "ligand" (e.g., avidin) which is conjugated to a cytotoxic agent (e.g., a radionucleotide).

9. Immunoliposomes

The antibodies disclosed herein may also be formulated as immunoliposomes. Liposomes containing the antibody are prepared by methods known in the art, such as described in Epstein *et al.*, Proc. Natl. Acad. Sci. USA, 82:3688 (1985); Hwang *et al.*, Proc. Natl. Acad. Sci. USA, 77:4030 (1980); and U.S. Patent Nos. 4,485,045 and 4,544,545. Liposomes with enhanced circulation time are disclosed in U.S. Patent No. 5,013,556.

Particularly useful liposomes can be generated by the reverse phase evaporation method with a lipid composition comprising phosphatidylcholine, cholesterol and PEG-derivatized phosphatidylethanolamine (PEG-PE). Liposomes are extruded through filters of defined pore size to yield liposomes with the desired diameter. Fab' fragments of the antibody of the present invention can be conjugated to the liposomes as described in Martin *et al.*, J. Biol. Chem., 257:286-288 (1982) via a disulfide interchange reaction. A chemotherapeutic agent (such as Doxorubicin) is optionally contained within the liposome. See Gabizon *et al.*, J. National Cancer Inst., 81(19):1484 (1989).

N. Pharmaceutical Compositions

Antibodies specifically binding the product of an amplified gene identified herein, as well as other molecules identified by the screening assays disclosed hereinbefore, can be administered for the treatment of tumors, including cancers, in the form of pharmaceutical compositions.

If the protein encoded by the amplified gene is intracellular and whole antibodies are used as inhibitors, internalizing antibodies are preferred. However, lipofections or liposomes can also be used to deliver the antibody,

or an antibody fragment, into cells. Where antibody fragments are used, the smallest inhibitory fragment which specifically binds to the binding domain of the target protein is preferred. For example, based upon the variable region sequences of an antibody, peptide molecules can be designed which retain the ability to bind the target protein sequence. Such peptides can be synthesized chemically and/or produced by recombinant DNA technology (see, *e.g.*,
5 Marasco *et al.*, *Proc. Natl. Acad. Sci. USA*, 90:7889-7893 [1993]).

Therapeutic formulations of the antibody are prepared for storage by mixing the antibody having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers (Remington's Pharmaceutical Sciences, 16th edition, Osol, A. Ed. [1980]), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations
10 employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and *m*-cresol); low molecular weight (less than about
15 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (*e.g.*, Zn-protein complexes); and/or non-ionic surfactants such as TWEEN™, PLURONICS™ or polyethylene glycol (PEG).

20 Non-antibody compounds identified by the screening assays of the present invention can be formulated in an analogous manner, using standard techniques well known in the art.

The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. Alternatively, or in addition, the composition may comprise a cytotoxic agent, cytokine or growth inhibitory agent.
25 Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques
30 are disclosed in Remington's Pharmaceutical Sciences, 16th edition, Osol, A. Ed. (1980).

The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the
35 form of shaped articles, *e.g.*, films or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919), copolymers of L-glutamic acid and ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT™ (injectable microspheres composed of lactic

acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated antibodies remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37°C, resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies can be devised for stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

10

O. Methods of Treatment

It is contemplated that the antibodies and other anti-tumor compounds of the present invention may be used to treat various conditions, including those characterized by overexpression and/or activation of the amplified genes identified herein. Exemplary conditions or disorders to be treated with such antibodies and other compounds, including, but not limited to, small organic and inorganic molecules, peptides, antisense molecules, etc., include benign or malignant tumors (*e.g.*, renal, liver, kidney, bladder, breast, gastric, ovarian, colorectal, prostate, pancreatic, lung, vulval, thyroid, hepatic carcinomas; sarcomas; glioblastomas; and various head and neck tumors); leukemias and lymphoid malignancies; other disorders such as neuronal, glial, astrocytal, hypothalamic and other glandular, macrophagal, epithelial, stromal and blastocoelic disorders; and inflammatory, angiogenic and immunologic disorders.

The anti-tumor agents of the present invention, *e.g.*, antibodies, are administered to a mammal, preferably a human, in accord with known methods, such as intravenous administration as a bolus or by continuous infusion over a period of time, by intramuscular, intraperitoneal, intracerebrospinal, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. Intravenous administration of the antibody is preferred.

Other therapeutic regimens may be combined with the administration of the anti-cancer agents, *e.g.*, antibodies of the instant invention. For example, the patient to be treated with such anti-cancer agents may also receive radiation therapy. Alternatively, or in addition, a chemotherapeutic agent may be administered to the patient. Preparation and dosing schedules for such chemotherapeutic agents may be used according to manufacturers' instructions or as determined empirically by the skilled practitioner. Preparation and dosing schedules for such chemotherapy are also described in Chemotherapy Service Ed., M.C. Perry, Williams & Wilkins, Baltimore, MD (1992). The chemotherapeutic agent may precede, or follow administration of the anti-tumor agent, *e.g.*, antibody, or may be given simultaneously therewith. The antibody may be combined with an anti-oestrogen compound such as tamoxifen or an anti-progesterone such as onapristone (see, EP 616812) in dosages known for such molecules.

It may be desirable to also administer antibodies against other tumor associated antigens, such as antibodies which bind to the ErbB2, EGFR, ErbB3, ErbB4, or vascular endothelial factor (VEGF). Alternatively, or in addition, two or more antibodies binding the same or two or more different antigens disclosed herein may be co-administered to the patient. Sometimes, it may be beneficial to also administer one or more cytokines to the patient. In a preferred embodiment, the antibodies herein are co-administered with a growth inhibitory agent. For example,

the growth inhibitory agent may be administered first, followed by an antibody of the present invention. However, simultaneous administration or administration of the antibody of the present invention first is also contemplated. Suitable dosages for the growth inhibitory agent are those presently used and may be lowered due to the combined action (synergy) of the growth inhibitory agent and the antibody herein.

5 For the prevention or treatment of disease, the appropriate dosage of an anti-tumor agent, *e.g.*, an antibody herein will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether the agent is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the agent, and the discretion of the attending physician. The agent is suitably administered to the patient at one time or over a series of treatments.

10 For example, depending on the type and severity of the disease, about 1 $\mu\text{g}/\text{kg}$ to 15 mg/kg (*e.g.*, 0.1-20 mg/kg) of antibody is an initial candidate dosage for administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. A typical daily dosage might range from about 1 $\mu\text{g}/\text{kg}$ to 100 mg/kg or more, depending on the factors mentioned above. For repeated administrations over several days or longer, depending on the condition, the treatment is sustained until a desired suppression of disease symptoms
15 occurs. However, other dosage regimens may be useful. The progress of this therapy is easily monitored by conventional techniques and assays.

P. Articles of Manufacture

In another embodiment of the invention, an article of manufacture containing materials useful for the
20 diagnosis or treatment of the disorders described above is provided. The article of manufacture comprises a container and a label. Suitable containers include, for example, bottles, vials, syringes, and test tubes. The containers may be formed from a variety of materials such as glass or plastic. The container holds a composition which is effective for diagnosing or treating the condition and may have a sterile access port (for example the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection
25 needle). The active agent in the composition is usually an anti-tumor agent capable of interfering with the activity of a gene product identified herein, *e.g.*, an antibody. The label on, or associated with, the container indicates that the composition is used for diagnosing or treating the condition of choice. The article of manufacture may further comprise a second container comprising a pharmaceutically-acceptable buffer, such as phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other materials desirable from a commercial and user
30 standpoint, including other buffers, diluents, filters, needles, syringes, and package inserts with instructions for use.

Diagnosis and Prognosis of Tumors

While cell surface proteins, such as growth receptors overexpressed in certain tumors are excellent targets for drug candidates or tumor (*e.g.*, cancer) treatment, the same proteins along with secreted proteins encoded by the
35 genes amplified in tumor cells find additional use in the diagnosis and prognosis of tumors. For example, antibodies directed against the protein products of genes amplified in tumor cells can be used as tumor diagnostics or prognostics.

For example, antibodies, including antibody fragments, can be used to qualitatively or quantitatively detect

the expression of proteins encoded by the amplified genes ("marker gene products"). The antibody preferably is equipped with a detectable, *e.g.*, fluorescent label, and binding can be monitored by light microscopy, flow cytometry, fluorimetry, or other techniques known in the art. These techniques are particularly suitable, if the amplified gene encodes a cell surface protein, *e.g.*, a growth factor. Such binding assays are performed essentially
5 as described in section 5 above.

In situ detection of antibody binding to the marker gene products can be performed, for example, by immunofluorescence or immunoelectron microscopy. For this purpose, a histological specimen is removed from the patient, and a labeled antibody is applied to it, preferably by overlaying the antibody on a biological sample. This procedure also allows for determining the distribution of the marker gene product in the tissue examined. It
10 will be apparent for those skilled in the art that a wide variety of histological methods are readily available for *in situ* detection.

The following examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way.

All patent and literature references cited in the present specification are hereby incorporated by reference
15 in their entirety.

EXAMPLES

Commercially available reagents referred to in the examples were used according to manufacturer's instructions unless otherwise indicated. The source of those cells identified in the following examples, and
20 throughout the specification, by ATCC accession numbers is the American Type Culture Collection, 10801 University Blvd., Manassas, VA 20110-2209. All original deposits referred to in the present application were made under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and the Regulations thereunder (Budapest Treaty). This assures maintenance of a viable culture of the deposit for 30 years from the date of deposit. The deposit will be made available by ATCC
25 under the terms of the Budapest Treaty, and subject to an agreement between Genentech, Inc., and ATCC, which assures permanent and unrestricted availability of the progeny of the culture of the deposit to the public upon issuance of the pertinent U.S. patent or upon laying open to the public of any U.S. or foreign patent application, whichever comes first, and assures availability of the progeny to one determined by the U.S. Commissioner of Patents and Trademarks to be entitled thereto according to 35 USC § 122 and the Commissioner's rules pursuant
30 thereto (including 37 CFR § 1.14 with particular reference to 886 OG 638).

Unless otherwise noted, the present invention uses standard procedures of recombinant DNA technology, such as those described hereinabove and in the following textbooks: Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press N.Y., 1989; Ausubel *et al.*, Current Protocols in Molecular Biology, Green Publishing Associates and Wiley Interscience, N.Y., 1989; Innis *et al.*, PCR Protocols: A Guide to Methods
35 and Applications, Academic Press, Inc., N.Y., 1990; Harlow *et al.*, Antibodies: A Laboratory Manual, Cold Spring Harbor Press, Cold Spring Harbor, 1988; Gait, Oligonucleotide Synthesis, IRL Press, Oxford, 1984; R.I. Freshney, Animal Cell Culture, 1987; Coligan *et al.*, Current Protocols in Immunology, 1991.

EXAMPLE 1

Isolation of cDNA clones Encoding PRO212

The extracellular domain (ECD) sequences (including the secretion signal sequence, if any) from about 950 known secreted proteins from the Swiss-Prot public database were used to search EST databases. The EST
5 databases included public EST databases (e.g., GenBank), a proprietary EST database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA), and a proprietary EST database from Genentech. The search was performed using the computer program BLAST or BLAST2 [Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)] as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequences. Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered
10 and assembled into consensus DNA sequences with the program "phrap" (Phil Green, University of Washington, Seattle, Washington).

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described above. The ESTs included an EST proprietary to Genentech. In some cases, the consensus sequence derives from an intermediate consensus DNA sequence which was extended using repeated cycles of BLAST and phrap to extend
15 that intermediate consensus sequence as far as possible using the sources of EST sequences discussed above.

Based on the consensus sequence, oligonucleotides were synthesized : 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO212. Forward and reverse PCR primers generally range from 20 to 30 nucleotides and are often designed to give a PCR product of about 100-1000 bp in length. The probe sequences are typically 40-55 bp in length. In
20 some cases, additional oligonucleotides are synthesized when the consensus sequence is greater than about 1-1.5kbp. In order to screen several libraries for a full-length clone, DNA from the libraries was screened by PCR amplification, as per Ausubel *et al.*, Current Protocols in Molecular Biology, *supra*, with the PCR primer pair. A positive library was then used to isolate clones encoding the gene of interest using the probe oligonucleotide and one of the primer pairs.

25 PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-CACGCTGGTTTCTGCTTGGAG-3' (SEQ ID NO:3)

reverse PCR primer 5'-AGCTGGTGCACAGGGTGTCTATG-3' (SEQ ID NO:4)

Additionally, a synthetic oligonucleotide hybridization probe was constructed which had the following nucleotide sequence:

30 hybridization probe

5'-CCCAGGCACCTTCTCAGCCAGCCAGCAGCTCCAGCTCAGAGCAGTGCCAGCCC-3' (SEQ ID NO:5)

RNA for construction of the cDNA libraries was isolated from human fetal lung tissue. The cDNA libraries used to isolate the cDNA clones were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA. The cDNA was primed with oligo dT containing a NotI site, linked with
35 blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRKD; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes *et al.*, Science, 253:1278-1280 (1991)) in the unique XhoI and NotI sites.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for a full-

length PRO212 polypeptide (designated herein as DNA30942-1134 [Figure 1, SEQ ID NO:1]) and the derived protein sequence for that PRO212 polypeptide.

The full length clone identified above contained a single open reading frame with an apparent translational initiation site at nucleotide positions 101-103 and a stop signal at nucleotide positions 1001-1003 (Figure 1, SEQ ID NO:1). The predicted polypeptide precursor is 300 amino acids long. Analysis of the full length PRO212 sequence shown in Figure 2 (SEQ ID NO:2) evidences the presence of a variety of important polypeptide domains as shown in Figure 2, wherein the locations given for those important polypeptide domains are approximate as described above. The N-terminus of the PRO212 sequence contains a typical secretion signal (amino acids 1-23 of Figure 2; SEQ ID NO:2). It is believed that amino acids 1-215 of PRO212 (Figure 2; SEQ ID NO:2) represents an extracellular domain. PRO212 has one potential N-linked glycosylation site at amino acid residue 173 (Figure 2; SEQ ID NO:2). Clone DNA30942-1134 has been deposited with ATCC on September 16, 1997 and is assigned ATCC deposit no. 209254. PRO212 shows some amino acid sequence identity to TNFR2 (28.7%) and to OPG (31%).

EXAMPLE 2

Isolation of cDNA Clones Encoding PRO290

An expressed sequence tag (EST) DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) was searched and an EST was identified which showed homology to NTII-1 (a nerve protein which facilitates regeneration), FAN and beige.

RNA for construction of cDNA libraries was then isolated from human fetal kidney tissue. The cDNA libraries used to isolate the cDNA clones encoding human PRO290 were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA. The cDNA was primed with oligodT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRK5D; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes *et al.*, Science, 253:1278-1280 (1991)) in the unique XhoI and NotI.

The human fetal kidney cDNA libraries (prepared as described above), were screened by hybridization with a synthetic oligonucleotide probe:

5'-TGACTGCACTACCCCGTGGCAAGCTGTTGAGCCAGCTCAGCTG-3' (SEQ ID NO:8)

A cDNA clone was sequenced in entirety and is herein designated as DNA35680-1212. The entire nucleotide sequence of clone DNA35680-1212 is shown in Figures 3A-3B (SEQ ID NO:6). Clone DNA35680-1212 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 293-295 and a stop signal at nucleotide positions 3002-3004 (Figures 3A-3B, SEQ ID NO:6). The predicted polypeptide precursor is 1003 amino acids long, has a calculated molecular weight of approximately 112,013 daltons and an estimated pI of approximately 6.4. Analysis of the full-length PRO290 sequence shown in Figure 4 (SEQ ID NO:7) evidences sequence identity between the PRO290 amino acid sequence and FAN and beige. Due to this sequence identity, it is currently believed that the PRO290 polypeptide is related to FAN and/or beige. Clone DNA35680-1212 has been deposited with ATCC on April 21, 1998 and is assigned ATCC deposit no. 209790.

EXAMPLE 3

Isolation of cDNA Clones Encoding a Human PRO341

1. Preparation of oligo dT primed cDNA library

mRNA was isolated from human placenta tissue using reagents and protocols from Invitrogen, San Diego, CA (Fast Track 2). This RNA was used to generate an oligo dT primed cDNA library in the vector pRK5D using reagents and protocols from Life Technologies, Gaithersburg, MD (Super Script Plasmid System). In this procedure, the double stranded cDNA was sized to greater than 1000 bp and the SalI/NotI linker cDNA was cloned into XhoI/NotI cleaved vector. pRK5D is a cloning vector that has an sp6 transcription initiation site followed by an SfiI restriction enzyme site preceding the XhoI/NotI cDNA cloning sites.

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2. Preparation of random primed cDNA library

A secondary cDNA library was generated in order to preferentially represent the 5' ends of the primary cDNA clones. Sp6 RNA was generated from the primary library (described above), and this RNA was used to generate a random primed cDNA library in the vector pSST-AMY.0 using reagents and protocols from Life Technologies (Super Script Plasmid System, referenced above). In this procedure the double stranded cDNA was sized to 500-1000 bp, linker with blunt to NotI adaptors, cleaved with SfiI, and cloned into SfiI/NotI cleaved vector. pSST-AMY.0 is a cloning vector that has a yeast alcohol dehydrogenase promoter preceding the cDNA cloning sites and the mouse amylase sequence (the mature sequence without the secretion signal) followed by the yeast alcohol dehydrogenase terminator, after the cloning sites. Thus, cDNAs cloned into this vector that are fused in frame with the amylase sequence will lead to the secretion of amylase from appropriately transfected yeast colonies.

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3. Transformation and Detection

DNA from the library described in paragraph 2 above was chilled on ice to which was added electrocompetent DH10B bacteria (Life Technologies, 20 ml). The bacteria and vector mixture was then electroporated as recommended by the manufacturer. Subsequently, SOC media (Life Technologies, 1 ml) was added and the mixture was incubated at 37°C for 30 minutes. The transformants were then plated onto 20 standard 150 mm LB plates containing ampicillin and incubated for 16 hours (37°C). Positive colonies were scraped off the plates and the DNA was isolated from the bacterial pellet using standard protocols, e.g., CsCl-gradient. The purified DNA was then carried on to the yeast protocols below.

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The yeast methods were divided into three categories: (1) Transformation of yeast with the plasmid/cDNA combined vector; (2) Detection and isolation of yeast colonies secreting amylase; and (3) PCR amplification of the insert directly from the yeast colony and purification of the DNA for sequencing and further analysis.

The yeast strain used was HD56-5A (ATCC-90785). This strain has the following genotype: MAT alpha, ura3-52, leu2-3, leu2-112, his3-11, his3-15, MAL⁺, SUC⁺, GAL⁺. Preferably, yeast mutants can be employed that have deficient post-translational pathways. Such mutants may have translocation deficient alleles in *sec71*, *sec72*, *sec62*, with truncated *sec71* being the most preferred. Alternatively, antagonists (including antisense nucleotides and/or ligands) which interfere with the normal operation of these genes, other proteins implicated in this post

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translational pathway (e.g., SEC61p, SEC72p, SEC62p, SEC63p, TDJ1p or SSA1p-4p) or the complex formation of these proteins may also be preferably employed in combination with the amylase-expressing yeast.

Transformation was performed based on the protocol outlined by Gietz *et al.*, Nucl. Acid. Res., **20**:1425 (1992). Transformed cells were then inoculated from agar into YEPD complex media broth (100 ml) and grown overnight at 30°C. The YEPD broth was prepared as described by Kaiser *et al.*, Methods in Yeast Genetics, Cold Spring Harbor Press, Cold Spring Harbor, NY, p. 207 (1994). The overnight culture was then diluted to about 2×10^6 cells/ml (approx. OD₆₀₀=0.1) into fresh YEPD broth (500 ml) and regrown to 1×10^7 cells/ml (approx. OD₆₀₀=0.4-0.5).

The cells were then harvested and prepared for transformation by transfer into GS3 rotor bottles in a Sorval GS3 rotor at 5,000 rpm for 5 minutes, the supernatant discarded, and then resuspended into sterile water, and centrifuged again in 50 ml falcon tubes at 3,500 rpm in a Beckman GS-6KR centrifuge. The supernatant was discarded and the cells were subsequently washed with LiAc/TE (10 ml, 10 mM Tris-HCl, 1 mM EDTA pH 7.5, 100 mM Li₂OOCCH₃), and resuspended into LiAc/TE (2.5 ml).

Transformation took place by mixing the prepared cells (100 µl) with freshly denatured single stranded salmon testes DNA (Lofstrand Labs, Gaithersburg, MD) and transforming DNA (1 µg, vol. < 10 µl) in microfuge tubes. The mixture was mixed briefly by vortexing, then 40% PEG/TE (600 µl, 40% polyethylene glycol-4000, 10 mM Tris-HCl, 1 mM EDTA, 100 mM Li₂OOCCH₃, pH 7.5) was added. This mixture was gently mixed and incubated at 30°C while agitating for 30 minutes. The cells were then heat shocked at 42°C for 15 minutes, and the reaction vessel centrifuged in a microfuge at 12,000 rpm for 5-10 seconds, decanted and resuspended into TE (500 µl, 10 mM Tris-HCl, 1 mM EDTA pH 7.5) followed by recentrifugation. The cells were then diluted into TE (1 ml) and aliquots (200 µl) were spread onto the selective media previously prepared in 150 mm growth plates (VWR).

Alternatively, instead of multiple small reactions, the transformation was performed using a single, large scale reaction, wherein reagent amounts were scaled up accordingly.

The selective media used was a synthetic complete dextrose agar lacking uracil (SCD-Ura) prepared as described in Kaiser *et al.*, Methods in Yeast Genetics, Cold Spring Harbor Press, Cold Spring Harbor, NY, p. 208-210 (1994). Transformants were grown at 30°C for 2-3 days.

The detection of colonies secreting amylase was performed by including red starch in the selective growth media. Starch was coupled to the red dye (Reactive Red-120, Sigma) as per the procedure described by Biely *et al.*, Anal. Biochem., **172**:176-179 (1988). The coupled starch was incorporated into the SCD-Ura agar plates at a final concentration of 0.15% (w/v), and was buffered with potassium phosphate to a pH of 7.0 (50-100 mM final concentration).

The positive colonies were picked and streaked across fresh selective media (onto 150 mm plates) in order to obtain well isolated and identifiable single colonies. Well isolated single colonies positive for amylase secretion were detected by direct incorporation of red starch into buffered SCD-Ura agar. Positive colonies were determined by their ability to break down starch resulting in a clear halo around the positive colony visualized directly.

4. Isolation of DNA by PCR Amplification

When a positive colony was isolated, a portion of it was picked by a toothpick and diluted into sterile water

(30 μ l) in a 96 well plate. At this time, the positive colonies were either frozen and stored for subsequent analysis or immediately amplified. An aliquot of cells (5 μ l) was used as a template for the PCR reaction in a 25 μ l volume containing: 0.5 μ l KlenTaq (Clontech, Palo Alto, CA); 4.0 μ l 10 mM dNTP's (Perkin Elmer-Cetus); 2.5 μ l KlenTaq buffer (Clontech); 0.25 μ l forward oligo 1; 0.25 μ l reverse oligo 2; 12.5 μ l distilled water. The sequence of the forward oligonucleotide 1 was:

5'-TGTAACGACGGCCAGTTAAATAGACCTGCAATTATTAATCT-3' (SEQ ID NO:11)

The sequence of reverse oligonucleotide 2 was:

5'-CAGGAAACAGCTATGACCACCTGCACACCTGCAAATCCATT-3' (SEQ ID NO:12)

PCR was then performed as follows:

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|----|----|---------------|------------------|
| 10 | a. | Dentaure | 92°C, 5 minutes |
| | b. | 3 cycles of: | |
| | | Denature | 92°C, 30 seconds |
| | | Anneal | 59°C, 30 seconds |
| | | Extend | 72°C, 60 seconds |
| 15 | c. | 3 cycles of: | |
| | | Denature | 92°C, 30 seconds |
| | | Anneal | 57°C, 30 seconds |
| | | Extend | 72°C, 60 seconds |
| 20 | d. | 25 cycles of: | |
| | | Denature | 92°C, 30 seconds |
| | | Anneal | 55°C, 30 seconds |
| | | Extend | 72°C, 60 seconds |
| 25 | e. | Hold | 4°C |

The underlined regions of the oligonucleotides disclosed above annealed to the ADH promoter region and the amylase region, respectively, and amplified a 307 bp region from vector pSST-AMY.0 when no insert was present. Typically, the first 18 nucleotides of the 5' end of these oligonucleotides contained annealing sites for the sequencing primers. Thus, the total product of the PCR reaction from an empty vector was 343 bp. However, signal sequence-fused cDNA resulted in considerably longer nucleotide sequences.

Following the PCR, an aliquot of the reaction (5 μ l) was examined by agarose gel electrophoresis in a 1 % agarose gel using a Tris-Borate-EDTA (TBE) buffering system as described by Sambrook *et al.*, *supra*. Clones resulting in a single strong PCR product larger than 400 bp were further analyzed by DNA sequencing after purification with a 96 Qiaquick PCR clean-up column (Quiagen Inc., Chatsworth, CA).

5. Identification of Full-length Clone

A cDNA sequence isolated in the above screen is herein designated as DNA25314. Probes were then generated from the sequence of the DNA25314 molecule and used to screen a human placenta cDNA library prepared as described in paragraph 1 above. The cloning vector was pRK5B (pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes *et al.*, *Science*, 253:1278-1280 (1991)), and the cDNA size cut was less

than 2800 bp. The oligonucleotide probes were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding region of PRO341. Forward and reverse PCR primers generally range from 20-30 nucleotides and are often designed to give a PCR product of about 100-1000 bp in length. The probe sequences are typically 40-55 bp in length. In order to screen several libraries for a full-length clone, DNA from the libraries was screened by PCR amplification, as per Ausubel *et al.*, Current Protocols in Molecular Biology, *supra*, with the PCR primer pair. A positive library was then used to isolate clones encoding the gene of interest using oligonucleotide primers based upon the DNA25314 sequence.

A full length clone was identified that contained a single open reading frame with an apparent translational initiation site at nucleotide positions 380-382 and a stop signal at nucleotide positions 1754-1756 (Figure 5, SEQ ID NO:9). The predicted polypeptide precursor is 458 amino acids long, has a calculated molecular weight of approximately 50,264 daltons and an estimated pI of approximately 8.17. Analysis of the full length PRO341 sequence shown in Figure 6 (SEQ ID NO:10) evidences the presence of a variety of important polypeptide domains as shown in Figure 6, wherein the locations given for those polypeptide domains are approximate as described above. The full-length PRO341 sequence evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 17, transmembrane domains from about amino acid 171 to about amino acid 190, from about amino acid 220 to about amino acid 239, from about amino acid 259 to about amino acid 275, from about amino acid 286 to about amino acid 305, from about amino acid 316 to about amino acid 335, from about amino acid 353 to about amino acid 378 and from about amino acid 396 to about amino acid 417 and potential N-glycosylation sites from about amino acid 145 to about amino acid 147 and from about amino acid 155 to about amino acid 158. Clone DNA26288-1239 has been deposited with ATCC on April 21, 1998 and is assigned ATCC deposit no. 209792.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 6 (SEQ ID NO:10), evidenced sequence identity between the PRO341 amino acid sequence and the following Dayhoff sequences: S75696, H69788, D69852, A69888, B64918, F64752, LPU89276_1, G64962, S52977 and S44253.

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EXAMPLE 4

Isolation of cDNA Clones Encoding a Human PRO535 Polypeptide

DNA49143-1429 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence

from the Incyte database, designated as ss.clu12694.init. This EST cluster sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (*e.g.*, GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altschul *et al.*, Methods in
5 Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained is herein designated DNA48352. In light of an observed sequence homology between the DNA48352 sequence and Merck EST clone no. H86994, the Merck EST clone H86994 was purchased and the cDNA insert
10 was obtained and sequenced. It was found herein that the cDNA insert encoded a full-length protein. The sequence of this cDNA insert is shown in Figure 7 (SEQ ID NO: 13) and is herein designated as DNA49143-1429.

Clone DNA49143-1429 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 78-80 and ending at the stop codon at nucleotide positions 681-683 (Figure 7). The predicted polypeptide precursor is 201 amino acids long (Figure 8). The full-length PRO535 protein shown in Figure 8 has
15 an estimated molecular weight of about 22,180 daltons and a pI of about 9.68. Analysis of the full-length PRO535 sequence shown in Figure 8 (SEQ ID NO:14) evidences the presence of a variety of important polypeptide domains as shown in Figure 8, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO535 sequence evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 25, a transmembrane domain from about amino acid 155 to
20 about amino acid 174, a potential N-glycosylation site from about amino acid 196 to about amino acid 199 and FKBP-type peptidyl-prolyl cis-trans isomer signature sequences from about amino acid 62 to about amino acid 77, from about amino acid 87 to about amino acid 123 and from about amino acid 128 to about amino acid 141. Clone DNA49143-1429 has been deposited with ATCC on June 23, 1998 and is assigned ATCC deposit no. 203013.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment
25 analysis of the full-length sequence shown in Figure 8 (SEQ ID NO:14), evidenced sequence identity between the PRO535 amino acid sequence and the following Dayhoff sequences: S71237, P_R93551, P_R28980, S71238, FKB2_HUMAN, CELC05C8_1, S55383, S72485, CELC50F2_6 and S75144.

EXAMPLE 5

30 Isolation of cDNA Clones Encoding PRO619

DNA49821-1562 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides
35 surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence

contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence from the Incyte database, designated as 88434 (also called DNA48290). This EST cluster sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). In light of the information and discoveries provided herein, the clone including EST 1656694 was purchased from Incyte (from a library constructed from ureter tumor tissue, URETTUT01) and the cDNA insert was obtained and sequenced. The sequence of this cDNA insert is shown in Figure 9 (SEQ ID NO: 15) and is herein designated as DNA49821-1562.

Clone DNA49821-1562 shown in Figure 9 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 81-83 and ending at the stop codon at nucleotide positions 450-452 (Figure 9; SEQ ID NO:15). The predicted polypeptide precursor is 123 amino acids long including a predicted signal peptide at about amino acids 1-20 (Figure 10). The full-length PRO619 protein shown in Figure 10 (SEQ ID NO:16) has an estimated molecular weight of about 13,710 daltons and an estimated pI of about 5.19. Clone DNA49821-1562 has been deposited with ATCC on June 16, 1998 and is assigned ATCC deposit no. 209981.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 10 (SEQ ID NO:16), evidenced sequence identity between the PRO619 amino acid sequence and the following Dayhoff sequences: S35302, D87009_1, HSU93494_1, HUMIGLAM5_1, D86999_2, HUMIGLYM1_1, HUMIGLYMKE_1, A29491_1, A29498_1, and VPR2_MOUSE.

EXAMPLE 6

Isolation of cDNA Clones Encoding Human PRO717

The extracellular domain (ECD) sequences (including the secretion signal sequence, if any) from about 950 known secreted proteins from the Swiss-Prot public database were used to search EST databases. The EST databases included public EST databases (e.g., GenBank) and a proprietary EST database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA). The search was performed using the computer program BLAST or BLAST2 [Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)] as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequences. Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into consensus DNA sequences with the program "phrap" (Phil Green, University of Washington, Seattle, Washington).

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described above and is herein designated DNA42829. In some cases, the DNA42829 consensus sequence derives from an intermediate consensus DNA sequence which was extended using repeated cycles of BLAST and phrap to extend

that intermediate consensus sequence as far as possible using the sources of EST sequences discussed above.

Based on the DNA42829 consensus sequence, oligonucleotides were synthesized : 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO717. Forward and reverse PCR primers generally range from 20 to 30 nucleotides and are often designed to give a PCR product of about 100-1000 bp in length. The probe sequences are typically 40-55 bp in length. In some cases, additional oligonucleotides are synthesized when the consensus sequence is greater than about 1-1.5kbp. In order to screen several libraries for a full-length clone, DNA from the libraries was screened by PCR amplification, as per Ausubel *et al.*, Current Protocols in Molecular Biology, *supra*, with the PCR primer pair. A positive library was then used to isolate clones encoding the gene of interest using the probe oligonucleotide and one of the primer pairs.

PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-AGCTTCTCAGCCCTCCTGGAGCAG-3' (SEQ ID NO:19)

reverse PCR primer 5'-CGGGTCAATAAACCTGGACGCTTGG-3' (SEQ ID NO:20)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA42829 sequence which had the following nucleotide sequence:

hybridization probe

5'-TATGTGGACCGGACCAAGCACTTCACTGAGGCCACCAAGATTG-3' (SEQ ID NO:21)

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue (LIB229). The cDNA libraries used to isolate the cDNA clones were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA. The cDNA was primed with oligo dT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRKD; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes *et al.*, Science, 253:1278-1280 (1991)) in the unique XhoI and NotI sites.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for a full-length PRO717 polypeptide (designated herein as DNA50988-1326 [Figure 11, SEQ ID NO:17]) and the derived protein sequence for that PRO717 polypeptide.

The full length clone identified above contained a single open reading frame with an apparent translational initiation site at nucleotide positions 17-19 and a stop signal at nucleotide positions 1697-1699 (Figure 11, SEQ ID NO:17). The predicted polypeptide precursor is 560 amino acids long, has a calculated molecular weight of approximately 58,427 daltons and an estimated pI of approximately 6.86. Analysis of the full length PRO717 sequence shown in Figure 12 (SEQ ID NO:18) suggests that PRO717 may be a novel 12 transmembrane receptor. The reverse complement strand of DNA50988 has a stretch that matches identically with human regulatory myosin light strand. Further analysis of the full-length PRO717 sequence shown in Figure 12 (SEQ ID NO:18) evidences the presence of a variety of important polypeptide domains as shown in Figure 12, wherein the locations given for those important polypeptide domains are approximate as described above. Transmembrane domains are from about amino acid 30 to about amino acid 50, from about amino acid 61 to about amino acid 79, from about amino acid 98 to about amino acid 112, from about amino acid 126 to about amino acid 146, from about amino acid 169 to about

amino acid 182, from about amino acid 201 to about amino acid 215, from about amino acid 248 to about amino acid 268, from about amino acid 280 to about amino acid 300, from about amino acid 318 to about amino acid 337, from about amino acid 341 to about amino acid 357, from about amino acid 375 to about amino acid 387, and from amino acid 420 to about amino acid 441 (Figure 12; SEQ ID NO:18). N-glycosylation sites are from about amino acid 40 to about amino acid 43 and from about amino acid 43 to about amino acid 46 (Figure 12; SEQ ID NO:18). A glycosaminoglycan attachment site is from about amino acid 468 to about amino acid 471 (Figure 12; SEQ ID NO:18). Clone DNA50988-1326 has been deposited with the ATCC on April 28, 1998 and is assigned ATCC deposit no. 209814.

EXAMPLE 7

Isolation of cDNA Clones Encoding PRO809

DNA57836-1338 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST sequence from the Incyte database. This EST sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (*e.g.*, GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST-2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained is herein designated DNA56418. Based on the discoveries and information provided herein, Merck clone 230132 including EST H74302 (from library 305 a human fetal liver spleen library) was further examined. DNA sequencing gave the full-length DNA sequence for PRO809 shown in Figure 13 (SEQ ID NO:22) (herein designated as DNA57836-1338) and the derived protein sequence for PRO809.

Clone DNA57836-1338 (Figure 13; SEQ ID NO:22) contains a single open reading frame with an apparent translational initiation site at nucleotide positions 63-65 and ending at the stop codon at nucleotide positions 858-860 (Figure 13; SEQ ID NO:22). The predicted polypeptide precursor is 265 amino acids long (Figure 14). The full-length PRO809 protein shown in Figure 14 (SEQ ID NO:23) has an estimated molecular weight of about 29,061 daltons and a pI of about 9.18. Analysis of the full-length PRO809 sequence shown in Figure 14 (SEQ ID NO:23)

evidences the presence of important polypeptide domains as shown in Figure 14, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO809 sequence shown in Figure 14 evidences the following: a signal peptide from about amino acid 1 to about amino acid 18 and a potential N-glycosylation sites from about amino acid 86 to about amino acid 89, from about amino acid 132 to about amino acid 135 and from amino acid 181 to about amino acid 184. Clone DNA57836-1338 has been deposited with ATCC on June 23, 1998 and is assigned ATCC deposit no. 203025.

Analysis of the amino acid sequence of the full-length PRO809 polypeptide suggests that it possesses some sequence similarity to the heparin sulfate proteoglycan and to endothelial cell adhesion molecule-1. More specifically, an analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 14 (SEQ ID NO:23), evidenced sequence identity between the PRO809 amino acid sequence and the following Dayhoff sequences: PGBM_MOUSE, D82082_1 and PW14158.

EXAMPLE 8

Isolation of cDNA Clones Encoding PRO830

DNA56866-1342 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence from the Incyte database, designated 20251. This EST cluster sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (*e.g.*, GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST-2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained is herein designated DNA55733. In light of an observed sequence homology between DNA55733 sequence and Merck EST clone no. H78534, from the Merck database, the Merck EST clone no. H78534 was purchased and the cDNA insert was obtained and sequenced. It was found herein that the cDNA insert encoded a full-length protein. The sequence of this cDNA insert is shown in Figure 15 and is herein designated as DNA56866-1342.

Clone DNA56866-1342 (Figure 15; SEQ ID NO:24) contains a single open reading frame with an apparent

translational initiation site at nucleotide positions 154-156 and ending at the stop codon at nucleotide positions 415-417 (Figure 15; SEQ ID NO:24). The predicted polypeptide precursor is 87 amino acids long (Figure 16). The full-length PRO830 protein shown in Figure 16 (SEQ ID NO:25) has an estimated molecular weight of about 9,272 daltons and a pI of about 9.19. Analysis of the full-length PRO830 sequence shown in Figure 16 (SEQ ID NO:25) evidences the presence of important polypeptide domains as shown in Figure 16, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO830 sequence shown in Figure 16 evidences the following: a signal peptide from about amino acid 1 to about amino acid 33, potential N-myristoylation sites from about amino acid 2 to about amino acid 7 and from about amino acid 8 to about amino acid 13 and a thioredoxin family of proteins homology block from about amino acid 23 to about amino acid 39. Clone DNA56866-1342 has been deposited with ATCC on June 23, 1998 and is assigned ATCC deposit no. 203023.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 16 (SEQ ID NO:25), evidenced sequence identity between the PRO830 amino acid sequence and the following Dayhoff sequences: HSU88154_1, HSU88153_1, SAPKSGENE_1, HPU31791_5, GGCNOT2_1, CPU91421_1, CHKESTPC09_1, PQ0769, U97553_79 and B60095.

EXAMPLE 9

Isolation of cDNA Clones Encoding Human PRO848

DNA59839-1461 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc. (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Candidate sequences, also called initial consensus sequences, with a sufficient score were further examined with the computer program BLAST or BLAST-2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). In this case, an initial consensus sequence formed from an Incyte database of sequences identified from the signal algorithm is designated herein as DNA55999.init. This sequence was extended using repeated cycles of BLAST and "phrap" to extend the sequence as far as possible using public and Incyte EST sequences to form a consensus sequence herein designated DNA55999. Based on the discoveries and information provided herein, the clone including Incyte EST 2768571 was further examined. DNA sequencing gave the sequence herein designated as DNA59839-1461.

Clone DNA59839-1461 (Figure 17; SEQ ID NO:26) contains a single open reading frame with an apparent translational initiation site at nucleotide positions 146-148 and ending at the stop codon at nucleotide positions 1946-1948 (Figure 17; SEQ ID NO:26). The predicted polypeptide precursor is 600 amino acids long (Figure 18). The full-length PRO848 protein shown in Figure 18 (SEQ ID NO:27) has an estimated molecular weight of about 68,536 daltons and a pI of about 10.20. Analysis of the full-length PRO848 sequence shown in Figure 18 (SEQ ID NO:27) evidences the presence of important polypeptide domains as shown in Figure 18, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO848 sequence shown in Figure 18 evidences the following: a cytoplasmic domain from about amino acid 1 to about amino acid 10, a Type II transmembrane domain from about amino acid 11 to about amino acid 35, a luminal catalytic domain from about amino acid 36 to about amino acid 600, a ribonucleotide reductase small subunit signature from about amino acid 481 to about amino acid 496 and N-glycosylation sites from about amino acid 300 to about amino acid 303, from about amino acid 311 to about amino acid 314, from about amino acid 331 to about amino acid 334, from about amino acid 375 to about amino acid 378 and from about amino acid 460 to about amino acid 463 (Figure 18; SEQ ID NO:27). Clone DNA59839-1461 has been deposited with ATCC on June 16, 1998 and is assigned ATCC deposit no. 209988.

An analysis of the amino acid sequence of the full-length PRO848 polypeptide suggests that it may be a novel sialyltransferase. More specifically, an analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 18 (SEQ ID NO:27), evidenced sequence identity between the PRO848 amino acid sequence and the following Dayhoff sequences: P_R78619 (GalNAc-alpha-2, 6-sialyltransferase), CAAG5_CHICK (alpha-N-acetylgalactosamide alpha-2, 6-sialyltransferase), HSU14550_1, CAG6-HUMAN and P_R63217 (human alpha-2, 3-sialyltransferase).

EXAMPLE 10

Isolation of cDNA Clones Encoding Human PRO943

The extracellular domain (ECD) sequences (including the secretion signal sequence, if any) from about 950 known secreted proteins from the Swiss-Prot public database were used to search EST databases. The EST databases included public EST databases (*e.g.*, GenBank) and a proprietary EST database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA). The search was performed using the computer program BLAST or BLAST2 [Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)] as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequences. Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into consensus DNA sequences with the program "phrap" (Phil Green, University of Washington, Seattle, Washington).

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described above and is herein designated DNA36360. In some cases, the DNA36360 consensus sequence derives from an intermediate consensus DNA sequence which was extended using repeated cycles of BLAST and phrap to extend that intermediate consensus sequence as far as possible using the sources of EST sequences discussed above.

Based on the DNA36360 consensus sequence, oligonucleotides were synthesized : 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length

coding sequence for PRO943. Forward and reverse PCR primers generally range from 20 to 30 nucleotides and are often designed to give a PCR product of about 100-1000 bp in length. The probe sequences are typically 40-55 bp in length. In some cases, additional oligonucleotides are synthesized when the consensus sequence is greater than about 1-1.5kbp. In order to screen several libraries for a full-length clone, DNA from the libraries was screened by
5 PCR amplification, as per Ausubel *et al.*, Current Protocols in Molecular Biology, *supra*, with the PCR primer pair. A positive library was then used to isolate clones encoding the gene of interest using the probe oligonucleotide and one of the primer pairs.

PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-CGAGATGACGCCGAGCCCCC-3' (SEQ ID NO:30)

10 reverse PCR primer 5'-CGGTTCGACACGCGGCAGGTG-3' (SEQ ID NO:31)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA36360 sequence which had the following nucleotide sequence:

hybridization probe

5'-TGCTGCTCCTGCTGCCGCCGCTGCTGCTGGGGGCTTCCCGCCGG-3' (SEQ ID NO:32)

15 RNA for construction of the cDNA libraries was isolated from human fetal brain tissue. The cDNA libraries used to isolate the cDNA clones were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA. The cDNA was primed with oligo dT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRKD; pRK5B is a precursor of
20 pRK5D that does not contain the SfiI site; see, Holmes *et al.*, Science, 253:1278-1280 (1991)) in the unique XhoI and NotI sites.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for a full-length PRO943 polypeptide (designated herein as DNA52192-1369 [Figures 19A-19B, SEQ ID NO:28]) and the derived protein sequence for that PRO943 polypeptide.

25 The full length clone identified above contained a single open reading frame with an apparent translational initiation site at nucleotide positions 150-152 and a stop signal at nucleotide positions 1662-1664 (Figures 19A-19B, SEQ ID NO:28). The predicted polypeptide precursor is 504 amino acids long (Figure 20), has a calculated molecular weight of approximately 54,537 daltons and an estimated pI of approximately 10.04. Analysis of the full length PRO943 sequence shown in Figure 20 (SEQ ID NO:29) evidences the presence of a variety of important
30 polypeptide domains as shown in Figure 20, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO943 sequence shown in Figure 20 evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 17, a transmembrane domain from about amino acid 376 to about amino acid 396, tyrosine kinase phosphorylation sites from about amino acid 212 to about amino acid 219 and from about amino acid 329 to about amino acid 336, potential N-glycosylation
35 sites from about amino acid 111 to about amino acid 114, from about amino acid 231 to about amino acid 234, from about amino acid 255 to about amino acid 258 and from about amino acid 293 to about amino acid 296 and an immunoglobulin and MHC protein sequence homology block from about amino acid 219 to about amino acid 236. Clone DNA52192-1369 has been deposited with the ATCC on July 1, 1998 and is assigned ATCC deposit no.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 20 (SEQ ID NO:29) evidenced sequence identity between the PRO943 amino acid sequence and the following Dayhoff sequences: B49151, A39752, FGR1_XENLA, S38579, 5 RATHBFGFRB_1, TVHU2F, FGR2_MOUSE, CEK3_CHICK, P_R21080 and A27171_1.

EXAMPLE 11

Isolation of cDNA Clones Encoding Human PRO1005

DNA57708-1411 was identified by applying a proprietary signal sequence finding algorithm developed by 10 Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence 15 acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster from the 20 Incyte database, designated herein as Incyte cluster sequence no. 49243. This EST cluster sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (*e.g.*, GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altschul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 25 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA56380.

In light of an observed sequence homology between the DNA56380 sequence and Merck EST no. AA256657, from the Merck database, the Merck EST no. AA256657 was purchased and the cDNA insert was 30 obtained and sequenced. It was found herein that the cDNA insert encoded a full-length protein. The sequence of this cDNA insert is shown in Figure 21 (SEQ ID NO:33) and is herein designated as DNA57708-1411.

Clone DNA57708-1411 (Figure 21; SEQ ID NO:33) contains a single open reading frame with an apparent translational initiation site at nucleotide positions 30-32 and ending at the stop codon at nucleotide positions 585-587 (Figure 21; SEQ ID NO:33). The predicted polypeptide precursor is 185 amino acids long (Figure 22). The full- 35 length PRO1005 protein shown in Figure 22 (SEQ ID NO:34) has an estimated molecular weight of about 20,331 daltons and a pI of about 5.85. Analysis of the full-length PRO1005 sequence shown in Figure 22 (SEQ ID NO:34) evidences the presence of important polypeptide domains as shown in Figure 22, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1005

sequence shown in Figure 22 evidences the following: a signal peptide from about amino acid 1 to about amino acid 20, an N-myristoylation sites from about amino acid 67 to about amino acid 72, from about amino acid 118 to about amino acid 123 and from about amino acid 163 to about amino acid 168, and a flavodoxin protein homology from about amino acid 156 to about amino acid 174 (Figure 22; SEQ ID NO:34). Clone DNA57708-1411 has been deposited with ATCC on June 23, 1998 and is assigned ATCC deposit no. 203021.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 22 (SEQ ID NO:34), evidenced sequence identity between the PRO1005 amino acid sequence and the following Dayhoff sequences: DDU07187_1, DDU87912_1, CELD1007_14, A42239, DDU42597_1, CYAG_DICDI, S50452, MRKC_KLEPN, P-R41998, and XYNA_RUMFL.

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EXAMPLE 12

Isolation of cDNA Clones Encoding Human PRO1009

A cDNA clone 57129-1413 encoding a native human PRO1009 polypeptide was identified using a yeast screen, in a human SK-Lu-1 adenocarcinoma cell line cDNA library that preferentially represents the 5' ends of the primary cDNA clones.

Clone DNA57129-1413 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 41-43 and ending at the stop codon at nucleotide positions 1886-1888 (Figure 23; SEQ ID NO:35). The predicted polypeptide precursor is 615 amino acids long (Figure 24). The full-length PRO1009 protein shown in Figure 24 has an estimated molecular weight of about 68,125 daltons and a pI of about 7.82. Analysis of the full-length PRO1009 sequence shown in Figure 24 (SEQ ID NO:36) evidences the presence of a variety of important polypeptide domains as shown in Figure 24, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1009 sequence shown in Figure 24 evidences the following: a signal peptide from about amino acid 1 to about amino acid 22, transmembrane domains from about amino acid 140 to about amino acid 161, from about amino acid 213 to about amino acid 229 and from about amino acid 312 to about amino acid 334, a putative AMP-binding domain signature from about amino acid 260 to about amino acid 271, N-myristoylation sites from about amino acid 19 to about amino acid 24, from about amino acid 22 to about amino acid 27, from about amino acid 120 to about amino acid 125, from about amino acid 203 to about amino acid 208, from about amino acid 268 to about amino acid 273, from about amino acid 272 to about amino acid 277, from about amino acid 314 to about amino acid 319, from about amino acid 318 to about amino acid 323, from about amino acid 379 to about amino acid 384, from about amino acid 380 to about amino acid 385 and from about amino acid 409 to about amino acid 414, and an N-glycosylation site from about amino acid 282 to about amino acid 285. Clone DNA57129-1413 has been deposited with ATCC on June 16, 1998 and is assigned ATCC deposit no. 209977.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 24 (SEQ ID NO:36), evidenced sequence identity between the PRO1009 amino acid sequence and the following Dayhoff sequences: F69893, CEF28F8_2, BSY13917_7, D69187, D69649, XCRPFB_1, E64928, YDID_ECOLI, BNACSF8_1 and RPU75363_2.

EXAMPLE 13

Isolation of cDNA Clones Encoding Human PRO1025

DNA59622-1334 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither
10 meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence from the Incyte database from which a candidate sequence was formed designated herein as DNA56507.init. This
15 EST cluster sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (*e.g.*, GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altschul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and
20 assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA56507.

In light of an observed sequence homology between DNA56507 and Incyte EST 1324053 (from Incyte library LPARNOT 02 formed from cancerous human parotid gland tissue), Incyte EST 1324053 was further examined. DNA sequencing gave the full-length DNA sequence shown in Figure 25 (SEQ ID NO:37) and is herein
25 designated as DNA59622-1334.

Clone DNA59622-1334 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 67-69 and ending at the stop codon at nucleotide positions 814-816 (Figure 25; SEQ ID NO:37). The predicted polypeptide precursor is 249 amino acids long (Figure 26). The full-length PRO1025 protein shown in Figure 26 (SEQ ID NO:38) has an estimated molecular weight of about 27,011 daltons and a pI of about
30 5.48. Analysis of the full-length PRO1025 sequence shown in Figure 26 (SEQ ID NO:38) evidences the presence of important polypeptide domains as shown in Figure 26, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1025 sequence shown in Figure 26 evidences the following: a signal peptide from about amino acid 1 to about amino acid 14, N-glycosylation sites from about amino acid 124 to about amino acid 127 and from about amino acid 132 to about amino acid 135, and a
35 leucine zipper pattern from about amino acid 44 to about amino acid 65. Clone DNA59622-1334 has been deposited with ATCC on June 16, 1998 and is assigned ATCC deposit no. 209984.

An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 26 (SEQ ID NO:38), evidenced sequence identity between the

PRO1025 amino acid sequence and the following Dayhoff sequences: BTU79413_1, MMU1_1, A42337, MMU46068_1 and S68553.

EXAMPLE 14

5 Isolation of cDNA Clones Encoding Human PRO1030

DNA59485-1336 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (*e.g.*, GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides
10 surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG
15 codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm applied to Incyte fragments from the Incyte database, sequences fitting the algorithm were identified. From those sequences fitting the algorithm, a sequence was derived. This derived sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (*e.g.*, GenBank, Merck/Washington University) and a proprietary EST DNA database
20 (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein
25 designated DNA56450.

Based upon the assembly and other discoveries and information provided herein, a clone containing Merck EST AA625485 was further examined. DNA sequencing gave the full-length DNA sequence shown in Figure 27 (SEQ ID NO:39) and is herein designated as DNA59485-1336.

Clone DNA59485-1336 contains a single open reading frame with an apparent translational initiation site
30 at nucleotide positions 148-150 and ending at the stop codon at nucleotide positions 673-675 (Figure 27; SEQ ID NO:39). The predicted polypeptide precursor is 175 amino acids long (Figure 28). The full-length PRO1030 protein shown in Figure 28 (SEQ ID NO:40) has an estimated molecular weight of about 19,979 daltons and a pI of about 9.26. Analysis of the full-length PRO1030 sequence shown in Figure 28 (SEQ ID NO:40) evidences the presence of an important polypeptide domain as shown in Figure 28, wherein the location given for that important polypeptide
35 domain is approximate as described above. Analysis of the full-length PRO1030 sequence shown in Figure 28 evidences the presence of a signal peptide from about amino acid 1 to about amino acid 20. Clone DNA59485-1336 has been deposited with ATCC on June 23, 1998 and is assigned ATCC deposit no. 203015.

EXAMPLE 15

Isolation of cDNA Clones Encoding Human PRO1097

DNA59841-1460 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (e.g., GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Using of the above described signal sequence algorithm applied to Incyte fragments from the Incyte database, sequences fitting the algorithm were identified. From those sequences fitting the algorithm, a sequence was derived. This derived sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA56006.

Based on the information discovered and provided herein, the Incyte clone including EST 2408105, from Incyte brain stem library 311 "BSTMNON02" was further examined. DNA sequencing gave the full-length DNA sequence shown in Figure 29 (SEQ ID NO:41) and is herein designated as DNA59841-1460.

Clone DNA59841-1460 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 3-5 and ending at the stop codon at nucleotide positions 276-278 (Figure 29; SEQ ID NO:41). The predicted polypeptide precursor is 91 amino acids long (Figure 30). The full-length PRO1097 protein shown in Figure 30 (SEQ ID NO:42) has an estimated molecular weight of about 10,542 daltons and a pI of about 10.04. Analysis of the full-length PRO1097 sequence shown in Figure 30 (SEQ ID NO:42) evidences the presence of important polypeptide domains as shown in Figure 30, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1097 sequence shown in Figure 30 evidences the following: a signal peptide from about amino acid 1 to about amino acid 20, glycoprotease family protein domain starting from about amino acid 56, and an acyltransferase ChoActase/COT/CPT family peptide starting from about amino acid 49 (Figure 30; SEQ ID NO:42). Clone DNA59841-1460 has been deposited with ATCC on July 1, 1998 and is assigned ATCC deposit no. 203044.

EXAMPLE 16

Isolation of cDNA Clones Encoding Human PRO1107

DNA59606-1471 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (e.g., GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence from the Incyte database, designated herein as DNA56402.init. This EST cluster sequence was then compared to a variety of expressed sequence tag (EST) databases which included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altschul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA56402.

In light of an observed sequence homology between the DNA56402 and an Incyte EST no. 3203694, (from Incyte library PENCNOT02, 309, a caversnosum tissue library from the same donor), Incyte EST no. 3203694 was further examined. DNA sequencing gave the full-length DNA sequence and is herein designated as DNA59606-1471 shown in Figure 31 (SEQ ID NO:43).

Clone DNA59606-1471 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 244-246 and ending at the stop codon at nucleotide positions 1675-1677 (Figure 31; SEQ ID NO:43). The predicted polypeptide precursor is 477 amino acids long (Figure 32). The full-length PRO1107 protein shown in Figure 32 (SEQ ID NO:44) has an estimated molecular weight of about 54,668 daltons and a pI of about 6.33. Analysis of the full-length PRO1107 sequence shown in Figure 32 (SEQ ID NO:44) evidences the presence of important polypeptide domains as shown in Figure 32, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1107 sequence shown in Figure 32 evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 22, a transmembrane domain from about amino acid 429 to about amino acid 452, N-glycosylation sites from about amino acid 101 to about amino acid 104, from about amino acid 158 to about amino acid 161, from about amino acid 292 to about amino acid 295, from about amino acid 329 to about amino acid 332, from about amino acid 362 to about amino acid 365, from about amino acid 369 to about amino acid 372, from about amino acid 382 to about amino acid 385 and from about amino acid 389 to about amino acid 392, a somatomedin B domain protein from

about amino acid 69 to about amino acid 85 and a sulfatase protein region from about amino acid 212 to about amino acid 241. Clone DNA59606-1471 has been deposited with ATCC on June 9, 1998 and is assigned ATCC deposit no. 209945.

Analysis of the amino acid sequence of the full-length PRO1107 polypeptide (Figure 32) suggests that it possesses significant sequence similarity to phosphodiesterase I/nucleotide pyrophosphatase, human insulin receptor tyrosine kinase inhibitor, alkaline phosphodiesterase and autotaxin, thereby indicating that PRO1107 may be a novel phosphodiesterase. More specifically, an analysis of the Dayhoff database (version 35.45 SwissProt), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 32 (SEQ ID NO:44), evidenced sequence identity between the PRO1107 amino acid sequence and the following Dayhoff sequences: AF005632_1, P_R79148, RNU78787_1, AF060218_4, A57080 and HUMATXT_1.

EXAMPLE 17

Isolation of cDNA Clones Encoding a Human PRO1111

An expressed sequence tag (EST) DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) was searched and an EST was identified which showed homology to insulin-like growth factor binding protein.

RNA for construction of cDNA libraries was then isolated from human fetal brain tissue. The cDNA libraries used to isolate the cDNA clones encoding human PRO1111 were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA. The cDNA was primed with oligodT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRK5D; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes *et al.*, Science, 253:1278-1280 (1991)) in the unique XhoI and NotI.

The human fetal brain cDNA libraries (prepared as described above), were screened by hybridization with a synthetic oligonucleotide probe:

5'-CCACCACCTGGAGGTCCTGCAGTTGGGCAGGAAGTCCATCCGGCAGATTG-3'
(SEQ ID NO:47) based on the identified EST described above.

A cDNA clone was sequenced in entirety and is herein designated as DNA58721-1475. The entire nucleotide sequence of clone DNA58721-1475 is shown in Figure 33 (SEQ ID NO:45). Clone DNA58721-1475 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 57-59 and a stop signal at nucleotide positions 2016-2018 (Figure 33, SEQ ID NO:45). The predicted polypeptide precursor is 653 amino acids long, has a calculated molecular weight of approximately 72,717 daltons and an estimated pI of approximately 6.99. Analysis of the full-length PRO1111 sequence shown in Figure 34 (SEQ ID NO:46) evidences the presence of a variety of important polypeptide domains as shown in Figure 34, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1111 sequence evidences the presence of the following: transmembrane domains from about amino acid 21 to about amino acid 40 and from about amino acid 528 to about amino acid 548, N-glycosylation sites from about amino acid 277 to about amino acid 280, from about amino acid 322 to about amino acid 325, from about amino acid 363 to about amino acid 366, from about amino acid 388 to about amino acid 391, from about amino acid 410 to about amino

acid 413, from about amino acid 434 to about amino acid 437, from about amino acid 440 to about amino acid 443, from about amino acid 447 to about amino acid 450 and from about amino acid 450 to about amino acid 453, a glycosaminoglycan attachment site from about amino acid 591 to about amino acid 594, and a leucine zipper pattern from about amino acid 271 to about amino acid 292. Clone DNA58721-1475 has been deposited with ATCC on August 11, 1998 and is assigned ATCC deposit no. 203110.

An analysis of the Dayhoff database (version 35.45 SwissProt), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 34 (SEQ ID NO:46) evidenced sequence identity between the PRO1111 amino acid sequence and the following Dayhoff sequences: A58532, D86983_1, RNPLGPV_1, PGS2_HUMAN, AF038127_1, ALS_MOUSE, GPV_HUMAN, PGS2_BOVIN, ALS_PAPPA and I47020.

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EXAMPLE 18

Isolation of cDNA Clones Encoding a Human PRO1153

DNA59842-1502 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (e.g., GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence from the Incyte database. This EST cluster sequence was then compared to variety of expressed sequence tag (EST) databases which included public EST databases (e.g., GenBank, Merck/Washington University) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altschul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA56008.

In light of an observed sequence homology between the DNA56008 sequence and an Incyte clone, including Incyte EST 2472409 (from library 309 (THP1NOT03) constructed from THP-1 cells derived from the blood of a monocytic leukemia patient), the Incyte clone, including Incyte EST 2472409 was purchased and the cDNA insert was obtained and sequenced. It was found that the cDNA insert encoded a full-length protein. The sequence of this cDNA insert is shown in Figure 35 (SEQ ID NO:48) and is herein designated as DNA59842-1502.

Clone DNA59842-1502 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 92-94 and ending at the stop codon at nucleotide positions 683-685 (Figure 35; SEQ ID

NO:48). The predicted polypeptide precursor is 197 amino acids long (Figure 36). The full-length PRO1153 protein shown in Figure 36 (SEQ ID NO:49) has an estimated molecular weight of about 21,540 daltons and a pI of about 8.31. Analysis of the full-length PRO1153 sequence shown in Figure 36 (SEQ ID NO:49) evidences the presence of important polypeptide domains as shown in Figure 36, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1153 sequence shown in Figure 36 evidences the presence of the following: transmembrane domains from about amino acid 10 to about amino acid 28 and from about amino acid 85 to about amino acid 110, an N-glycosylation site from about amino acid 38 to about amino acid 41, and N-myristoylation sites from about amino acid 5 to about amino acid 10 and from about amino acid 88 to about amino acid 93. Clone DNA59842-1502 has been deposited with ATCC on June 16, 1998 and is assigned ATCC deposit no. 209982.

An analysis of the Dayhoff database (version 35.45 SwissProt), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 36 (SEQ ID NO:49), evidenced sequence identity between the PRO1153 amino acid sequence and the following Dayhoff sequences: S57447; SOYHRGPC_1; S46965; P_P82971; VCPHEROPH_1; EXTN_TOBAC; MLCB2548_9; ANXA_RABIT; JC5437 and SSGP_VOLCA.

EXAMPLE 19

Isolation of cDNA Clones Encoding a Human PRO1182

DNA59848-1512 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (e.g., GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence from the Incyte database, designated herein as 146647. This EST cluster sequence was then compared to variety of expressed sequence tag (EST) databases which included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA56033.

In light of an observed sequence homology between the DNA56033 sequence and the Incyte EST clone no. 2595195, the Incyte EST clone 2595195 was purchased and the cDNA insert was obtained and sequenced. It

was found that the cDNA insert encoded a full-length protein. The sequence of this cDNA insert is shown in Figure 37 (SEQ ID NO:50) and is herein designated as DNA59848-1512.

Clone DNA59848-1512 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 67-69 and ending at the stop codon at nucleotide positions 880-882 (Figure 37; SEQ ID NO:50). The predicted polypeptide precursor is 271 amino acids long (Figure 38). The full-length PRO1182 protein shown in Figure 38 (SEQ ID NO:51) has an estimated molecular weight of about 28,665 daltons and a pI of about 5.33. Analysis of the full-length PRO1182 sequence shown in Figure 38 (SEQ ID NO:51) evidences the presence of important polypeptide domains as shown in Figure 38, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1182 sequence shown in Figure 38 evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 25, an amino acid block having homology to C-type lectin domain proteins from about amino acid 247 to about amino acid 256 and an amino acid sequence block having homology to C1q domain proteins from about amino acid 44 to about amino acid 77. Clone DNA59848-1512 has been deposited with ATCC on August 4, 1998 and is assigned ATCC deposit no. 203088.

An analysis of the Dayhoff database (version 35.45 SwissProt), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 38 (SEQ ID NO:51), evidenced sequence identity between the PRO1182 amino acid sequence and the following Dayhoff sequences: PSPD_BOVIN, CL43_BOVIN, CONG_BOVIN, P_W18780, P_R45005, P_R53257 and CELEGAP7_1.

20

EXAMPLE 20

Isolation of cDNA Clones Encoding Human PRO1184

DNA59220-1514 was identified by applying a proprietary signal sequence finding algorithm developed by Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (e.g., GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

Use of the above described signal sequence algorithm allowed identification of an EST sequence from the Incyte database, designated herein as DNA56375.init. This EST sequence was then compared to variety of expressed sequence tag (EST) databases which included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using the computer program BLAST or BLAST2 (Altschul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the

program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA56375.

In light of an observed sequence homology between the DNA56375 sequence and an Incyte clone, including Incyte EST no. 1428374 (from an ileum tissue library (39, SINTBST01), the Incyte clone, including
5 Incyte EST no. 1428374, was further examined, and the cDNA insert was obtained and sequenced. It was found herein that the cDNA insert encoded a full-length protein. The sequence of this cDNA insert is shown in Figure 39 (SEQ ID NO:52) and is herein designated as DNA59220-1514.

Clone DNA59220-1514 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 106-108 and ending at the stop codon at nucleotide positions 532-534 (Figure 39; SEQ ID
10 NO:52). The predicted polypeptide precursor is 142 amino acids long (Figure 40). The full-length PRO1184 protein shown in Figure 40 (SEQ ID NO:53) has an estimated molecular weight of about 15,690 daltons and a pI of about 9.64. Analysis of the full-length PRO1184 sequence shown in Figure 40 (SEQ ID NO:53) evidences the presence of an important polypeptide domain as shown in Figure 40, wherein the location given for that important polypeptide domain is approximate as described above. Analysis of the full-length PRO1184 sequence shown in Figure 40
15 evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 38. Clone DNA59220-1514 has been deposited with ATCC on June 9, 1998 and is assigned ATCC deposit no. 209962.

Analysis of the amino acid sequence of the full-length PRO1184 polypeptide suggests that it possesses some sequence identity with a protein called TIM from *Drosophila virilis*. More specifically, an analysis of the Dayhoff database (version 35.45 SwissProt), using the ALIGN-2 sequence alignment analysis of the full-length sequence
20 shown in Figure 40 (SEQ ID NO:53), evidenced sequence identity between the PRO1184 amino acid sequence and the following Dayhoff sequences: WIS1_SCHPO, F002186_1, ATAC00239124 and MSAIPRP_1.

EXAMPLE 21

Isolation of cDNA Clones Encoding PRO1187

DNA62876-1517 was identified by applying a proprietary signal sequence finding algorithm developed by
25 Genentech, Inc., (South San Francisco, CA) upon ESTs as well as clustered and assembled EST fragments from public (e.g., GenBank) and/or private (LIFESEQ™, Incyte Pharmaceuticals, Inc., Palo Alto, CA) databases. The signal sequence algorithm computes a secretion signal score based on the character of the DNA nucleotides surrounding the first and optionally the second methionine codon(s) (ATG) at the 5'-end of the sequence or sequence
30 fragment under consideration. The nucleotides following the first ATG must code for at least 35 unambiguous amino acids without any stop codons. If the first ATG has the required amino acids, the second is not examined. If neither meets the requirement, the candidate sequence is not scored. In order to determine whether the EST sequence contains an authentic signal sequence, the DNA and corresponding amino acid sequences surrounding the ATG codon are scored using a set of seven sensors (evaluation parameters) known to be associated with secretion signals.

35 Use of the above described signal sequence algorithm allowed identification of an EST cluster sequence from the Incyte database. This EST cluster sequence was then compared to variety of expressed sequence tag (EST) databases which included public EST databases (e.g., GenBank) and a proprietary EST DNA database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA) to identify existing homologies. The homology search was performed using

the computer program BLAST or BLAST2 (Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)). Those comparisons resulting in a BLAST score of 70 (or in some cases, 90) or greater that did not encode known proteins were clustered and assembled into a consensus DNA sequence with the program "phrap" (Phil Green, University of Washington, Seattle, Washington). The consensus sequence obtained therefrom is herein designated DNA57726.

5 In light of an observed sequence homology between the DNA57726 sequence and the Incyte EST 358563 sequence, the clone (from a library constructed using RNA from synovial membrane tissue) including Incyte EST 358563 was purchased and the cDNA insert was obtained and sequenced. The sequence of this cDNA insert is shown in Figure 41 (SEQ ID NO:54) and is herein designated as DNA62876-1517.

10 Clone DNA62876-1517 contains a single open reading frame with an apparent translational initiation site at nucleotide positions 121-123 and ending at the stop codon at nucleotide positions 481-483 (Figure 41; SEQ ID NO:54). The predicted polypeptide precursor is 120 amino acids long (Figure 42). The full-length PRO1187 protein shown in Figure 42 (SEQ ID NO:55) has an estimated molecular weight of about 12,925 daltons and a pI of about 9.46. Analysis of the full-length PRO1187 sequence shown in Figure 42 (SEQ ID NO:55) evidences the presence of important polypeptide domains as shown in Figure 42, wherein the locations given for those important
15 polypeptide domains are approximate as described above. Analysis of the full-length PRO1187 sequence shown in Figure 42 evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 17, and N-myristoylation sites from about amino acid 58 to about amino acid 63, from about amino acid 63 to about amino acid 68, from about amino acid 64 to about amino acid 69, from about amino acid 83 to about amino acid 88, from about amino acid 111 to about amino acid 116 and from about amino acid 115 to about amino acid 120.
20 Clone DNA62876-1517 has been deposited with ATCC on August 4, 1998 and is assigned ATCC deposit no. 203095.

An analysis of the Dayhoff database (version 35.45 SwissProt), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 42 (SEQ ID NO:55), evidenced sequence identity between the PRO1187 amino acid sequence and the following Dayhoff sequences: MGNENDOBX_1, CELF41G3_9,
25 AMPG_STRLI, HSBBOVHERL_2, LEEXTEN10_1, AF029958_1 and P_W04957.

EXAMPLE 22

Isolation of cDNA Clones Encoding a Human PRO1281

The extracellular domain (ECD) sequences (including the secretion signal sequence, if any) from about 950
30 known secreted proteins from the Swiss-Prot public database were used to search EST databases. The EST databases included public EST databases (*e.g.*, GenBank) and a proprietary EST database (LIFESEQ™, Incyte Pharmaceuticals, Palo Alto, CA). The search was performed using the computer program BLAST or BLAST2 [Altshul *et al.*, Methods in Enzymology, 266:460-480 (1996)] as a comparison of the ECD protein sequences to a 6 frame translation of the EST sequences. Those comparisons resulting in a BLAST score of 70 (or in some cases,
35 90) or greater that did not encode known proteins were clustered and assembled into consensus DNA sequences with the program "phrap" (Phil Green, University of Washington, Seattle, Washington).

A consensus DNA sequence was assembled relative to other EST sequences using phrap as described above and is herein designated DNA35720. In some cases, the DNA35720 consensus sequence derives from an

intermediate consensus DNA sequence which was extended using repeated cycles of BLAST and phrap to extend that intermediate consensus sequence as far as possible using the sources of EST sequences discussed above.

Based on the DNA35720 consensus sequence, oligonucleotides were synthesized : 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence for PRO1281. Forward and reverse PCR primers generally range from 20 to 30 nucleotides and are often designed to give a PCR product of about 100-1000 bp in length. The probe sequences are typically 40-55 bp in length. In some cases, additional oligonucleotides are synthesized when the consensus sequence is greater than about 1-1.5kbp. In order to screen several libraries for a full-length clone, DNA from the libraries was screened by PCR amplification, as per Ausubel *et al.*, Current Protocols in Molecular Biology, *supra*, with the PCR primer pair.

A positive library was then used to isolate clones encoding the gene of interest using the probe oligonucleotide and one of the primer pairs.

PCR primers (forward and reverse) were synthesized:

forward PCR primers (35720.f1) 5'-TGGAAGGCTGCCGCAACGACAATC-3' (SEQ ID NO:58)
(35720.f2) 5'-CTGATGTGGCCGATGTTCTG-3' (SEQ ID NO:59)
15 (35720.f3) 5'-ATGGCTCAGTGTGCAGACAG-3' (SEQ ID NO:60)
reverse PCR primers (35720.r1) 5'-GCATGCTGCTCCGTGAAGTAGTCC-3' (SEQ ID NO:61)
(32720.r2) 5'-ATGCATGGGAAAGAAGGCCTGCCC-3' (SEQ ID NO:62)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA35720 sequence which had the following nucleotide sequence:

20 hybridization probe

5'-TGCACCTGGTGACCACGAGGGGGTGCACCTATAGCCATCTGGAGCTGAG-3' (SEQ ID NO:63)

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue. The cDNA libraries used to isolate the cDNA clones were constructed by standard methods using commercially available reagents such as those from Invitrogen, San Diego, CA. The cDNA was primed with oligo dT containing a NotI site, linked with blunt to SalI hemikinased adaptors, cleaved with NotI, sized appropriately by gel electrophoresis, and cloned in a defined orientation into a suitable cloning vector (such as pRKB or pRKD; pRK5B is a precursor of pRK5D that does not contain the SfiI site; see, Holmes *et al.*, Science, 253:1278-1280 (1991)) in the unique XhoI and NotI sites.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for a full-length PRO1281 polypeptide (designated herein as DNA59820-1549 [Figures 43A-43B, SEQ ID NO:56]) and the derived protein sequence for that PRO1281 polypeptide.

The full length clone identified above contained a single open reading frame with an apparent translational initiation site at nucleotide positions 228-230 and a stop signal at nucleotide positions 2553-2555 (Figures 43A-43B, SEQ ID NO:56). The predicted polypeptide precursor is 775 amino acids long (Figure 44), has a calculated molecular weight of approximately 85,481 daltons and an estimated pI of approximately 6.92. Analysis of the full length PRO1281 sequence shown in Figure 44 (SEQ ID NO:57) evidences the presence of a variety of important polypeptide domains as shown in Figure 44, wherein the locations given for those important polypeptide domains are approximate as described above. Analysis of the full-length PRO1281 sequence shown in Figure 44 evidences the presence of the following: a signal peptide from about amino acid 1 to about amino acid 15, and potential N-

glycosylation sites from about amino acid 138 to about amino acid 141 and from about amino acid 361 to about amino acid 364. Clone DNA59820-1549 has been deposited with the ATCC on August 18, 1998 and is assigned ATCC deposit no. 203129.

5 An analysis of the Dayhoff database (version 35.45 SwissProt 35), using the ALIGN-2 sequence alignment analysis of the full-length sequence shown in Figure 44 (SEQ ID NO:57) evidenced sequence identity between the PRO1281 amino acid sequence and the following Dayhoff sequences: S44860, CET24D1_1, CEC38H2_3, CAC2_HAECO, B3A2_HUMAN, S22373, CEF38A3_2, CEC34F6_2, CEC34F6_3 and CELT22B11_3.

EXAMPLE 23

10 Gene Amplification

This example shows that the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding genes are amplified in the genome of certain human lung, colon and/or
15 breast cancers and/or cell lines. Amplification is associated with overexpression of the gene product, indicating that the polypeptides are useful targets for therapeutic intervention in certain cancers such as colon, lung, breast and other cancers. Therapeutic agents may take the form of antagonists of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710,
20 PRO2094, PRO2145 or PRO2198 polypeptide, for example, murine-human chimeric, humanized or human antibodies against a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

25 The starting material for the screen was genomic DNA isolated from a variety cancers. The DNA is quantitated precisely, *e.g.*, fluorometrically. As a negative control, DNA was isolated from the cells of ten normal healthy individuals which was pooled and used as assay controls for the gene copy in healthy individuals (not shown). The 5' nuclease assay (for example, TaqMan™) and real-time quantitative PCR (for example, ABI Prizm 7700 Sequence Detection System™ (Perkin Elmer, Applied Biosystems Division, Foster City, CA)), were used to
30 find genes potentially amplified in certain cancers. The results were used to determine whether the DNA encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 is over-represented in any of the primary lung or colon cancers or cancer cell lines or breast cancer cell lines that were screened. The
35 primary lung cancers were obtained from individuals with tumors of the type and stage as indicated in Table 2. An explanation of the abbreviations used for the designation of the primary tumors listed in Table 2 and the primary tumors and cell lines referred to throughout this example has been given hereinbefore.

The results of the Taqman™ are reported in delta (Δ) Ct units. One unit corresponds to 1 PCR cycle or

approximately a 2-fold amplification relative to normal, two units corresponds to 4-fold, 3 units to 8-fold amplification and so on. Quantitation was obtained using primers and a TaqmanTM fluorescent probe derived from the PRO212-, PRO290-, PRO341-, PRO535-, PRO619-, PRO717-, PRO809-, PRO830-, PRO848-, PRO943-, PRO1005-, PRO1009-, PRO1025-, PRO1030-, PRO1097-, PRO1107-, PRO1111-, PRO1153-, PRO1182-, PRO1184-, PRO1187-, PRO1281-, PRO23-, PRO39-, PRO834-, PRO1317-, PRO1710-, PRO2094-, PRO2145- or PRO2198-encoding gene. Regions of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 which are most likely to contain unique nucleic acid sequences and which are least likely to have spliced out introns are preferred for the primer and probe derivation, *e.g.*, 3-untranslated regions. The sequences for the primers and probes (forward, reverse and probe) used for the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 gene amplification analysis were as follows:

15

PRO212 (DNA30942-1134)

30942.3unt-probe:

5'-CCCTGTGCACTGATCCTGGCCC-3' (SEQ ID NO:80)

30942.3unt-5:

20 5'-AGCGGAGCGTCCGTGA-3' (SEQ ID NO:81)

30942.3unt-3:

5'-CCACTTGCACTGAAAGAGGCT-3' (SEQ ID NO:82)

30942.tm.p2:

5'-ACACGATGCGTGCTCCAAGCAGAA-3' (SEQ ID NO:83)

25 30942.tm.f2:

5'-CTTCTTCGCGCACGCTG-3' (SEQ ID NO:84)

30942.tm.r2:

5'-ATCACGCCGGCACCAG-3' (SEQ ID NO:85)

30942.dom1.f3

30 5'-ACGCGGAGTGGCAGAAAC-3' (SEQ ID NO:86)

30942.dom1.r3:

5'-CACTGGGCGCACACCA-3' (SEQ ID NO:87)

30942.dom1.p3:

5'-TACCCCTGGCGGGACGCAGAG-3' (SEQ ID NO:88)

35 30942.dom4a.f4:

5'-CACCTTCTCAGCCAGCAGCT-3' (SEQ ID NO:89)

30942.dom4a.r4:

5'-GAGGAAGAGCCTGGCACATT-3' (SEQ ID NO:90)

- 30942.dom4a.p4:
5'-AACTGCACGGCCCTGGGCCT-3' (SEQ ID NO:91)
- 30942.dom4b.f5:
5'-CCTCAATGTGCCAGGCTCTT-3' (SEQ ID NO:92)
- 5 30942.dom4b.r5:
5'-CCTGGTACCCTGGTGCTGA-3' (SEQ ID NO:93)
- 30942.dom4b.p5:
5'-CACCAGCTGCACTGGCTTCCCC-3' (SEQ ID NO:94)
- 10 PRO290 (DNA35680-1212):
35680.tm.p:
5'-CCACCAATGGCAGCCCCACCT-3' (SEQ ID NO:95)
35680.tm.f:
5'-GACTGCCCTCCCTGCCA-3' (SEQ ID NO:96)
- 15 35680.tm.r:
5'-CAAAAAGCCTGGAAGTCTTCAAAG-3' (SEQ ID NO:97)
- PRO341 (DNA26288-1239):
26288.tm.fl:
- 20 5'-CAGCTGGACTGCAGGTGCTA-3' (SEQ ID NO:98)
26288.tm.rl:
5'-CAGTGAGCACAGCAAGTGTCT-3' (SEQ ID NO:99)
26288.tm.pl:
5'-GGCCACCTCCTTGAGTCTTCAGTTCCT-3' (SEQ ID NO:100)
- 25 PRO535 (DNA49143-1429):
49143.tm.fl:
5'-CAACTACTGGCTAAAGCTGGTGAA-3' (SEQ ID NO:101)
49143.tm.rl:
- 30 5'-CCTTTCTGTATAGGTGATACCCAATGA-3' (SEQ ID NO:102)
49143.tm.pl:
5'-TGGCCATCCCTACCAGAGGCAAAA-3' (SEQ ID NO:103)
- PRO619 (DNA49821-1562):
- 35 49821.tm.fl:
5'-CTGAAGACGACGCGGATTACTA-3' (SEQ ID NO:104)
49821.tm.rl:
5'-GGCAGAAATGGGAGGCAGA-3' (SEQ ID NO:105)

49821.tm.p1:

5'-TGCTCTGTTGGCTACGGCTTTAGTCCCTAG-3' (SEQ ID NO:106)

PRO717 (DNA50988-1326):

5 50988.tm.f3:

5'-CAAGCGTCCAGGTTTATTGA-3' (SEQ ID NO:107)

50988.tm.r3:

5'-GACTACAAGGCGCTCAGCTA-3' (SEQ ID NO:108)

50988.tm.p3:

10 5'-CCGGCTGGGTCTCACTCCTCC-3' (SEQ ID NO:109)

PRO809 (DNA57836-1338):

57836.tm.f1:

5'-AGCAGCAGCCATGTAGAATGAA-3' (SEQ ID NO:110)

15 57836.tm.r1:

5'-AATACGAACAGTGCACGCTGAT-3' (SEQ ID NO:111)

57836.tm.p1:

5'-TCCAGAGAGCCAAGCACGGCAGA-3' (SEQ ID NO:112)

20 PRO830 (DNA56866-1342):

56866.tm.f1:

5'-TCTAGCCAGCTTGGCTCCAATA-3' (SEQ ID NO:113)

56866.tm.r1:

5'-CCTGGCTCTAGCACCAACTCATA-3' (SEQ ID NO:114)

25 56866.tm.p1:

5'-TCAGTGGCCCTAAGGAGATGGGCCT-3' (SEQ ID NO:115)

PRO848 (DNA59839-1461):

59839.tm.f1:

30 5'-CAGGATACAGTGGGAATCTTGAGA-3' (SEQ ID NO:116)

59839.tm.r1:

5'-CCTGAAGGGCTTGGAGCTTAGT-3' (SEQ ID NO:117)

59839.tm.p1:

5'-TCTTTGGCCATTTCCCATGGCTCA-3' (SEQ ID NO:118)

35

PRO943 (DNA52192-1369):

52192.tm.f1:

5'-CCCATGGCGAGGAGGAAT-3' (SEQ ID NO:119)

52192.tm.r1:

5'-TGCGTACGTGTGCCTTCAG-3' (SEQ ID NO:120)

52192.tm.p1:

5'-CAGCACCCCAGGCAGTCTGTGTGT-3' (SEQ ID NO:121)

5

PRO1005 (DNA57708-1411):

57708.tm.f1:

5'-AACGTGCTACACGACCAGTGTACT-3' (SEQ ID NO:122)

57708.tm.r1:

10 5'-CACAGCATATTCAGATGACTAAATCCA-3' (SEQ ID NO:123)

57708.tm.p1:

5'-TTGTTTAGTTCTCCACCGTGTCTCCACAGAA-3' (SEQ ID NO:124)

PRO1009 (DNA57129-1413):

15 57129.tm.f1:

5'-TGTCAGAATGCAACCTGGCTT-3' (SEQ ID NO:125)

57129.tm.r1:

5'-TGATGTGCCTGGCTCAGAAC-3' (SEQ ID NO:126)

57129.tm.p1:

20 5'-TGCACCTAGATGTCCCCAGCACCC-3' (SEQ ID NO:127)

PRO1025 (DNA59622-1334):

59622.tm.f1:

5'-TGATTGGTTCCCAGTGGCTT-3' (SEQ ID NO:128)

25 59622.tm.r1:

5'-GTGGTGGCAGCAGCTTCCT-3' (SEQ ID NO:129)

59622.tm.p1:

5'-CCCCACCCCCTTATAGCATCTCCCTC-3' (SEQ ID NO:130)

30 PRO1030 (DNA59485-1336):

59485.tm.f1:

5'-AATCTCCAAGCCCTTCTGTCTGT-3' (SEQ ID NO:131)

59485.tm.r1:

5'-TGCTTCCACACTAGCCAGTCTTC-3' (SEQ ID NO:132)

35 59485.tm.p1:

5'-CACACTTCTTCTGGTTTCAAGTCTCAAGGCCT-3' (SEQ ID NO:133)

PRO1097 (DNA59841-1460):

59841.tm.fl:

5'-AAGATGCGCCAGGCTTCTTA-3' (SEQ ID NO:134)

59841.tm.rl:

5 5'-CTCCTGTACGGTCTGCTCACTTAT-3' (SEQ ID NO:135)

59841.tm.pl:

5'-TGGCTGTCTAGTCCAGTGTGCATGG-3' (SEQ ID NO:136)

PRO1107 (DNA59606-1471):

10 59606.tm.fl:

5'-GCATAGGGATAGATAAGATCCTGCTTTAT-3' (SEQ ID NO:137)

59606.tm.rl:

5'-CAAATTAAAGTACCCATCAGGAGAGAA-3' (SEQ ID NO:138)

59606.tm.pl:

15 5'-AAGTTGCTAAATATATACATTATCTGCGCCAAGTCCA-3' (SEQ ID NO:139)

PRO1111 (DNA58721-1475):

58721.tm.fl:

5'-GTGCTGCCCAACAATTCATGA-3' (SEQ ID NO:140)

20 58721.tm.rl:

5'-GTCCTTGGTATGGGTCTGAATTATAT-3' (SEQ ID NO:141)

58721.tm.pl:

5'-ACTCTCTGCACCCACAGTCACCACTATCTC-3' (SEQ ID NO:142)

25 PRO1153 (DNA59842-1502):

59842.tm.fl:

5'-CTGAGGAACCAGCCATGTCTCT-3' (SEQ ID NO:143)

59842.tm.rl:

5'-GACCAGATGCAGGTACAGGATGA-3' (SEQ ID NO:144)

30 59842.tm.pl:

5'-CTGCCCCTTCAGTGATGCCAACCTT-3' (SEQ ID NO:145)

PRO1182 (DNA59848-1512):

59848.tm.fl:

35 5'-GGGTGGAGGCTCACTGAGTAGA-3' (SEQ ID NO:146)

59848.tm.rl:

5'-CAATACAGGTAATGAACTCTGCTTCTT-3' (SEQ ID NO:147)

59848.tm.pl:

5'-TCCTCTTAAGCATAGGCCATTTTCTCAGTTTAGACA-3' (SEQ ID NO:148)

PRO1184 (DNA59220-1514):

5 59220.tm.fl:

5'-GGTGGTCTTGCTTGGTCTCAC-3' (SEQ ID NO:149)

59220.tm.rl:

5'-CCGTCGTTTCAGCAACATGAC-3' (SEQ ID NO:150)

59220.tm.pl:

10 5'-ACCGCCTACCGCTGTGCCCA-3' (SEQ ID NO:151)

PRO1187 (DNA62876-1517):

62876.tm.fl:

5'-CAGTAAAACACAGGCTGGATTT-3' (SEQ ID NO:152)

15 62876.tm.rl:

5'-CCTGAGAGCAAGAAGGTTGAGAAT-3' (SEQ ID NO:153)

62876.tm.pl:

5'-TAGACAGGGACCATGGCCCGCA-3' (SEQ ID NO:154)

20 PRO1281 (DNA59820-1549):

59820.tm.fl:

5'-TGGGCTGTAGAAGAGTTGTTG-3' (SEQ ID NO:155)

59820.tm.rl:

5'-TCCACACTTGCCAGTTTAT-3' (SEQ ID NO:156)

25 59820.tm.pl:

5'-CCCAACTTCTCCCTTTTGGACCCT-3' (SEQ ID NO:157)

PRO23 (DNA36640):

36640.tm.fl:

30 5'-TTTCATGACCTCCGAGAAGA-3' (SEQ ID NO:158)

36640.tm.rl:

5'-TTGTAGGCGTTCTTGATGATG-3' (SEQ ID NO:159)

36640.tm.pl:

5'-CCCCTGGTGACGCTGTTCAAAAA-3' (SEQ ID NO:160)

35

PRO39 (DNA36651):

36651.tm.fl:

5'-GCGCGTGGACACCAAGTC-3' (SEQ ID NO:161)

36651.tm.r1:

5'-CGAAGCAGCCCTTGGACA-3' (SEQ ID NO:162)

36651.tm.p1:

5'-ACCCCAACGCGCGCAAATACAAA-3' (SEQ ID NO:163)

5

PRO834 (DNA 56538):

56538.tm.fl:

5'-GGGAGATGTCCCTGGAAGAA-3' (SEQ ID NO:164)

56538.tm.r1:

10 5'-TGTCTGGCTTCACAGCACTGT-3' (SEQ ID NO:165)

56538.tm.p1:

5'-TTTTTTTAGGTCTGCATTACACTGGCTGT-3' (SEQ ID NO:166)

PRO1317 (DNA71166):

15 71166.tm.fl:

5'-CCCTAGCTGACCCCTTCA-3' (SEQ ID NO:167)

71166.tm.r1:

5'-TCTGACAAGCAGTTTTCTGAATC-3' (SEQ ID NO:168)

71166.tm.p1:

20 5'-CTCTCCCCCTCCCTTTTCCTTTGTTT-3' (SEQ ID NO:169)

PRO1710 (DNA82331):

82331.tm.fl:

5'-TGCTTCGGGCCCCAATATCT-3' (SEQ ID NO:170)

25 82331.tm.r1:

5'-CGACGGCAGGTAGTTCTCCT-3' (SEQ ID NO:171)

82331.tm.p1:

5'-CGTGGGCACCGCCGAAGC-3' (SEQ ID NO:172)

30 PRO2094 (DNA83123):

83123.tm.fl:

5'-CCCATGTTCAATGCAGTTTG-3' (SEQ ID NO:173)

83123.tm.r1:

5'-AGTCCTTGGAGGTCCTATCCA-3' (SEQ ID NO:174)

35 83123.tm.p1:

5'-AAGCAAGGCTGATAAAAAAGCTGCCC-3' (SEQ ID NO:175)

PRO2145 (DNA88050):

88050.tm.fl:

5'-GGCCTTCTCGGTCTTCAC-3' (SEQ ID NO:176)

88050.tm.rl:

5 5'-AAGTTGGGAGCCGACATC-3' (SEQ ID NO:177)

88050.tm.pl:

5'-ATCATCTGCACCATCGGCATCC-3' (SEQ ID NO:178)

PRO2198 (DNA88153):

10 88153.tm.fl:

5'-TGGAGGCTGCAGATAGTGA-3' (SEQ ID NO:179)

88153.tm.rl:

5'-CGTCACCCTCGTAGTCATAGAT-3' (SEQ ID NO:180)

88153.tm.pl:

15 5'-CCCAGTGTGCCGCCTTACGA-3' (SEQ ID NO:181)

The 5' nuclease assay reaction is a fluorescent PCR-based technique which makes use of the 5' exonuclease activity of Taq DNA polymerase enzyme to monitor amplification in real time. Two oligonucleotide primers are used to generate an amplicon typical of a PCR reaction. A third oligonucleotide, or probe, is designed to detect nucleotide sequence located between the two PCR primers. The probe is non-extendible by Taq DNA polymerase enzyme, and is labeled with a reporter fluorescent dye and a quencher fluorescent dye. Any laser-induced emission from the reporter dye is quenched by the quenching dye when the two dyes are located close together as they are on the probe. During the amplification reaction, the Taq DNA polymerase enzyme cleaves the probe in a template-dependent manner. The resultant probe fragments disassociate in solution, and signal from the released reporter dye is free from the quenching effect of the second fluorophore. One molecule of reporter dye is liberated for each new molecule synthesized, and detection of the unquenched reporter dye provides the basis for quantitative interpretation of the data.

The 5' nuclease procedure is run on a real-time quantitative PCR device such as the ABI Prism 7700TM Sequence Detection. The system consists of a thermocycler, laser, charge-coupled device (CCD) camera and computer. The system amplifies samples in a 96-well format on a thermocycler. During amplification, laser-induced fluorescent signal is collected in real-time through fiber optics cables for all 96 wells, and detected at the CCD. The system includes software for running the instrument and for analyzing the data.

5' Nuclease assay data are initially expressed as Ct, or the threshold cycle. This is defined as the cycle at which the reporter signal accumulates above the background level of fluorescence. The ΔC_t values are used as quantitative measurement of the relative number of starting copies of a particular target sequence in a nucleic acid sample when comparing cancer DNA results to normal human DNA results.

Table 2 describes the stage, T stage and N stage of various primary tumors which were used to screen the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005,

PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 compounds of the invention.

Table 2
Primary Lung and Colon Tumor Profiles

	<u>Primary Tumor</u>	<u>Stage</u>	<u>Other Stage</u>	<u>Dukes Stage</u>	<u>T Stage</u>	<u>N Stage</u>
5	Human lung tumor AdenoCa (SRCC724) [LT1]	IIA			T1	N1
	Human lung tumor SqCCa (SRCC725) [LT1a]	IIB			T3	N0
	Human lung tumor AdenoCa (SRCC726) [LT2]	IB			T2	N0
	Human lung tumor AdenoCa (SRCC727) [LT3]	IIIA			T1	N2
	Human lung tumor AdenoCa (SRCC728) [LT4]	IB			T2	N0
10	Human lung tumor SqCCa (SRCC729) [LT6]	IB			T2	N0
	Human lung tumor Aden/SqCCa (SRCC730) [LT7]	IA			T1	N0
	Human lung AdenoCa (SRCC825) [LT8]	-			-	-
	Human lung tumor AdenoCa (SRCC731) [LT9]	IB			T2	N0
	Human lung tumor SqCCa (SRCC732) [LT10]	IIB			T2	N1
15	Human lung tumor SqCCa (SRCC733) [LT11]	IIA			T1	N1
	Human lung tumor AdenoCa (SRCC734) [LT12]	IV			T2	N0
	Human lung tumor AdenoSqCCa (SRCC735)[LT13]	IB			T2	N0
	Human lung tumor SqCCa (SRCC736) [LT15]	IB			T2	N0
	Human lung tumor SqCCa (SRCC737) [LT16]	IB			T2	N0
20	Human lung tumor SqCCa (SRCC738) [LT17]	IIB			T2	N1
	Human lung tumor SqCCa (SRCC739) [LT18]	IB			T2	N0
	Human lung tumor SqCCa (SRCC740) [LT19]	IB			T2	N0
	Human lung tumor LCCa (SRCC741) [LT21]	IIB			T3	N1
	Human lung AdenoCa (SRCC811) [LT22]	IA			T1	N0
25	Human colon AdenoCa (SRCC742) [CT2]		M1	D	pT4	N0
	Human colon AdenoCa (SRCC743) [CT3]			B	pT3	N0
	Human colon AdenoCa (SRCC 744) [CT8]			B	T3	N0
	Human colon AdenoCa (SRCC745) [CT10]			A	pT2	N0
	Human colon AdenoCa (SRCC746) [CT12]		MO, R1	B	T3	N0
30	Human colon AdenoCa (SRCC747) [CT14]		pMO, RO	B	pT3	pN0
	Human colon AdenoCa (SRCC748) [CT15]		M1, R2	D	T4	N2
	Human colon AdenoCa (SRCC749) [CT16]		pMO	B	pT3	pN0
	Human colon AdenoCa (SRCC750) [CT17]			C1	pT3	pN1
	Human colon AdenoCa (SRCC751) [CT1]		MO, R1	B	pT3	N0
35	Human colon AdenoCa (SRCC752) [CT4]			B	pT3	M0
	Human colon AdenoCa (SRCC753) [CT5]		G2	C1	pT3	pN0
	Human colon AdenoCa (SRCC754) [CT6]		pMO, RO	B	pT3	pN0
	Human colon AdenoCa (SRCC755) [CT7]		G1	A	pT2	pN0
	Human colon AdenoCa (SRCC756) [CT9]		G3	D	pT4	pN2
40	Human colon AdenoCa (SRCC757) [CT11]			B	T3	N0
	Human colon AdenoCa (SRCC758) [CT18]		MO, RO	B	pT3	pN0

DNA Preparation:

45 DNA was prepared from cultured cell lines, primary tumors, normal human blood. The isolation was performed using purification kit, buffer set and protease and all from Quiagen, according to the manufacturer's instructions and the description below.

Cell culture lysis:

50 Cells were washed and trypsinized at a concentration of 7.5×10^8 per tip and pelleted by centrifuging at 1000 rpm for 5 minutes at 4°C, followed by washing again with 1/2 volume of PBS recentrifugation. The pellets were washed a third time, the suspended cells collected and washed 2x with PBS. The cells were then suspended into 10 mL PBS. Buffer C1 was equilibrated at 4°C. Qiagen protease #19155 was diluted into 6.25 ml cold ddH₂O

to a final concentration of 20 mg/ml and equilibrated at 4°C. 10 mL of G2 Buffer was prepared by diluting Qiagen RNase A stock (100 mg/ml) to a final concentration of 200 µg/ml.

Buffer C1 (10 mL, 4°C) and ddH₂O (40 mL, 4°C) were then added to the 10 mL of cell suspension, mixed by inverting and incubated on ice for 10 minutes. The cell nuclei were pelleted by centrifuging in a Beckman swinging bucket rotor at 2500 rpm at 4°C for 15 minutes. The supernatant was discarded and the nuclei were suspended with a vortex into 2 mL Buffer C1 (at 4°C) and 6 mL ddH₂O, followed by a second 4°C centrifugation at 2500 rpm for 15 minutes. The nuclei were then resuspended into the residual buffer using 200 µl per tip. G2 buffer (10 ml) was added to the suspended nuclei while gentle vortexing was applied. Upon completion of buffer addition, vigorous vortexing was applied for 30 seconds. Quiagen protease (200 µl, prepared as indicated above) was added and incubated at 50°C for 60 minutes. The incubation and centrifugation was repeated until the lysates were clear (e.g., incubating additional 30-60 minutes, pelleting at 3000 x g for 10 min., 4°C).

Solid human tumor sample preparation and lysis:

Tumor samples were weighed and placed into 50 ml conical tubes and held on ice. Processing was limited to no more than 250 mg tissue per preparation (1 tip/preparation). The protease solution was freshly prepared by diluting into 6.25 ml cold ddH₂O to a final concentration of 20 mg/ml and stored at 4°C. G2 buffer (20 ml) was prepared by diluting DNase A to a final concentration of 200 mg/ml (from 100 mg/ml stock). The tumor tissue was homogenated in 19 ml G2 buffer for 60 seconds using the large tip of the polytron in a laminar-flow TC hood to order to avoid inhalation of aerosols, and held at room temperature. Between samples, the polytron was cleaned by spinning at 2 x 30 seconds each in 2L ddH₂O, followed by G2 buffer (50 ml). If tissue was still present on the generator tip, the apparatus was disassembled and cleaned.

Quiagen protease (prepared as indicated above, 1.0 ml) was added, followed by vortexing and incubation at 50°C for 3 hours. The incubation and centrifugation was repeated until the lysates were clear (e.g., incubating additional 30-60 minutes, pelleting at 3000 x g for 10 min., 4°C).

Human blood preparation and lysis:

Blood was drawn from healthy volunteers using standard infectious agent protocols and citrated into 10 ml samples per tip. Quiagen protease was freshly prepared by dilution into 6.25 ml cold ddH₂O to a final concentration of 20 mg/ml and stored at 4°C. G2 buffer was prepared by diluting RNase A to a final concentration of 200 µg/ml from 100 mg/ml stock. The blood (10 ml) was placed into a 50 ml conical tube and 10 ml C1 buffer and 30 ml ddH₂O (both previously equilibrated to 4°C) were added, and the components mixed by inverting and held on ice for 10 minutes. The nuclei were pelleted with a Beckman swinging bucket rotor at 2500 rpm, 4°C for 15 minutes and the supernatant discarded. With a vortex, the nuclei were suspended into 2 ml C1 buffer (4°C) and 6 ml ddH₂O (4°C). Vortexing was repeated until the pellet was white. The nuclei were then suspended into the residual buffer using a 200 µl tip. G2 buffer (10 ml) were added to the suspended nuclei while gently vortexing, followed by vigorous vortexing for 30 seconds. Quiagen protease was added (200 µl) and incubated at 50°C for 60 minutes. The incubation and centrifugation was repeated until the lysates were clear (e.g., incubating additional 30-60 minutes, pelleting at 3000 x g for 10 min., 4°C).

Purification of cleared lysates:

(1) Isolation of genomic DNA:

Genomic DNA was equilibrated (1 sample per maxi tip preparation) with 10 ml QBT buffer. QF elution buffer was equilibrated at 50°C. The samples were vortexed for 30 seconds, then loaded onto equilibrated tips and
5 drained by gravity. The tips were washed with 2 x 15 ml QC buffer. The DNA was eluted into 30 ml silanized, autoclaved 30 ml Corex tubes with 15 ml QF buffer (50°C). Isopropanol (10.5 ml) was added to each sample, the tubes covered with parafin and mixed by repeated inversion until the DNA precipitated. Samples were pelleted by centrifugation in the SS-34 rotor at 15,000 rpm for 10 minutes at 4°C. The pellet location was marked, the supernatant discarded, and 10 ml 70% ethanol (4°C) was added. Samples were pelleted again by centrifugation on
10 the SS-34 rotor at 10,000 rpm for 10 minutes at 4°C. The pellet location was marked and the supernatant discarded. The tubes were then placed on their side in a drying rack and dried 10 minutes at 37°C, taking care not to overdry the samples.

After drying, the pellets were dissolved into 1.0 ml TE (pH 8.5) and placed at 50°C for 1-2 hours. Samples were held overnight at 4°C as dissolution continued. The DNA solution was then transferred to 1.5 ml tubes with
15 a 26 gauge needle on a tuberculin syringe. The transfer was repeated 5x in order to shear the DNA. Samples were then placed at 50°C for 1-2 hours.

(2) Quantitation of genomic DNA and preparation for gene amplification assay:

The DNA levels in each tube were quantified by standard A260, A280 spectrophotometry on a 1:20 dilution (5 µl DNA + 95 µl ddH₂O) using the 0.1 ml quartz cuvetts in the Beckman DU640 spectrophotometer.
20 A260/A280 ratios were in the range of 1.8-1.9. Each DNA samples was then diluted further to approximately 200 ng/ml in TE (pH 8.5). If the original material was highly concentrated (about 700 ng/µl), the material was placed at 50°C for several hours until resuspended.

Fluorometric DNA quantitation was then performed on the diluted material (20-600 ng/ml) using the manufacturer's guidelines as modified below. This was accomplished by allowing a Hoeffer DyNA Quant 200
25 fluorometer to warm-up for about 15 minutes. The Hoechst dye working solution (#H33258, 10 µl, prepared within 12 hours of use) was diluted into 100 ml 1 x TNE buffer. A 2 ml cuvette was filled with the fluorometer solution, placed into the machine, and the machine was zeroed. pGEM 3Zf(+) (2 µl, lot #360851026) was added to 2 ml of fluorometer solution and calibrated at 200 units. An additional 2 µl of pGEM 3Zf(+) DNA was then tested and the reading confirmed at 400 +/- 10 units. Each sample was then read at least in triplicate. When 3 samples were found
30 to be within 10% of each other, their average was taken and this value was used as the quantification value.

The fluorometrically determined concentration was then used to dilute each sample to 10 ng/µl in ddH₂O. This was done simultaneously on all template samples for a single TaqMan plate assay, and with enough material to run 500-1000 assays. The samples were tested in triplicate with TaqmanTM primers and probe both B-actin and GAPDH on a single plate with normal human DNA and no-template controls. The diluted samples were used
35 provided that the CT value of normal human DNA subtracted from test DNA was +/- 1 Ct. The diluted, lot-qualified genomic DNA was stored in 1.0 ml aliquots at -80°C. Aliquots which were subsequently to be used in the gene amplification assay were stored at 4°C. Each 1 ml aliquot is enough for 8-9 plates or 64 tests.

Gene amplification assay:

The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198

5 compounds of the invention were screened in the following primary tumors and the resulting ΔC_t values are reported in Tables 3A-3D.

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Table 3A

 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	LT-1	0.15 -1.05 -0.46 -0.55 -2.42 -1.99 -1.75 -1.68 -1.56 -2.02 -0.17 -0.44	0.26	-0.17	---	0.39 -0.02	---	---	0.69	0.44
15										
20										
	LT-1a	-0.12 0.55 0.80 -0.56 -0.58 -0.49 -0.27 -0.18 -0.46 1.34 0.63	0.63	0.47	---	0.83 0.36	---	---	1.13	0.67
25										
30										
35	LT2	-0.50 0.35 -0.91 -0.13 0.13 0.12 -0.44 -0.04 -0.46	-0.21	0.43	---	0.34 0.53	---	---	0.44	0.78
40										

Table 3A Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	LT3	0.56 0.64 0.69 0.85 0.84 1.45 0.29 0.34 0.66 0.88 0.57	0.80	0.33	---	1.04 1.68	---	---	0.89	0.99
15										
20	LT4	-0.27 0.33 -1.25 -0.50 -0.37 0.79 -0.73 -0.60 -0.17	0.37	0.74	---	0.77 0.02	---	---	0.43	0.49
25										
30	LT6	0.02 0.26 -0.58 -1.10 -0.99 -0.31 -0.51 -0.45 -0.20 0.43 -0.15	0.50	0.51	---	0.70 0.41	---	---	0.89	0.26
35										
40										

Table 3A Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	LT7	0.07 -0.13 -0.41 -1.62 -0.63 -0.75 -0.52 -0.49 1.15 0.47	0.71	0.17	---	1.21 1.34	---	---	0.95	0.82
15										
20	LT9	-0.52 -0.26 0.15 -1.35 -1.27 0.51 -1.12 -1.21 0.38 -0.31 -0.69	0.73	0.21	---	1.19 1.63	---	---	0.65	0.44
25										
30										
35	LT10	0.03 0.15 0.26 -0.31 -0.17 1.17 -0.25 -0.28 0.91 1.91 -0.12	0.75	0.24	---	1.41 2.02	---	---	0.67	0.76
40										
45										

Table 3A Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	LT11	1.56 1.96 1.16 0.66 0.70 -0.06 0.92 0.01 1.68 0.35	1.63	0.64 0.91	1.40 0.66	1.69 1.57 0.18	---	1.40	1.25	1.04
15										
20	LT12	4.19 6.41 3.87 0.20 3.28 3.06 3.58 2.90 3.87 2.71 5.40 4.44	0.95	0.69 0.32	0.96 0.51	1.81 0.10	---	1.61	1.35	1.22
25										
30										
35	LT13	2.61 3.91 2.01 0.09 1.47 1.39 1.89 1.10 2.32 1.54 1.97	1.47	0.17 -0.05	1.37 0.67	2.13 1.74 0.26	7.03	1.03 -0.09	0.98	0.74
40										
45										

Table 3A Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	LT15	3.25	1.67	0.72	0.65	2.08	2.71	0.76	0.88	0.63
		4.02		0.54	0.35	1.52		0.45		
		3.17				0.21				
		-0.16								
		1.77								
15		2.26								
		2.64								
		2.30								
		3.28								
		2.31								
20	LT16	-0.67								
		0.87	0.33	1.12	0.82	0.85	5.48	0.30	0.69	0.71
		0.30		0.14	0.50	0.74		-0.37		
		1.85				0.24				
		0.53								
25		1.12								
		1.56								
		0.50								
		1.13								
		0.88								
30		1.35								
		0.89								
35	LT17	1.16	1.22	1.33	1.42	1.83	---	1.10	1.17	0.78
		3.83		-0.36	0.63	1.67				
		0.93				0.46				
		0.38								
		1.12								
40		0.80								
		1.06								
		0.31								
		1.38								
		0.34								
45										

Table 3A Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	LT18	0.22	-0.31	0.57	-0.08	1.32	---	0.33	0.06	0.42
		0.05		0.34	0.05	1.14				
		0.90				0.65				
		0.14								
		0.07								
15		0.29								
		-0.44								
		0.19								
		-0.51								
		-0.06								
20	LT19	3.53	2.07	0.42	0.88	2.33	---	0.63	1.31	0.76
		3.67		0.12	0.74	1.90				
		0.28				0.10				
		2.27								
		2.92								
		3.27								
25										
	LT21	0.45	0.83	1.15	0.82	1.15	---	1.05	0.80	1.07
		0.02		0.43	0.69	1.09				
		0.06				0.27				
30		0.30								
		0.08								
		0.24								
35	CT2	4.44	1.56	---	0.80	1.22	---	0.07	---	0.37
		3.20				0.19				
		4.66								
		3.93								
		4.49								
40		4.43								
		4.64								
		5.03								
45										

Table 3A Cont'd
ΔCt values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	CT3	1.76 0.15 1.62 0.71 1.06 1.23 1.76 1.87	0.17	---	1.28	1.49 -0.24	---	-0.01	---	0.07
15										
20	CT8	1.17 0.53 1.06 1.95 1.55 1.38 1.39 1.30	0.16	---	0.95	0.67 -0.06	---	0.08	---	-0.11
25										
30	CT10	1.92 1.28 -0.71 2.43 2.04 2.55 2.98 2.70	0.16	---	1.34	0.65 -0.62	---	-0.11	---	-0.05
35										
40	CT12	1.07 0.86 1.47 2.57 2.08 1.77 1.40 1.29	0.21	---	0.91	0.93 -0.07	---	0.17	---	0.10
45										

Table 3A Cont'd
ΔCt values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	CT14	1.68 1.27 2.20 2.52 2.38 2.53 2.45 2.36	0.70	---	1.29	0.72 -0.71	---	0.25	---	0.35
15										
20	CT15	1.51 0.60 -0.67 2.83 2.41 2.30 1.08	0.36	---	1.10	1.00 -0.50	2.61	0.07	---	---
25										
30	CT16	0.65 -0.90 1.02 2.53 2.17 2.10 0.09	0.50	---	1.35	1.02 -0.11	2.21	0.01	---	---
35										
40	CT17	0.93 -0.01 0.75 2.83 2.97 2.78 0.37	0.95	---	1.26	1.23 -0.10	3.31	-0.17	---	---
45										

Table 3A Cont'd
ΔCt values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	CT1	1.60 0.54 -0.21 -0.70 0.73 0.94 1.93 1.62	0.78	---	0.64	1.12 -1.53	---	-0.01	---	0.24
15										
20	CT4	2.68 2.19 1.42 2.11 2.01 2.22 2.67 2.40	0.76	---	1.03	1.25 -1.25	---	-0.04	---	0.19
25										
30	CT5	2.63 1.87 0.84 1.10 1.68 2.11	0.83	---	0.99	1.34 -0.61	---	---	---	---
35										
40	CT6	0.91 1.66 0.51 1.12 0.86 1.19	0.72	---	1.00	1.06 -2.42	---	---	---	---
45										

Table 3A Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	CT7	-0.18 0.41 0.51 0.74 0.79 1.38	0.17	---	0.55	0.88 -1.98	---	---	---	---
15	CT9	1.43 1.21 -1.12 0.20 -0.10 0.20 2.29 1.46	0.52	---	0.82	0.61 -1.49	---	0.09	---	0.21
20										
25	CT11	1.64 1.13 1.14 1.13 1.30 2.03	1.16	---	1.25	1.80 -1.13	---	---	---	---
30										
35	CT18	0.89 -0.96 -0.42 -0.91 -2.46 0.58	0.56	---	0.59	0.69 -1.43	---	---	---	---
40	A549	-0.92 -1.45	---	---	---	---	1.66	0.23	---	---
	Calu-1	0.03	---	---	---	---	1.77	---	---	---
45	Calu-6	-0.17	---	---	---	---	0.91	---	---	---

Table 3A Cont'd
ΔCt values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	H157	0.05	---	---	---	---	0.89	---	---	---
	H441	0.13	---	---	---	---	2.57	---	---	---
	H460	-1.91	---	---	---	---	2.50	---	---	---
15	SKMES1	-0.72	---	---	---	---	2.15	---	---	---
	SW900	-0.56	---	---	---	---	---	---	---	---
20	SW480	2.17	---	---	---	---	---	---	---	---
	SW620	1.04	---	---	---	---	2.36	---	---	---
	Colo320	-1.11 0.85	---	---	---	---	1.99 2.73	0.38	---	---
25	HT29	---	---	---	---	---	1.71	---	---	---
	HM7	-0.05	---	---	---	---	---	---	---	---
30	WiDr	1.21	---	---	---	---	---	---	---	---
	HCT116	2.02 0.38	---	---	---	---	1.90	-0.19	---	---
35	SKCO1	---	---	---	---	---	3.13	---	---	---
	SW403	0.93	---	---	---	---	2.09	---	---	---
40	LS174T	1.24	---	---	---	---	2.90	---	---	---

Table 3A Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO212	PRO290	PRO341	PRO535	PRO619	PRO717	PRO809	PRO830	PRO848
10	LT22	0.00	---	---	---	---	---	---	---	---
		-0.45	---	---	---	---	---	---	---	---
		-0.91	---	---	---	---	---	---	---	---
		-0.92	---	---	---	---	---	---	---	---
15	LT8	1.66	---	---	---	---	---	---	---	---
		0.53	---	---	---	---	---	---	---	---
20	Colo205	---	---	---	---	---	1.48	---	---	---
		---	---	---	---	---	1.46	---	---	---
		---	---	---	---	---	1.20	---	---	---
		---	---	---	---	---	1.67	---	---	---
25	H522	---	---	---	---	---	3.78	---	---	---
		---	---	---	---	---	1.84	---	---	---
	H810	---	---	---	---	---	---	---	---	---

Table 3B
ΔCt values in lung and colon primary tumors and cell line models

Primary Tumor	PRO943	PRO1005	PRO1009	PRO1025	PRO1030	PRO1097	PRO1107	PRO1111	PRO1153
5									
LT-1	-0.01 -2.67	1.07	---	0.82 0.03	0.62	0.07	0.21	0.34	0.48
10									
LT-1a	0.84 -1.01	3.87	---	0.85 -0.26	0.75	-0.17	-0.22	0.47	0.76
15									
LT2	-0.34 -1.33	---	---	0.79 -0.20	0.07	1.23	0.09	0.61	0.83
LT3	0.95 0.38	1.61	---	1.40 0.15	0.78	0.07	-0.19	1.39	0.22
20									
LT4	0.48 0.20	---	---	0.51 -0.14	1.79	0.83	-0.16	1.49	1.01
LT6	-0.52 -1.52	1.29	---	1.16 -0.49	0.82	0.04	-0.35	0.43	-0.06
25									
LT7	0.47 -0.45	0.00	---	1.13 0.10	1.09	0.50	0.35	1.58	1.52
30									
LT9	0.61 -0.05	2.50	---	2.05 -0.38	1.07	1.21	-0.65	1.44	0.26
LT10	0.95 0.24	0.23	---	1.07 0.33	1.15	0.90	-0.10	1.05	0.98
35									
LT11	2.06 0.84	---	---	0.41 0.09	0.76	---	---	1.45	---
LT12	1.94 0.62	1.21	---	0.71 0.25	0.23	---	---	0.52	---
40									
LT13	1.64 1.27	2.30	0.21	0.66 0.06	0.63	---	3.55	0.55	0.24
45									
LT15	2.05 0.81	1.03	0.08	0.53 -0.37	0.88	---	2.47	0.44	-0.09

Table 3B Cont'd
 ΔC_t values in lung and colon primary tumors and cell line models

Primary Tumor	PRO943	PRO1005	PRO1009	PRO1025	PRO1030	PRO1097	PRO1107	PRO1111	PRO1153
5									
LT16	0.49 0.27	1.05	0.00	0.53 -0.96	0.46	---	2.45	0.74	0.01
10									
LT17	1.93 0.95	---	---	0.52 0.05	0.81	---	---	1.47	---
15									
LT18	-0.09 -0.65	---	---	-0.24 0.06	0.10	---	---	0.84	---
15									
LT19	2.90 -0.91	---	---	0.52 0.42	0.64	---	---	---	---
20									
LT21	0.96	---	---	0.67	0.70	---	---	---	---
20									
CT2	1.92 1.70	---	2.00 0.04	1.88 1.11	1.03	0.13	4.75	0.30	0.75
25									
CT3	0.31 0.04	---	1.75 -0.18	1.77 0.54	1.75	0.43	1.52	0.90	0.33
30									
CT8	1.37 1.12	---	1.29 -0.07	0.81 -0.26	0.82	0.43	0.60	0.62	-0.21
35									
CT10	2.13 1.67	---	1.73 0.21	1.28 -0.05	1.19	0.95	2.82	0.56	0.09
40									
CT12	1.43 0.40	---	1.92 0.23	0.90 0.34	1.28	0.49	0.93	0.87	-0.14
35									
CT14	1.46 0.56	---	2.10 0.42	1.58 0.34	1.92	1.08	1.54	1.38	0.06
40									
CT15	0.72 0.43	-0.71	2.02 -0.37	0.73 -0.45	1.46	0.55	0.48	0.30	0.24
45									
CT16	-0.26 0.13	-0.95	1.56 0.02	0.78 -0.36	1.57	1.11	0.36	0.74	0.09

Table 3B Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

Primary Tumor	PRO943	PRO1005	PRO1009	PRO1025	PRO1030	PRO1097	PRO1107	PRO1111	PRO1153
5									
CT17	1.30 0.99	-0.78	1.76 -0.30	0.50 -0.19	1.64	1.34	0.80	0.83	-0.48
10									
CT1	0.79 1.36	---	0.89 -0.14	1.11 0.63	0.78	-0.89	1.57	0.36	-0.21
CT4	-1.34 -0.43	---	1.06 -0.21	1.46 0.75	0.90	-0.39	1.59	0.43	0.02
15									
CT5	1.88 2.51	---	1.43	0.73 -0.12	1.05	-1.29	---	0.33	---
CT6	1.41 1.75	---	0.79	0.48 -0.20	0.57	-0.91	---	0.19	---
20									
CT7	-0.98 -0.48	---	0.74	1.00 0.55	0.47	-0.69	---	1.16	---
CT9	0.16 0.15	---	-0.18 0.07	1.13 -0.46	0.93	-1.27	0.44	0.85	0.19
25									
CT11	2.80 2.61	---	1.83	0.71 -0.44	0.94	0.19	---	1.17	---
30									
CT18	1.30 0.96	---	0.32	0.73 0.01	0.63	-0.47	---	1.05	---
35									
A549	---	0.23	0.21	---	---	---	0.34	---	0.24
Calu-1	---	---	---	---	---	---	---	---	---
Calu-6	---	---	---	---	---	---	---	---	---
40									
H157	---	---	---	---	---	---	---	---	---
H441	---	---	---	---	---	---	---	---	---

Table 3B Cont'd
 ΔC_t values in lung and colon primary tumors and cell line models

Primary Tumor	PRO943	PRO1005	PRO1009	PRO1025	PRO1030	PRO1097	PRO1107	PRO1111	PRO1133
5 H460	---	---	---	---	---	---	---	---	---
10 SKMES1	---	---	---	---	---	---	---	---	---
SW900	---	---	---	---	---	---	---	---	---
SW480	---	---	---	---	---	---	---	---	---
15 SW620	---	---	---	---	---	---	---	---	---
Colo320	---	-0.51	0.32	---	---	---	0.93	---	0.10
20 HT29	---	---	---	---	---	---	---	---	---
HM7	---	---	---	---	---	---	---	---	---
WiDr	---	---	---	---	---	---	---	---	---
25 HCT116	---	-2.48	-0.47	---	---	---	-0.73	---	-0.49
SKCO1	---	---	---	---	---	---	---	---	---
SW403	---	---	---	---	---	---	---	---	---
30 LS174T	---	---	---	---	---	---	---	---	---
LT22	---	---	---	---	---	---	---	0.82	---
35 LT8	---	0.48	---	---	---	---	---	0.82	---
Colo205	---	---	---	---	---	---	---	---	---
HCT15	---	---	---	---	---	---	---	---	---
40 HCC2998	---	---	---	---	---	---	---	---	---
KM12	---	---	---	---	---	---	---	---	---

Table 3B Cont'd
ΔCt values in lung and colon primary tumors and cell line models

5	Primary Tumor	PRO943	PRO1005	PRO1009	PRO1025	PRO1030	PRO1097	PRO1107	PRO1111	PRO1153
	H522	---	---	---	---	---	---	---	---	---
10	H810	---	---	---	---	---	---	---	---	---
15										
20										
25										
30										
35										
40										
45										

Table 3C
 Δ Ct values in lung and colon primary tumors and cell line models

Primary Tumor	PRO1182	PRO1184	PRO1187	PRO1281	PRO23	PRO39	PRO834	PRO1317	PRO1710
5	LT-1	1.81 0.50 0.42 -0.29	-0.88	---	0.02	0.78	-0.09	1.15	-1.78
10	LT-1a	0.73 1.14 1.09 0.26	-0.03	---	0.55	1.33	1.20	1.49	-0.89
15	LT2	-0.07 -0.32 -0.40 -0.88	-0.14	---	-0.06	1.09	0.27	0.83	-0.18
20	LT3	0.70 0.58 0.55 -0.42	0.28	---	0.92	-0.17	0.56	0.67	-0.17
25	LT4	1.43 1.37 1.18 -0.05	-0.36	---	0.48	1.01	0.14	0.98	-1.43
30	LT6	0.04 1.78 1.66 1.05	-0.69	---	0.73	0.59	0.66	0.93	-0.35
35	LT7	0.47 0.58 0.40 -0.57	-0.07	---	0.13	-0.30	0.65	0.85	-0.93
40	LT9	1.43 0.43 0.58 -0.34	-0.39	---	0.13	0.87	0.75	1.26	-1.20
	LT10	0.91 0.62 0.60 0.01	-0.51	---	0.56	-10.46	0.63	1.68	-0.32

Table 3C Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

Primary Tumor	PRO1182	PRO1184	PRO1187	PRO1281	PRO23	PRO39	PRO834	PRO1317	PRO1710
5									
LT11	0.62	0.73 0.96 -0.28	0.49	---	0.52	0.27 -0.43	---	-0.44 -0.37	0.61 0.34
10									
LT12	0.43	2.47 2.61 1.80	1.17	---	0.65	2.57 2.29	---	-0.13 0.01	1.27 1.26
15									
LT13	0.34	0.54 0.71 -0.73 -0.38	0.92	---	1.43	2.22 2.26	---	0.54 -0.51	2.22 2.53
20									
LT15	0.20	0.46 0.67 -0.41 0.61	1.55	---	2.41	3.24 3.04	---	1.16 -0.84	3.78 3.52
25									
LT16	0.42	1.01 1.07 -0.31 0.18	1.33	---	1.64	3.06 2.70	---	0.97 0.41	3.84 3.55
30									
LT17	0.53	0.67 0.87 -0.67	0.57	---	0.96	-0.21 -0.99	---	1.74 -0.26	-0.72 -0.73
35									
LT18	0.46	1.07 1.13 0.18	0.48	---	0.81	-0.92 -1.43	---	0.86 -0.12	-0.90 -0.34
40									
LT19	0.78	1.19 1.35 1.02	---	---	---	---	---	---	---
LT21	0.60	1.00 1.20	---	---	---	---	---	---	---

Table 3C Cont'd
ΔCt values in lung and colon primary tumors and cell line models

Primary Tumor	PRO1182	PRO1184	PRO1187	PRO1281	PRO23	PRO39	PRO834	PRO1317	PRO1710
5									
CT2	0.09	0.43 0.41	---	1.15	-0.06	3.45	2.66	0.03	4.95
10									
CT3	-0.21	0.44 0.22	---	0.46	-0.27	0.15	0.74	0.27	0.17
15									
CT8	-0.48	0.32 -0.03	---	0.01	0.08	1.12	1.13	0.05	1.34
CT10	-0.04	0.57 0.32	---	-0.01	0.17	2.15	1.89	-0.07	1.94
20									
CT12	-0.45	0.10 -0.18	---	1.07	0.66	0.86	1.56	-0.44	0.43
CT14	-0.04	-0.28 -0.51	---	-0.05	0.88	2.58	1.96	0.60	1.56
25									
CT15	---	0.49 0.07	---	---	---	---	1.88	---	---
30									
CT16	---	-0.28 -0.48	---	---	0.45	0.77	1.34	0.29	1.19
CT17	---	-0.08 -0.30	---	---	1.35	0.86	0.96	-0.44	0.78
35									
CT1	-0.21	0.49 0.08	---	-0.76	1.06	0.58	1.53	-0.12	0.85
CT4	0.26	-0.02 -0.16	---	0.98	1.22	1.18	1.19	-0.06	2.28
40									
CT5	---	-0.12	---	---	1.87	2.70	1.63	-0.31	2.67
CT6	---	-0.20	---	---	0.63	-0.35	1.85	-0.13	-1.38

Table 3C Cont'd
 Δ Ct values in lung and colon primary tumors and cell line models

Primary Tumor	PRO1182	PRO1184	PRO1187	PRO1281	PRO23	PRO39	PRO834	PRO1317	PRO1710
5 CT7	---	-0.08	---	---	1.56	1.02	0.88	0.47	1.61
10 CT9	-0.09	-0.17 -0.26	---	-0.01	-0.45	-1.60	0.32	-0.25	-0.68
CT11	---	-0.25	---	---	1.40	2.25	2.72	-0.54	2.17
15 CT18	---	-0.15	---	---	0.63	-0.61	0.92	-0.06	-0.58
A549	---	0.26	---	---	---	---	---	---	---
Calu-1	---	---	---	---	0.24	---	---	---	1.49
20 Calu-6	---	---	---	---	0.11	---	---	---	0.12
H157	---	---	---	---	-0.08	---	---	---	0.61
25 H441	---	---	---	---	1.19	---	---	---	0.07
H460	---	---	---	---	0.85	---	---	---	0.50
30 SKMES1	---	---	---	---	0.30	---	---	---	0.89
SW900	---	---	---	---	---	---	---	---	---
SW480	---	---	---	---	---	---	---	---	---
35 SW620	---	---	---	---	-0.04	---	---	---	2.22
Colo320	---	-0.20	---	---	1.90	1.02	---	---	---
40 HT29	---	---	---	---	1.76	2.43	---	---	---
HM7	---	---	---	---	---	---	---	---	---
WiDr	---	---	---	---	---	---	---	---	---

Table 3C Cont'd
 ΔC_t values in lung and colon primary tumors and cell line models

Primary Tumor	PRO1182	PRO1184	PRO1187	PRO1281	PRO23	PRO39	PRO834	PRO1317	PRO1710
5 HCT116	---	-0.36	---	---	---	---	---	---	---
10 SKCO1	---	---	---	---	1.60	1.28	---	---	---
SW403	---	---	---	---	3.13	1.29	---	---	---
LS174T	---	---	---	---	2.43	1.05	---	---	---
15 LT22	---	---	-0.99	---	-0.87	-0.64 -1.63	---	2.69 0.70	-0.52 -0.97
LT8	---	---	---	---	---	---	---	---	---
20 Colo205	---	---	---	---	0.66	-0.11	---	---	---
HCT15	---	---	---	---	3.11	0.48	---	---	---
25 HCC2998	---	---	---	---	1.44	0.74	---	---	---
KM12	---	---	---	---	1.18	0.17	---	---	---
H522	---	---	---	---	0.74	---	---	---	-0.18
30 H810	---	---	---	---	0.93	---	---	---	-0.21

35

40

Table 3D
 ΔC_t values in lung and colon primary tumors and cell line models

Primary Tumor	PRO2094	PRO2145	PRO2198
5 LT-1	0.74	1.11	0.44
10 LT-1a	0.83	1.11	1.32
LT2	---	---	---
LT3	0.95	1.23	0.67
15 LT4	---	---	---
LT6	0.90	0.36	-0.12
LT7	2.10	1.28	0.26
20 LT9	1.13	3.29	-0.19
LT10	-1.24	0.40	0.04
25 LT11	---	---	---
LT12	-1.64	3.22	-0.11
30 LT13	0.41	2.40	2.92
			-0.48
			0.34
			-0.09
			0.43
			1.12
			-0.04
			-0.22
35			
40			

Table 3D Cont'd
ΔCt values in lung and colon primary tumors and cell line models

Primary Tumor	PRO2094	PRO2145	PRO2198
5			
LT15	1.48	1.26	0.16
			0.84
			0.44
10			-0.09
			0.31
			0.36
			0.48
			0.56
15			
LT16	-1.00	0.54	1.69
			0.42
			0.62
20			-0.25
			0.47
			0.79
			0.27
			-0.33
25			
LT17	---	---	---
LT18	---	---	---
LT19	---	---	---
30			
LT21	---	---	---
CT2	-1.37	---	2.27
35			
CT3	-1.38	---	0.45
CT8	1.20	---	0.43
CT10	0.11	---	1.26
40			
CT12	-8.76	---	0.13
CT14	1.38	---	1.46

Table 3D Cont'd
ΔCt values in lung and colon primary tumors and cell line models

Primary Tumor	PRO2094	PRO2145	PRO2198
5	CT15	1.16	0.51
			0.78
			-0.23
			0.21
			-0.40
			0.67
			1.43
			0.21
			0.05
10			
			0.74
			-0.23
			1.31
			-0.22
			0.24
			2.58
			0.08
			-0.05
15			
	CT16	0.30	0.28
			0.49
			2.33
			1.50
			-0.58
			0.41
			0.85
			-0.01
			-0.12
			0.90
20			
			0.97

			0.41
25	CT17	1.12	0.20
30			
	CT1	1.70	---
	CT4	1.34	---
	CT5	---	---
	CT6	---	---
	CT7	---	---
	CT9	1.33	---

Table 3D Cont'd
ΔCt values in lung and colon primary tumors and cell line models

Primary Tumor	PRO2094	PRO2145	PRO2198
5 CT11	---	---	---
CT18	---	---	---
10 A549	-11.06	2.41	1.49
			-13.07
			1.31
			-0.59
			-0.27
15			1.75
			0.27
			-0.04
20 Calu-1	---	---	0.36
Calu-6	---	---	0.56
H157	---	---	0.84
25 H441	---	---	0.78
H460	---	---	0.52
30 SKMES1	---	---	0.31
SW900	---	---	---
SW480	---	---	---
35 SW620	---	---	0.59

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Table 3D Cont'd
ΔCt values in lung and colon primary tumors and cell line models

	Primary Tumor	PRO2094	PRO2145	PRO2198
5	Colo320	-1.33	0.31	2.10 0.86 -0.81 -0.58 -0.11 0.09 0.46 -0.13
10				
15	HT29	---	---	---
	HM7	---	---	---
20	WiDr	---	---	---
	HCT116	0.86	1.03	1.32 1.60 0.39 -0.10 0.55 1.89 -0.21 -0.39
25				
30	SKCO1	---	---	---
	SW403	---	---	---
35	LS174T	---	---	---
	LT22	---	---	---
	LT8	0.66	1.35	0.92
40	Colo205	---	---	---

Table 3D Cont'd
ΔCt values in lung and colon primary tumors and cell line models

Primary Tumor	PRO2094	PRO2145	PRO2198
5 HCT15	---	---	---
10 HCC2998	---	---	---
KM12	---	---	---
H522	---	---	0.45
15 H810	---	---	-0.72

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PRO717:

PRO717 (UNQ385) (DNA50988-1326) was also reexamined along with selected tumors from the above initial screen with framework mapping. Figure 63 and Table 4 indicate the chromosomal mapping of the framework markers that were used in the present example. The framework markers are located approximately every 20 megabases along Chromosome 4, and are used to control aneuploidy.

PRO717 (UNQ385) (DNA50988-1326) was also reexamined with epicenter mapping. The epicenter markers indicated in Table 5 are located in close proximity (in the genome) to DNA50988, and are used to assess the relative amplification in the immediate vicinity of Chromosome 4 wherein the molecule is located. The distance between markers is measured in centirays (cR), which is a radiation breakage unit approximately equal to a 1% chance of a breakage between two markers. One cR is very roughly equivalent to 20 kilobases. The marker WI-13023 is the marker found to be the closest to the location on Chromosome 4 where DNA50988-1326 maps. DNA52192 maps to the same marker (WI-13023) at which DNA50988 maps.

The ΔC_t values of the described framework markers (Table 4) along with Chromosome 4 relative to PRO717 are indicated for selected tumors in Table 6.

Table 7 indicates the ΔC_t values for results of epicenter mapping to DNA50988, indicating the relative amplification in the region more immediate to the actual location of DNA50988 along Chromosome 4.

Table 4
Framework Markers Along Chromosome 4

Map Position on Chromosome 4	Stanford Human Genome Center Marker Name
D8	SHGC4-312
D69	SHGC4-370
D134	SHGC4-666
D186	SHGC-34520
D233	SHGC-10570
D283	WI-9138
D344	SHGC-36904
D397	SHGC-11799
D457	SHGC-23843
D502	CdyOce04
D561	SHGC4-1073
D611	SHGC4-359
D667	FB12A4
D705	W88774

Table 5
Epicenter Markers Along Chromosome 4 used for DNA50988

	Map Position on Chromosome 4	Stanford Human Genome Center Marker Name	Distance to Next Marker (cR)
5	D2	WI-15821	4
	D3	SHGC-32625	4
	D4	SHGC4-207	19
	D5	WI-13023	11
	DNA52192	WI-13023	?
10	D6	SHGC4-224	28
	D7	SHGC4-1127	24
	D8	SHGC4-312	-

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Table 6

Amplification of Framework Markers Relative to DNA50988 (ΔCt)

Tumor	Framework Markers																
	DNA 50988	DNA 52192	D8	D69	D134	D186	D233	D283	D344	D397	D457	D502	D561	D611	D667	D705	
5	CT15	2.97	0.61	-0.18	-0.51	-0.34	-0.36	0.02	-0.50	-0.46	1.23	-0.48	0.00	-0.29	-0.39	-0.48	-0.45
	CT16	3.38	0.20	-0.07	-16.48	-0.23	0.06	0.12	-0.05	0.02	1.97	-0.06	0.00	-0.04	-0.07	0.13	0.07
	CT17	3.97	0.38	0.13	-16.48	0.04	-0.26	0.48	-0.12	-0.09	2.19	-0.01	0.00	-0.30	-0.09	-0.30	-0.10
	LT13	6.43	2.37	0.18	0.07	-0.20	-0.04	4.30	-0.34	0.01	3.18	-0.22	0.00	-0.33	-0.18	-0.12	0.24
10	LT15	3.45	0.38	0.29	0.10	-0.09	-0.40	1.86	-0.04	-0.30	2.67	-0.23	0.00	-0.72	-0.33	-0.16	-0.19
	LT16	5.65	0.77	0.01	-0.29	-0.23	-0.31	4.69	-0.30	-0.27	3.71	-0.32	0.00	-0.81	-0.85	-0.66	-0.64
	SW480	3.00	0.13	-0.07	-0.55	-0.27	-0.33	-0.40	-0.07	-0.37	1.72	-0.47	0.00	-1.00	-0.36	-0.73	-0.29
	HCT 116	2.64	0.45	0.29	0.12	0.18	0.07	1.22	0.13	0.18	1.97	0.05	0.00	-0.52	-0.03	-0.66	0.04
15	A549	1.57	0.18	-0.10	-0.55	-0.32	-0.68	0.60	-0.64	-0.44	1.69	-0.57	0.00	-1.08	-0.63	-0.98	-0.65
	CT1	3.49	0.56	0.43	-0.25	-0.04	0.00	1.41	-0.02	0.10	0.37	0.03	0.00	-0.15	0.14	0.00	-0.13
	CT2	5.65	0.59	0.02	-0.22	-0.25	-0.12	3.43	-0.27	-0.12	1.31	-0.22	0.00	-0.24	-0.20	-0.13	-0.26
	CT3	2.46	0.97	0.13	0.27	0.14	0.07	1.40	0.15	0.17	0.20	0.33	0.00	-0.03	0.06	0.04	0.10
20	CT4	3.49	0.52	-0.02	-0.27	-0.27	-0.33	0.97	-0.59	0.04	0.11	-0.15	0.00	-0.35	-0.03	0.05	-0.08
	CT8	3.35	0.70	0.34	0.22	0.01	0.00	0.80	-0.27	-0.02	0.52	-0.05	0.00	-0.01	0.02	0.14	-0.02

Tumor	Framework Markers															
	DNA 50988	DNA 52192	D8	D69	D134	D186	D233	D283	D344	D397	D457	D502	D561	D611	D667	D705
CT9	2.77	-0.12	0.16	-0.18	-0.07	-0.11	0.69	-0.54	0.03	0.14	-0.04	0.00	0.00	-0.03	0.03	-0.20
CT10	4.59	1.23	0.34	0.14	-0.01	0.13	2.01	-0.27	0.15	0.32	0.24	0.00	0.25	0.04	0.07	-0.05
CT12	2.52	0.73	0.18	-0.11	-0.01	0.25	1.88	-0.22	0.10	0.54	-0.11	0.00	-0.28	-0.18	0.00	-0.24
CT14	3.99	1.09	0.45	-0.20	0.17	0.25	0.50	0.50	-0.30	1.16	-0.50	0.00	-0.72	-0.48	-0.82	-0.82

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Table 7
Amplification of Epicenter Markers Relative to DNA50988 (ΔC_t)

Tumor	D2	D3	D4	D5	DNA 50988	D6	D7	D8
CT15	-0.16	-0.25	-0.21	-0.56	2.25	-1.14	-0.35	0.31
CT16	-0.50	-0.67	-0.78	-0.88	1.32	-0.77	-1.05	-0.59
CT17	0.18	0.07	-0.10	0.00	2.60	0.88	-0.18	0.04
LT13	-0.50	-0.69	-0.83	0.00	6.70	0.09	-0.89	0.25
LT15	0.19	0.05	0.38	0.10	2.71	0.04	0.40	0.34
LT16	-0.20	-0.36	-0.34	0.00	5.44	0.23	-0.30	0.09
SW480	-0.36	-0.54	-0.91	-0.25	2.22	0.20	-0.06	0.40

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Tumor	D2	D3	D4	D5	DNA 50988	D6	D7	D8
HCT116	-0.15	-0.27	-0.21	-0.09	2.50	-0.50	-0.09	0.09
A549	-0.54	-0.83	-0.57	-0.56	1.98	1.37	-0.43	-0.11
CT1	0.54	0.52	0.56	0.56	4.54	0.00	0.69	0.75
CT2	-0.09	-0.01	0.05	-0.06	6.33	0.00	0.10	0.16
CT3	-0.34	-0.45	-0.45	-0.43	2.50	0.00	-0.28	-0.32
CT4	-0.19	-0.04	0.18	-0.02	4.10	0.00	0.26	0.34
CT8	0.34	0.34	0.70	0.49	4.42	0.00	0.81	0.75
CT9	0.16	0.14	0.52	0.20	3.96	0.00	0.46	0.41
CT10	0.69	0.73	0.86	0.72	5.62	0.00	0.94	0.75
CT12	0.75	0.70	0.70	0.58	4.14	0.00	0.80	0.70
CT14	0.54	0.57	0.18	0.74	6.23	0.00	0.90	0.90

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PRO1005:

PRO1005 (UNQ489) (DNA57708) was also reexamined with epicenter mapping. The epicenter markers indicated in Table 8 are located in close proximity (in the genome) to DNA57708, and are used to assess the relative amplification in the immediate vicinity of Chromosome 2 wherein the molecule is located.

- 5 The distance between individual markers is measured in centirays (cR), which is a radiation breakage unit approximately equal to a 1% chance of breakage between two markers. One cR is very roughly equivalent to 20 kilobases. The marker SHGC-30955 is the marker found to be the closest to the location on Chromosome 2 where DNA57708 maps.

- 10 Table 9 indicates the ΔC_t values for results of epicenter mapping relative to DNA57708, indicating the relative amplification in the region more immediate to the actual location of DNA57708 along Chromosome 2.

Table 8
Epicenter Markers Along Chromosome 2 Used for DNA57708

15	Map Position on Chromosome 2	Stanford Human Genome Center Marker Name	Distance to next Marker (cR)
	B140	SHGC-35027	19
	B141	SHGC-30955	7
	B142	SHGC-33686	7
20	B143	CHLC.GATA72A05	7
	B144	SHGC-11271	24
	B145	AFM154xf8	104
	B149	SHGC-31078	-

Table 9
Amplification of Epicenter Markers Relative to DNA57129 (ΔC_t)

	Tumor	B140	B141	DNA 57708	B143	B144	B145	B149
	HF613	-0.15	0.09	-0.43	0.11	0.00	0.44	0.03
	HF854	-0.19	0.09	0.11	0.28	-0.07	-0.34	-0.01
30	HF545	-0.09	-0.20	1.60	-0.04	-0.12	-0.74	-0.73
	HF539	0.10	-0.04	1.29	0.51	0.09	-0.26	0.02
	HF575	-0.18	0.04	0.83	0.28	0.00	-0.10	-0.09
	HF499	-0.01	0.11	0.67	0.47	0.04	-0.13	0.10
	HF733	0.48	0.57	4.61	0.65	0.47	-0.04	0.23

Tumor	B140	B141	DNA 57708	B143	B144	B145	B149
HF716	-0.14	-0.28	3.67	0.79	0.09	-1.37	-0.56

PRO2094 (DNA83123):

5 PRO2094 (DNA83123) was also reexamined along with selected tumors from the above initial screen with framework mapping. Figure 65 and Table 10 indicate the chromosomal mapping of the framework markers that were used in the present example. The framework markers are located approximately every 20 megabases along Chromosome 6, and are used to control aneuploidy.

10 PRO2094 (DNA83123) was also reexamined with epicenter mapping. The epicenter markers indicated in Table 11 are located in close proximity (in the genome) to DNA83123, and are used to assess the relative amplification in the immediate vicinity of Chromosome 6 wherein the molecule is located. The distance between markers is measured in centirays (cR), which is a radiation breakage unit approximately equal to a 1% of a breakage between two markers. One cR is very roughly equivalent to 20 kilobases. The marker SHGC-1259 is the marker found to be the closest to the location on Chromosome 6 where DNA83123 maps. DNA77568 maps
15 to the same marker (SHGC-1259) at which DNA83123 maps.

The ΔC_t values of the described framework markers (Table 10) along Chromosome 6 relative to PRO2094 is indicated for selected tumors in Table 12.

Table 13 indicates the ΔC_t values for results of epicenter mapping relative to DNA83123, indicating the relative amplification in the region more immediate to the actual location of DNA83123 along Chromosome
20 6.

Table 10
Framework Markers Along Chromosome 6

Map Position on Chromosome 6	Stanford Human Genome Center Marker Name
F1	SHGC-37322
F38	AFM242za5
F84	SHGC-11076
F136	SHGC-11518
F186	SHGC-10380
F228	SHGC-35377
F259	SHGC-10217
F307	SHGC-15962
F348	SHGC-36574
F394	SHGC-34515

Table 11
Epicenter Markers Along Chromosome 6 Used for DNA83123

	Map Position on Chromosome 6	Stanford Human Genome Center Marker Name	Distance to Next Marker (cR)
5	F146	SHGC-9402	46
	F147	SHGC-3143	14
	F148	SHGC-1259	3
	DNA77568	SHGC-1259	?
	F149	AFM301wb9	28
10	F150	GGAT4C01	-

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Table 12

Amplification of Framework Markers Relative to DNA83123 (Δ Ct)

Tumor	Framework Markers									
	F1	F38	DNA 83123	DNA 77568	F84	F228	F259	F307	F348	F394
CT15	-0.02	0.03	1.99	-0.23	0.41	0.31	0.02	-0.23	0.21	0.35
CT16	0.46	0.16	1.11	0.57	0.68	0.42	0.31	0.23	0.46	0.48
CT17	-0.32	-0.59	1.31	-0.20	-0.18	-0.24	-0.57	-0.52	-0.24	-0.25
LT13	0.94	0.24	1.36	-0.13	0.66	0.70	0.32	0.54	1.81	0.54
LT15	0.43	0.11	1.42	0.37	0.34	0.59	0.47	0.54	1.20	0.50
LT16	0.65	-0.16	-0.04	-0.22	0.04	0.09	0.04	0.21	0.74	-0.02
SW480	-0.64	-0.78	1.56	-0.66	-0.09	0.29	-0.31	-0.29	-0.25	-0.24
HCT 116	0.64	0.32	-0.13	-0.22	0.41	0.71	0.70	0.38	0.68	0.60
A549	-0.31	-0.40	-12.69	-1.08	0.29	0.26	-0.11	0.02	0.02	-0.09
Colo 320	0.66	-0.44	1.49	-0.74	0.53	-0.02	-0.05	-	-9.73	-
Colo205	-0.20	-0.17	-12.89	0.13	-0.22	-0.77	-1.21	-	-1.33	-
HCT75	-0.20	-0.06	1.14	0.03	0.15	-0.23	-0.30	-	-0.56	-

Tumor	Framework Markers									
	F1	F38	DNA 83123	DNA 77568	F84	F228	F259	F307	F348	F394
HCC 2998	0.42	-0.16	1.22	-0.23	0.45	-0.20	-0.42	-	-0.82	-
CT29	-0.35	-0.88	-14.11	0.36	0.09	-0.32	-0.73	-	0.08	-
KM12	-0.18	-1.57	1.30	-0.34	-1.27	-0.28	-0.55	-	-0.70	-
Ls174T	-0.03	-1.54	-0.54	-2.83	-0.52	0.08	-0.67	-	-1.65	-
SKCO1	0.12	-0.53	1.14	-0.32	-0.36	-0.50	-0.76	-	-0.68	-
SW403	-0.53	-0.20	-0.22	0.38	0.48	-0.13	-0.50	-	-0.18	-
SW620	0.22	-0.02	1.40	-1.35	-0.23	-0.02	-0.13	-	-1.01	-
H522	0.11	0.34	-12.43	0.25	-0.88	0.05	0.04	-	-0.84	-
H460	-0.07	-0.41	-12.33	-1.03	-0.29	-0.70	-1.01	-	-0.50	-
SKMES1	-0.05	-0.44	-2.94	-0.27	-0.48	-0.61	-0.43	-	-0.37	-
Calu-1	-0.71	-0.11	0.23	0.10	0.48	-0.07	-0.22	-	-0.37	-
Calu-6	-1.02	-0.07	1.50	-0.12	0.39	-0.70	-0.57	-	-0.68	-
H157	-0.75	0.38	1.50	-0.80	0.25	-0.65	-0.79	-	-1.02	-
H441	-0.68	-0.21	1.04	-0.35	-0.84	-0.77	-1.03	-	-0.52	-
H810	-0.36	-0.63	-0.22	-0.32	-0.77	-0.82	-1.44	-	-0.93	-

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Table 13

Amplification of Epicenter Markers Relative to DNA83123 (Δ Ct)

Tumor	F150	F149	DNA 83123	F148	F147	F146	F144
Kidney 613	0.00	0.00	1.21	-0.18	-0.08	0.07	-0.57
Lymphoma 854	0.00	0.00	0.58	-0.26	-0.93	-0.56	-0.47
Breast 545	0.00	0.00	-0.41	-0.51	-0.19	0.22	-0.13
Colon 539	0.00	0.00	-0.27	0.02	0.24	0.13	-0.29
Colon 575	0.00	0.00	1.22	-0.16	-0.19	0.05	0.10
Colon 698	0.00	0.00	0.00	-0.34	-0.28	0.22	-0.27
Colon 499	0.00	0.00	0.35	0.25	-0.10	-0.08	-0.59
Testes 733	0.00	0.00	1.61	-0.52	-0.03	0.40	-0.15
Testes 716	0.00	0.00	0.92	-0.88	-0.14	-0.07	-0.75

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PRO2198:

PRO2198(DNA88153) was also reexamined along with selected tumors from the above initial screen with framework mapping. Figure 66 and Table 14 indicate the chromosomal mapping of the framework markers that were used in the present example. The framework markers are located approximately every 20 megabases along Chromosome 16, and are used to control aneuploidy.

PRO2198 (DNA88153) was also reexamined with epicenter mapping. The epicenter markers indicated in Table 15 are located in close proximity (in the genome) to DNA88153, and are used to assess the relative amplification in the immediate vicinity of Chromosome 16 wherein the molecule is located. The distance between markers is measured in centirays (cR), which is a radiation breakage unit approximately equal to a 1% chance of a breakage between two markers. One cR is very roughly equivalent to 20 kilobases. The marker AFM156xb8 is the marker found to be the closest to the location on Chromosome 16 where DNA88153 maps. DNA62377 maps to the same marker (AFM156xb8) at which DNA88153 maps.

The ΔC_t values of the described framework markers (Table 14) along Chromosome 16 relative to PRO2198 is indicated for selected tumors in Table 16.

Table 17 indicates the ΔC_t values for results of epicenter mapping relative to DNA88153, indicating the relative amplification in the region more immediate to the actual location of DNA88153 along Chromosome 16.

Table 14
Framework Markers Along Chromosome 16

Map Position on Chromosome 16	Stanford Human Genome Center Marker Name
P7	SHGC-2835
P19	SHGC-6150
P55	SHGC-9643
P99	GATA7B02
P154	SHGC-33727
P208	SHGC-13574

Table 15
Epicenter Markers Along Chromosome 16 used for DNA88153

Map Position on Chromosome 16	Stanford Human Genome Center Marker Name	Distance to Next Marker (cR)
P210	SHGC-32044	19
P211	HUM00KMB07	7
P212	AFM156xb8	-
DNA62377	AFM156xb8	?

Table 16
Amplification of Framework Markers Relative to DNA88153 (Δ Ct)

	Tumor	Framework Markers						
		P7	P19	P55	P99	P154	P208	DNA 88153
5								DNA 62377
	Colo320	-0.07	-0.08	-0.04	-0.07	-0.48	0.33	0.97
	Colo205	-0.61	-0.02	-0.26	-0.20	-0.52	0.45	0.09
	HCT15	0.26	-0.12	-0.40	-0.02	-0.25	0.10	0.29
10	HCC 2998	2.72	-0.41	-0.56	-0.43	-0.87	-0.56	2.67
	HT-29	2.12	0.03	-0.20	-0.61	-0.51	-0.54	-0.05
	KM12	1.03	-0.58	-0.71	-1.09	-1.14	-0.49	1.44
	LS174T	-1.75	-0.67	-0.54	-0.53	-0.55	-0.51	0.84
	SKC0-1	-0.97	-0.58	-0.29	-0.03	-0.48	0.43	2.14
15	SW403	-0.40	-0.19	0.02	-0.15	-0.18	0.25	0.90
	CT15	-0.14	-0.28	-0.34	-0.48	-0.05	-0.79	1.04
	CT16	-0.76	-0.40	-0.26	-0.65	-0.75	-0.77	0.30
	CT17	-0.34	-0.37	-0.01	-0.12	0.04	-0.15	0.21
	LT13	0.90	-0.39	-0.19	-0.20	-0.48	-0.61	2.75
20	LT15	0.63	0.01	-0.08	-1.22	-0.47	-0.77	-0.13
	LT16	0.93	-0.60	-0.39	-0.71	-0.55	-0.90	1.05
	SW480	-2.10	-0.63	-0.31	-0.45	-0.16	-0.36	1.19
	HCT116	-0.70	-0.49	-0.29	-0.26	-0.29	-0.31	1.95
	A459	-1.64	-0.71	-0.09	-0.39	-0.04	-0.25	1.10
25	SW620	1.22	-0.25	-0.07	0.10	1.17	-0.05	0.60
	H522	1.59	0.32	0.43	0.28	0.30	-0.16	0.27
	H460	1.34	-0.20	0.01	-0.16	0.57	-0.29	0.34
	SKMES-1	1.57	0.02	0.20	-0.15	0.65	-0.42	-0.06
	Calu-1	1.69	-0.04	-0.08	-1.01	-0.45	-0.53	0.53
30	Calu-6	1.11	0.00	0.14	0.06	0.22	-0.19	0.55
	H157	0.31	-0.36	-0.24	-0.01	0.50	0.38	0.87
	H441	2.04	-0.15	-0.06	0.00	0.04	-0.19	0.81
								-1.59

Tumor	Framework Markers							
	P7	P19	P55	P99	P154	P208	DNA 88153	DNA 62377
H810	-0.32	-0.59	-0.62	-0.89	-0.50	-0.57	-0.20	0.10

5

Table 17

Amplification of Epicenter Markers Relative to DNA88153 (Δ Ct)

	Tumor	P210	P211	P212	DNA 88153	DNA 62377
10	Lung Tumor #26	-0.36	-0.17	-0.20	-0.13	0.24
	Lung Tumor #27	-0.21	-0.34	-0.21	-1.21	-0.39
	Lung Tumor #29	-0.03	-0.31	-0.14	-1.50	-0.16
15	Lung Tumor #30	0.28	0.25	0.31	1.27	1.37
	Lung Tumor #31	0.29	0.23	0.18	-0.04	0.45
	Lung Tumor #33	0.29	0.08	0.36	0.02	0.99
20	Colon Tumor 25	0.64	0.45	0.47	-0.30	0.69
	Lung Tumor 28	0.72	0.60	0.54	0.23	1.35
	Colon Tumor 35	0.71	0.38	0.45	-0.87	-0.03
25	Kidney #613	-0.04	0.16	0.07	0.22	0.94
	Lymphoma #854	-0.79	-0.66	-0.42	0.31	1.20
	Breast #545	-0.07	-0.07	0.24	1.92	2.37
30	Colon #539	0.04	0.06	0.12	1.64	0.89
	Colon #575	0.13	0.00	-0.03	1.04	1.86

Tumor	P210	P211	P212	DNA 88153	DNA 62377
Colon #698	0.20	0.00	0.02	0.58	1.43
Colon #499	0.01	0.13	0.01	0.67	1.66
Testes #733	0.18	0.11	0.49	2.52	2.02
Testes #716	-0.62	-0.57	0.26	2.43	2.30

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DISCUSSION AND CONCLUSION:

PRO212 (DNA30942-1134):

10 The ΔC_t values for DNA30942-1134 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA30942-1134 encoding PRO212 occurred in primary lung tumors LT1a, LT3, LT7, LT8, LT10, LT11, LT12, LT13, LT15, LT16, LT17 and LT19; in primary colon tumors CT2, CT3, CT8, CT10, CT12, CT14, CT15, CT16, CT17, CT1, CT4, CT5, CT6, CT7, 15 CT9 and CT11; and in colon tumor cell lines SW480, SW620, WiDr, HCT116 and LS174T. Because amplification of DNA30942-1134 occurs in various tumors and tumor cell lines, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO212 would be expected to be useful in cancer therapy.

20 PRO290 (DNA35680-1212):

The ΔC_t values for DNA35680-1212 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA35680-1212 encoding PRO290 occurred in primary lung tumors LT11, LT13, LT15, LT17, and LT19; and in primary colon tumors CT2 and CT11. 25 Because amplification of DNA35680-1212 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO290 would be expected to be useful in cancer therapy.

PRO341 (DNA26288-1239):

30 The ΔC_t values for DNA26288-1239 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA26288-1239 encoding PRO341 occurred in primary lung tumors LT16, LT17, and LT21. Because amplification of DNA26288-1239 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) 35 directed against PRO341 would be expected to be useful in cancer therapy.

PRO535 (DNA49143-1429):

The ΔC_t values for DNA49143-1429 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA49143-1429 encoding PRO535 occurred in primary lung tumors LT11, LT13 and LT17; and in primary colon tumors CT3, CT10, CT14, CT15, CT16, CT17, CT4, CT6 and CT11. Because amplification of DNA49143-1429 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO535 would be expected to be useful in cancer therapy.

10 PRO619 (DNA49821-1562):

The ΔC_t values for DNA49821-1562 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA49821-1562 encoding PRO619 occurred in primary lung tumors LT3, LT7, LT9, LT10, LT11, LT12, LT13, LT15, LT17, LT18, LT19 and LT21; and in primary colon tumors CT2, CT3, CT15, CT16, CT 17, CT1, CT4, CT5, CT6 and CT11. Because amplification of DNA49821-1562 occurs in tumor, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO619 would be expected to be useful in cancer therapy.

PRO717 (DNA50988-1326):

20 The ΔC_t values for DNA50988-1326 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA50988-1326 encoding PRO717 occurred in primary lung tumors LT13, LT15 and LT16; in primary colon tumors CT15, CT16 and CT17; in lung tumor cell lines A549, Calu-1, H441, H460, SKMES1, H522 and H810; and in colon tumor cell lines SW620, Colo320, HT29, HCT116, SKCO1, SW403, LS174T Colo205, HCT15, HCC2998 and KM12.

Amplification has been confirmed by framework mapping for DNA50988: in primary colon tumors: CT15, CT16, CT17, CT1, CT2, CT3, CT4, CT8, CT9, CT10, CT12 and CT14; in primary lung tumors: LT13, LT15 and LT16; in colon tumor cell lines: SW480 and HCT116; and lung tumor cell line A549. Epicenter mapping for DNA50988 resulted in significant amplification: in primary colon tumors: CT15, CT16, CT17, CT1, CT2, CT3, CT4, CT8, CT9, CT10, CT12 and CT14; in primary lung tumors: LT13, LT15 and LT16; in colon tumor cell lines: SW480 and HCT116; and in lung tumor cell line A549. In contrast, the amplification of the closest known framework markers (with the exception of D233 and D397) (Table 6) or epicenter markers (with the exception of D6) (Table 7) does not occur to a greater extent than that of DNA50988. This strongly suggests that DNA50988 is the gene responsible for the amplification of the particular region on Chromosome 4. Because amplification of DNA50988 occurs in various lung and colon tumors and cell lines (especially colon), it is highly probable to play a significant role in tumor formation or growth. As a result, antagonists (*e.g.*, antibodies) directed against the protein encoded by DNA50988 (PRO717) would be expected to have utility in cancer therapy.

PRO809 (DNA57836-1338):

The ΔC_t values for DNA57836-1338 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA57836-1338 encoding PRO809 occurred in primary lung tumors LT11, LT12, LT13, LT17 and LT21. Because amplification of DNA57836-1338 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO809 would be expected to be useful in cancer therapy.

PRO830 (DNA56866-1342):

The ΔC_t values for DNA56866-1342 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA56866-1342 encoding PRO830 occurred in primary lung tumors LT1a, LT11, LT12, LT17 and LT19. Because amplification of DNA56866-1342 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO830 would be expected to be useful in cancer therapy.

PRO848 (DNA59839-1461):

The ΔC_t values for DNA59839-1461 in a variety of tumors are reported in Table 3A. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3A indicates that significant amplification of nucleic acid DNA59839-1461 encoding PRO848 occurred in primary lung tumors LT11, LT12 and LT21. Because amplification of DNA59839-1461 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO848 would be expected to be useful in cancer therapy.

PRO943 (DNA52192-1369):

The ΔC_t values for DNA52192-1369 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA52192-1369 encoding PRO943 occurred in primary lung tumors LT11, LT12, LT13, LT15, LT17 and LT19; and in primary colon tumors CT2, CT8, CT10, CT12, CT14, CT17, CT1, CT5, CT6, CT11 and CT18. Because amplification of DNA52192-1369 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO943 would be expected to be useful in cancer therapy.

PRO1005 (DNA57708-1411):

The ΔC_t values for DNA57708-1411 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA57708-1411 encoding PRO1005 occurred in primary lung tumors LT1, LT1a, LT3, LT6, LT9, LT12, LT13, LT15, and LT16.

Epicenter mapping for DNA57708 resulted in confirmation of significant amplification in lung tumor cell lines HF545, HF539, HF733 and HF716. In contrast, the amplification of the closest known epicenter markers does not occur to a greater extent than that of DNA57708. This strongly suggests that DNA57708 is the gene responsible for the amplification of the particular region on Chromosome 2. Because amplification of DNA57708 occurs in various lung tumors and cell lines, it is highly probable to play a role in tumor formation or growth. As a result, antagonists (*e.g.*, antibodies) directed against the protein encoded by DNA57708 (PRO1005) would be expected to have utility in cancer therapy.

PRO1009 (DNA57129-1413):

The ΔC_t values for DNA57129-1413 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA57129-1413 encoding PRO1009 occurred in primary colon tumors CT2, CT3, CT8, CT10, CT12, CT14, CT15, CT16, CT17, CT4, CT5 and CT11. Because amplification of DNA57129-1413 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1009 would be expected to be useful in cancer therapy.

PRO1025 (DNA59622-1334):

The ΔC_t values for DNA59622-1334 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA59622-1334 encoding PRO1025 occurred in primary lung tumors LT3, LT6, LT7, LT9, and LT10; and in primary colon tumors CT2, CT3, CT10, CT14, CT1, CT4, CT7 and CT9. Because amplification of DNA59622-1334 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1025 would be expected to be useful in cancer therapy.

PRO1030 (DNA59485-1336):

The ΔC_t values for DNA59485-1336 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA59485-1336 encoding PRO1030 occurred in primary lung tumors LT4, LT7, LT9 and LT10; and in primary colon tumors CT2, CT3, CT10, CT12, CT14, CT15, CT16, CT17 and CT5. Because amplification of DNA59485-1336 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1030 would be expected to be useful in cancer therapy.

PRO1097 (DNA59841-1460):

The ΔC_t values for DNA59841-1460 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table

3B indicates that significant amplification of nucleic acid DNA59841-1460 encoding PRO1097 occurred in primary lung tumors LT2 and LT9; and in primary colon tumors CT14, CT16 and CT17. Because amplification of DNA59841-1460 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1097 would be expected to be useful in cancer therapy.

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PRO1107 (DNA59606-1471):

The ΔC_t values for DNA59606-1471 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA59606-1471 encoding PRO1107 occurred in primary lung tumors LT13, LT15, and LT16; and in primary colon tumors CT2, CT3, CT10, CT14, CT1 and CT4. Because amplification of DNA59606-1471 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1107 would be expected to be useful in cancer therapy.

15 PRO1111 (DNA58721-1475):

The ΔC_t values for DNA58721-1475 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA58721-1475 encoding PRO1111 occurred in primary lung tumors LT3, LT4, LT7, LT9, LT10, LT11 and LT17; and in primary colon tumors CT14, CT7, CT11 and CT18. Because amplification of DNA58721-1475 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1111 would be expected to be useful in cancer therapy.

PRO1153 (DNA59842-1502):

25 The ΔC_t values for DNA59842-1502 in a variety of tumors are reported in Table 3B. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3B indicates that significant amplification of nucleic acid DNA59842-1502 encoding PRO1153 occurred in primary lung tumors LT4 and LT7. Because amplification of DNA59842-1502 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1153 would be expected to be useful in cancer therapy.

PRO1182 (DNA59848-1512):

35 The ΔC_t values for DNA59848-1512 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA59848-1512 encoding PRO1182 occurred in primary lung tumors LT1, LT4 and LT9. Because amplification of DNA59848-1512 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1182 would be expected to be useful in cancer therapy.

PRO1184 (DNA59220-1514):

The ΔC_t values for DNA59220-1514 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA encoding PRO1184 occurred in primary lung tumors LT1a, LT4, LT6, LT12, LT16, LT18, LT19 and LT21. Because amplification of DNA59220-1514 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1184 would be expected to be useful in cancer therapy.

PRO1187 (DNA62876-1517):

The ΔC_t values for DNA62876-1517 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA62876-1517 encoding PRO1187 occurred in primary lung tumors LT12, LT15 and LT16. Because amplification of DNA62876-1517 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1187 would be expected to be useful in cancer therapy.

PRO1281 (DNA59820-1549):

The ΔC_t values for DNA59820-1549 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA59820-1549 encoding PRO1281 occurred in primary colon tumors CT2 and CT12. Because amplification of DNA59820-1549 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1281 would be expected to be useful in cancer therapy.

PRO23 (DNA36640):

The ΔC_t values for DNA36640 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA36640 encoding PRO23 occurred in primary lung tumors LT13, LT15 and LT16; in primary colon tumors CT17, CT1, CT4, CT5, CT7 and CT11; in lung tumor cell line H441; and in colon tumor cell lines Colo320, HT29, SKCO1, SW403, LS174T, HCT15, HCC2998 and KM12. Because amplification of DNA36640 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO23 would be expected to be useful in cancer therapy.

PRO39 (DNA36651):

The ΔC_t values for DNA36651 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA36651 encoding PRO39 occurred in primary lung

tumors LT1a, LT2, LT4, LT12, LT13, LT15 and LT16; in primary colon tumors CT2, CT8, CT10, CT14, CT4, CT5, CT7 and CT11; and colon tumor cell lines Colo320, HT29, SKCO1, SW403 and LS174T. Because amplification of DNA36651 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO39 would be expected to be useful in cancer therapy.

PRO834 (DNA56538):

The ΔC_t values for DNA56538 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA56538 encoding PRO834 occurred in primary lung tumor LT1a; and primary colon tumors CT2, CT8, CT10, CT12, CT14, CT15, CT16, CT1, CT4, CT5, CT6 and CT11. Because amplification of DNA56538 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO834 would be expected to be useful in cancer therapy.

PRO1317 (DNA71166):

The ΔC_t values for DNA71166 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA71166 encoding PRO1317 occurred in primary lung tumors LT1, LT1a, LT9, LT10, LT15, LT17 and LT22. Because amplification of DNA71166 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1317 would be expected to be useful in cancer therapy.

PRO1710 (DNA82331):

The ΔC_t values for DNA82331 in a variety of tumors are reported in Table 3C. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3C indicates that significant amplification of nucleic acid DNA82331 encoding PRO1710 occurred in primary lung tumors LT12, LT13, LT15 and LT16; in primary colon tumors CT2, CT8, CT10, CT14, CT16, CT4, CT5, CT7 and CT11; in lung tumor cell line Calu-1; and in colon tumor cell line SW620. Because amplification of DNA82331 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO1710 would be expected to be useful in cancer therapy.

PRO2094 (DNA83123):

The ΔC_t values for DNA83123 in a variety of tumors are reported in Table 3D. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3D indicates that significant amplification of nucleic acid DNA83123 encoding PRO2094 occurred in primary lung tumors LT7, LT9 and LT15; and in primary colon tumors CT8, CT14, CT15, CT17, CT1, CT4 and CT9.

Amplification has been confirmed by framework mapping for DNA83123. Table 12 indicates that

significant amplification for DNA83123 was confirmed in colon tumors CT15, CT16 and CT17; in lung tumors LT13 and LT15 ; in colon tumor cell lines: SW480, Colo320, HCT75, HCC2998, KM12, SKCO1 and SW620; and in lung tumor cell lines: Calu-6, H157 and H441. Epicenter mapping for DNA83123 resulted in significant amplification in: kidney cell line 613, colon cell line 575 and testes cell line 733. In contrast, the amplification of the closest known framework markers (with the exception of F348) (Table 12) and epicenter markers (Table 13) does not occur to a greater extent than that of DNA83123. This strongly suggests that DNA83123 is the gene responsible for the amplification of the particular region in Chromosome 6. Because amplification of DNA83123 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO2094 would be expected to be useful in cancer therapy.

PRO2145 (DNA88050):

The ΔC_t values for DNA88050 in a variety of tumors are reported in Table 3D. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3D indicates that significant amplification of nucleic acid DNA88050 encoding PRO2145 occurred in primary lung tumors LT1, LT1a, LT3, LT7, LT9, LT12, LT13, LT15 and LT8; in lung tumor cell line A549; and in colon tumor cell line HCT116. Because amplification of DNA88050 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO2145 would be expected to be useful in cancer therapy.

PRO2198 (DNA88153):

The ΔC_t values for DNA88153 in a variety of tumors are reported in Table 3D. A ΔC_t of >1 was typically used as the threshold value for amplification scoring, as this represents a doubling of gene copy. Table 3D indicates that significant amplification of nucleic acid DNA88153 encoding PRO2198 occurred in primary lung tumors LT1a, LT13 and LT16; in primary colon tumors CT2, CT10, CT14, CT15, CT16 and CT17; in lung tumor cell line A549; and in colon tumor cell lines Colo320 and HCT116.

Amplification has been confirmed by framework mapping for DNA88153: in primary colon tumor CT15; in primary lung tumors: LT13 and LT16; in colon tumor cell lines: HCC2998, KM12, SKC0-1, SW480 and HCT116; and lung tumor cell line A549. Epicenter mapping for DNA88153 resulted in significant amplification: in primary colon tumors: CT15, CT16, CT17, CT1, CT2, CT3, CT4, CT8, CT9, CT10, CT12 and CT14; in primary lung tumors: LT13, LT15 and LT16; in colon tumor cell lines: SW480 and HCT116; and in lung tumor cell line A549. In contrast, the amplification of the closest known framework markers (with the exception of P7 and P154)(Table 16) or epicenter markers (Table 17) does not occur to a greater extent than that of DNA. This strongly suggests that DNA88153 is the gene responsible for the amplification of the particular region on Chromosome 16. Because amplification of DNA88153 occurs in various tumors, it is likely associated with tumor formation and/or growth. As a result, antagonists (*e.g.*, antibodies) directed against PRO2198 would be expected to be useful in cancer therapy.

EXAMPLE 24

In situ Hybridization

In situ hybridization is a powerful and versatile technique for the detection and localization of nucleic acid sequences within cell or tissue preparations. It may be useful, for example, to identify sites of gene expression, analyze the tissue distribution of transcription, identify and localize viral infection, follow changes in specific mRNA synthesis and aid in chromosome mapping.

In situ hybridization was performed following an optimized version of the protocol by Lu and Gillett, Cell Vision, 1:169-176 (1994), using PCR-generated ^{33}P -labeled riboprobes. Briefly, formalin-fixed, paraffin-embedded human tissues were sectioned, deparaffinized, deproteinized in proteinase K (20 g/ml) for 15 minutes at 37°C, and further processed for *in situ* hybridization as described by Lu and Gillett, supra. A [^{33}P] UTP-labeled antisense riboprobe was generated from a PCR product and hybridized at 55°C overnight. The slides were dipped in Kodak NTB2 nuclear track emulsion and exposed for 4 weeks.

^{33}P -Riboprobe synthesis

6.0 μl (125 mCi) of ^{33}P -UTP (Amersham BF 1002, SA<2000 Ci/mmol) were speed vac dried. To each tube containing dried ^{33}P -UTP, the following ingredients were added:

2.0 μl 5x transcription buffer

1.0 μl DTT (100 mM)

2.0 μl NTP mix (2.5 mM : 10 μl ; each of 10 mM GTP, CTP & ATP + 10 μl H₂O)

1.0 μl UTP (50 μM)

1.0 μl Rnasin

1.0 μl DNA template (1 μg)

1.0 μl H₂O

1.0 μl RNA polymerase (for PCR products T3 = AS, T7 = S, usually)

The tubes were incubated at 37°C for one hour. 1.0 μl RQ1 DNase were added, followed by incubation at 37°C for 15 minutes. 90 μl TE (10 mM Tris pH 7.6/1mM EDTA pH 8.0) were added, and the mixture was pipetted onto DE81 paper. The remaining solution was loaded in a Microcon-50 ultrafiltration unit, and spun using program 10 (6 minutes). The filtration unit was inverted over a second tube and spun using program 2 (3 minutes). After the final recovery spin, 100 μl TE were added. 1 μl of the final product was pipetted on DE81 paper and counted in 6 ml of Biofluor II.

The probe was run on a TBE/urea gel. 1-3 μl of the probe or 5 μl of RNA Mrk III were added to 3 μl of loading buffer. After heating on a 37°C heat block for three minutes, the gel was immediately placed on ice. The wells of gel were flushed, the sample loaded, and run at 180-250 volts for 45 minutes. The gel was wrapped in saran wrap and exposed to XAR film with an intensifying screen in -70°C freezer one hour to overnight.

^{33}P -Hybridization

Pretreatment of frozen sections: The slides were removed from the freezer, placed on aluminium trays and thawed at room temperature for 5 minutes. The trays were placed in a 55°C incubator for five minutes to reduce condensation. The slides were fixed for 10 minutes in 4% paraformaldehyde on ice in the fume hood, and washed in 0.5 x SSC for 5 minutes, at room temperature (25 ml 20 x SSC + 975 ml SQ H₂O). After deproteinization in 0.5

$\mu\text{g/ml}$ proteinase K for 10 minutes at 37°C ($12.5\ \mu\text{l}$ of $10\ \text{mg/ml}$ stock in $250\ \text{ml}$ prewarmed RNase-free RNase buffer), the sections were washed in $0.5 \times \text{SSC}$ for 10 minutes at room temperature. The sections were dehydrated in 70%, 95%, 100% ethanol, 2 minutes each.

Pretreatment of paraffin-embedded sections: The slides were deparaffinized, placed in $\text{SQ H}_2\text{O}$, and rinsed twice in $2 \times \text{SSC}$ at room temperature, for 5 minutes each time. The sections were deproteinated in $20\ \mu\text{g/ml}$ proteinase K ($500\ \mu\text{l}$ of $10\ \text{mg/ml}$ in $250\ \text{ml}$ RNase-free RNase buffer; 37°C , 15 minutes) - human embryo, or $8 \times$ proteinase K ($100\ \mu\text{l}$ in $250\ \text{ml}$ RNase buffer, 37°C , 30 minutes) - formalin tissues. Subsequent rinsing in $0.5 \times \text{SSC}$ and dehydration were performed as described above.

Prehybridization: The slides were laid out in plastic box lined with Box buffer ($4 \times \text{SSC}$, 50% formamide) - saturated filter paper. The tissue was covered with $50\ \mu\text{l}$ of hybridization buffer ($3.75\ \text{g}$ Dextran Sulfate + $6\ \text{ml}$ $\text{SQ H}_2\text{O}$), vortexed and heated in the microwave for 2 minutes with the cap loosened. After cooling on ice, $18.75\ \text{ml}$ formamide, $3.75\ \text{ml}$ $20 \times \text{SSC}$ and $9\ \text{ml}$ $\text{SQ H}_2\text{O}$ were added, the tissue was vortexed well, and incubated at 42°C for 1-4 hours.

Hybridization: $1.0 \times 10^6\ \text{cpm}$ probe and $1.0\ \mu\text{l}$ tRNA ($50\ \text{mg/ml}$ stock) per slide were heated at 95°C for 3 minutes. The slides were cooled on ice, and $48\ \mu\text{l}$ hybridization buffer were added per slide. After vortexing, $50\ \mu\text{l}$ ^{32}P mix were added to $50\ \mu\text{l}$ prehybridization on slide. The slides were incubated overnight at 55°C .

Washes: Washing was done 2×10 minutes with $2 \times \text{SSC}$, EDTA at room temperature ($400\ \text{ml}$ $20 \times \text{SSC}$ + $16\ \text{ml}$ 0.25M EDTA, $V_f=4\text{L}$), followed by RNaseA treatment at 37°C for 30 minutes ($500\ \mu\text{l}$ of $10\ \text{mg/ml}$ in $250\ \text{ml}$ RNase buffer = $20\ \mu\text{g/ml}$). The slides were washed 2×10 minutes with $2 \times \text{SSC}$, EDTA at room temperature. The stringency wash conditions were as follows: 2 hours at 55°C , $0.1 \times \text{SSC}$, EDTA ($20\ \text{ml}$ $20 \times \text{SSC}$ + $16\ \text{ml}$ EDTA, $V_f=4\text{L}$).

DNA30942-1134 (TNF-R Homolog)

Expression in Human Tissues:

Expression in fetal and adult tissues was observed to give negative results. Fetal tissues examined included: placenta, umbilical cord, brain, spinal cord, eye, optic nerve, trachea, lung, heart, liver, spleen, esophagus, small intestine, pancreas, adrenals, thyroid, body wall and lower limb. Adult tissues examined included: liver, kidney, adrenals, myocardium, aorta, spleen, lymph node, pancreas, lung and skin.

Expression in Lung Carcinomas, Tumor Block and Breast Carcinomas:

Expression was observed in mononuclear phagocytes in the normal chimp thymus, as well as in gastric carcinoma, colorectal cancer, breast cancer and lung cancer. Expression was observed in malignant cells in an osteosarcoma and a poorly differentiated liposarcoma. A possible signal may be found in the malignant cells of a testicular teratoma and breast cancers. In one of the lung cancers, a scattered signal was seen over a high endothelial venule within pulmonary lymphoid tissue.

Fetal tissues examined (E12-E16 weeks) included: placenta, umbilical cord, liver, kidney, adrenals, thyroid, lung, heart, great vessels, esophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb. Adult human tissues examined included: liver, kidney, adrenals, myocardium, aorta, spleen, lung, skin, chondrosarcoma, eye, stomach, gastric carcinoma, colon, colonic carcinoma, renal cell carcinoma,

prostate, bladder mucosa and gall bladder. Acetaminophen induced liver injury and hepatic cirrhosis. Rhesus tissues examined included: cerebral cortex (rm) and hippocampus (rm). Chimp tissues examined included: thyroid, parathyroid, ovary, nerve, tongue, thymus, adrenals, gastric mucosa and salivary gland.

Expression in Lung Adenocarcinoma and Squamous Carcinoma:

5 Eight adenocarcinomas and seven squamous lung carcinomas were examined. Actins were strongly positive in all tumors, indicating that all are suitable for *in situ* hybridization analysis. Expression of DNA30942 was observed in 6 of the tumors as follows:

6727-95 / squamous carcinoma - strongly expressed over neoplastic epithelium

9558-95 / squamous carcinoma - expression over neoplastic epithelium

10 12235-95 / adenocarcinoma - expression over *in situ* and infiltrating tumor cells

6545-95 & 4187-96 / squamous carcinomas - expression over cells in tumor stroma, no expression seen over tumor cells

12954-94 / squamous carcinoma - possible weak expression over stromal cells

15 C-111V DNA30942-p1:

5'-GGATTCTAATACGACTCACTATAGGGCTCGCTGCTGTGCCTGGTGTG-3' (SEQ ID NO:182)

C-111W DNA30942-p2:

5'-CTATGAAATTAACCCTCACTAAAGGGACCGCTGCAGCCTCTTGATGGA-3' (SEQ ID NO:183)

20

DNA52192-1369 (FGF receptor-like molecule)

Expression was observed in fetal skeletal muscle and long bone cartilage. Elsewhere, a relatively high background signal is a problem. In one fetal liver, expression appears to occur at sites of hematopoiesis. The other fetal liver was negative. Fetal tissues examined (E12-E16 weeks) included: placenta, umbilical cord, liver, kidney,

25 adrenals, thyroid, lung, heart, great vessels, esophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb. Adult human tissues examined included: liver, kidney, adrenals, myocardium, aorta, spleen, lung, skin, chondrosarcoma, eye, stomach, gastric carcinoma, colon, colonic carcinoma, renal cell carcinoma, prostate, bladder mucosa and gall bladder. Acetaminophen induced liver injury and hepatic cirrhosis. Rhesus tissues examined included cerebral cortex (rm) and hippocampus (rm). Chimp tissues examined
30 included: thyroid, parathyroid, ovary, nerve, tongue, thymus, adrenals, gastric mucosa and salivary gland.

D-269E FGFr p1:

5'-GGATTCTAATACGACTCACTATAGGGCCGCTGACCATGTGGACCAAGG-3' (SEQ ID NO:184)

35 C-257I FGFr p2:

5'-CTATGAAATTAACCCTCACTAAAGGGATCTGGCAGCACGGTGAGGAAG-3' (SEQ ID NO:185)

EXAMPLE 25

Use of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 as a
5 hybridization probe

The following method describes use of a nucleotide sequence encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide as a hybridization probe.

10 DNA comprising the coding sequence of a full-length or mature "PRO" polypeptide as disclosed herein and/or fragments thereof may be employed as a probe to screen for homologous DNAs (such as those encoding naturally-occurring variants of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145
15 or PRO2198 in human tissue cDNA libraries or human tissue genomic libraries.

Hybridization and washing of filters containing either library DNAs is performed under the following high stringency conditions. Hybridization of radiolabeled PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094,
20 PRO2145 or PRO2198-derived probe to the filters is performed in a solution of 50% formamide, 5x SSC, 0.1% SDS, 0.1% sodium pyrophosphate, 50 mM sodium phosphate, pH 6.8, 2x Denhardt's solution, and 10% dextran sulfate at 42°C for 20 hours. Washing of the filters is performed in an aqueous solution of 0.1x SSC and 0.1% SDS at 42°C.

DNAs having a desired sequence identity with the DNA encoding full-length native sequence PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009,
25 PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can then be identified using standard techniques known in the art.

EXAMPLE 26

30 Expression of "PRO" Polypeptides in *E. coli*.

This example illustrates preparation of an unglycosylated form of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 by recombinant exoppression in *E. coli*.

35 The DNA sequence encoding the PRO polypeptide of interest is initially amplified using selected PCR primers. The primers should contain restriction enzyme sites which correspond to the restriction enzyme sites on the selected expression vector. A variety of expression vectors may be employed. An example of a suitable vector is pBR322 (derived from *E. coli*; see Bolivar *et al.*, Gene, 2:95 (1977)) which contains genes for ampicillin and

tetracycline resistance. The vector is digested with restriction enzyme and dephosphorylated. The PCR amplified sequences are then ligated into the vector. The vector will preferably include sequences which encode for an antibiotic resistance gene, a *trp* promoter, a poly-His leader (including the first six STII codons, poly-His sequence, and enterokinase cleavage site), the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 coding region, lambda transcriptional terminator, and an *argU* gene.

The ligation mixture is then used to transform a selected *E. coli* strain using the methods described in Sambrook *et al.*, *supra*. Transformants are identified by their ability to grow on LB plates and antibiotic resistant colonies are then selected. Plasmid DNA can be isolated and confirmed by restriction analysis and DNA sequencing.

Selected clones can be grown overnight in liquid culture medium such as LB broth supplemented with antibiotics. The overnight culture may subsequently be used to inoculate a larger scale culture. The cells are then grown to a desired optical density, during which the expression promoter is turned on.

After culturing the cells for several more hours, the cells can be harvested by centrifugation. The cell pellet obtained by the centrifugation can be solubilized using various agents known in the art, and the solubilized PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 protein can then be purified using a metal chelating column under conditions that allow tight binding of the protein.

PRO619, PRO1184 and PRO834 were successfully expressed in *E. coli* in a poly-His tagged form was accomplished by using the following procedure. The DNA encoding desired protein was initially amplified using selected PCR primers. The primers contained restriction enzyme sites which correspond to the restriction enzyme sites on the selected expression vector, and other useful sequences providing for efficient and reliable translation initiation, rapid purification on a metal chelation column, and proteolytic removal with enterokinase. The PCR-amplified, poly-His tagged sequences were then ligated into an expression vector, which was used to transform an *E. coli* host based on strain 52 (W3110 fuhA(tonA) lon galE rpoHts(htpRts) clpP(lacIq). Transformants were first grown in LB containing 50 mg/ml carbenicillin at 30°C with shaking until an O.D.600 of 3-5 was reached. Cultures were then diluted 50-100 fold into CRAP media (prepared by mixing 3.57 g (NH₄)₂SO₄, 0.71 g sodium citrate•2H₂O, 1.07 g KCl, 5.36 g Difco yeast extract, 5.36 g Sheffield hycase SF in 500 mL water, as well as 110 mM MPOS, pH 7.3, 0.55% (w/v) glucose and 7 mM MgSO₄) and grown for approximately 20-30 hours at 30°C with shaking. Samples were removed to verify expression by SDS-PAGE analysis, and the bulk culture is centrifuged to pellet the cells. Cell pellets were frozen until purification and refolding.

E. coli paste from 0.5 to 1 L fermentations (6-10 g pellets) was resuspended in 10 volumes (w/v) in 7 M guanidine, 20 mM Tris, pH 8 buffer. Solid sodium sulfite and sodium tetrathionate is added to make final concentrations of 0.1M and 0.02 M, respectively, and the solution was stirred overnight at 4°C. This step results in a denatured protein with all cysteine residues blocked by sulfitolization. The solution was centrifuged at 40,000 rpm in a Beckman Ultracentrifuge for 30 min. The supernatant was diluted with 3-5 volumes of metal chelate column buffer (6 M guanidine, 20 mM Tris, pH 7.4) and filtered through 0.22 micron filters to clarify. The clarified extract

was loaded onto a 5 ml Qiagen Ni-NTA metal chelate column equilibrated in the metal chelate column buffer. The column was washed with additional buffer containing 50 mM imidazole (Calbiochem, Utrol grade), pH 7.4. The protein was eluted with buffer containing 250 mM imidazole. Fractions containing the desired protein were pooled and stored at 4°C. Protein concentration was estimated by its absorbance at 280 nm using the calculated extinction coefficient based on its amino acid sequence.

The proteins were refolded by diluting sample slowly into freshly prepared refolding buffer consisting of: 20 mM Tris, pH 8.6, 0.3 M NaCl, 2.5 M urea, 5 mM cysteine, 20 mM glycine and 1 mM EDTA. Refolding volumes were chosen so that the final protein concentration was between 50 to 100 micrograms/ml. The refolding solution was stirred gently at 4°C for 12-36 hours. The refolding reaction was quenched by the addition of TFA to a final concentration of 0.4% (pH of approximately 3). Before further purification of the protein, the solution was filtered through a 0.22 micron filter and acetonitrile was added to 2-10% final concentration. The refolded protein was chromatographed on a Poros R1/H reversed phase column using a mobile buffer of 0.1% TFA with elution with a gradient of acetonitrile from 10 to 80%. Aliquots of fractions with A280 absorbance were analyzed on SDS polyacrylamide gels and fractions containing homogeneous refolded protein were pooled. Generally, the properly refolded species of most proteins are eluted at the lowest concentrations of acetonitrile since those species are the most compact with their hydrophobic interiors shielded from interaction with the reversed phase resin. Aggregated species are usually eluted at higher acetonitrile concentrations. In addition to resolving misfolded forms of proteins from the desired form, the reversed phase step also removes endotoxin from the samples.

Fractions containing the desired proteins were pooled and the acetonitrile removed using a gentle stream of nitrogen directed at the solution. Proteins were formulated into 20 mM Hepes, pH 6.8 with 0.14 M sodium chloride and 4% mannitol by dialysis or by gel filtration using G25 Superfine (Pharmacia) resins equilibrated in the formulation buffer and sterile filtered.

EXAMPLE 27

Expression of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 in mammalian cells

This example illustrates preparation of a potentially glycosylated form of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 by recombinant expression in mammalian cells.

The vector, pRK5 (see EP 307,247, published March 15, 1989), is employed as the expression vector. Optionally, the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 DNA is ligated into pRK5 with selected restriction enzymes to allow insertion of the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097,

PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 DNA using ligation methods such as described in Sambrook *et al.*, supra. The resulting vector is called pRK5-PRO212, pRK5-PRO290, pRK5-PRO341, pRK5-PRO535, pRK5-PRO619, pRK5-PRO717, pRK5-PRO809, pRK5-PRO830, pRK5-PRO848, pRK-PRO943, pRK5-PRO1005, pRK5-PRO1009, pRK5-PRO1025, pRK5-PRO1030, pRK5-PRO1097, pRK5-PRO1107, pRK5-PRO1111, pRK5-PRO1153, pRK5-PRO1182, pRK5-PRO1184, pRK5-PRO1187, pRK5-PRO1281, pRK5-PRO23, pRK5-PRO39, pRK5-PRO834, pRK5-1317, pRK5-PRO1710, pRK5-PRO2094, pRK5-PRO2145 or pRK5-PRO2198.

In one embodiment, the selected host cells may be 293 cells. Human 293 cells (ATCC CCL 1573) are grown to confluence in tissue culture plates in medium such as DMEM supplemented with fetal calf serum and optionally, nutrient components and/or antibiotics. About 10 μ g pRK5-PRO212, pRK5-PRO290, pRK5-PRO341, pRK5-PRO535, pRK5-PRO619, pRK5-PRO717, pRK5-PRO809, pRK5-PRO830, pRK5-PRO848, pRK-PRO943, pRK5-PRO1005, pRK5-PRO1009, pRK5-PRO1025, pRK5-PRO1030, pRK5-PRO1097, pRK5-PRO1107, pRK5-PRO1111, pRK5-PRO1153, pRK5-PRO1182, pRK5-PRO1184, pRK5-PRO1187, pRK5-PRO1281, pRK5-PRO23, pRK5-PRO39, pRK5-PRO834, pRK5-PRO1317, pRK5-PRO1710, pRK5-PRO2094, pRK5-PRO2145 or pRK5-PRO2198 DNA is mixed with about 1 μ g DNA encoding the VA RNA gene [Thimmappaya *et al.*, Cell, 31:543 (1982)] and dissolved in 500 μ l of 1 mM Tris-HCl, 0.1 mM EDTA, 0.227 M CaCl₂. To this mixture is added, dropwise, 500 μ l of 50 mM HEPES (pH 7.35), 280 mM NaCl, 1.5 mM NaPO₄, and a precipitate is allowed to form for 10 minutes at 25°C. The precipitate is suspended and added to the 293 cells and allowed to settle for about four hours at 37°C. The culture medium is aspirated off and 2 ml of 20% glycerol in PBS is added for 30 seconds. The 293 cells are then washed with serum free medium, fresh medium is added and the cells are incubated for about 5 days.

Approximately 24 hours after the transfections, the culture medium is removed and replaced with culture medium (alone) or culture medium containing 200 μ Ci/ml ³⁵S-cysteine and 200 μ Ci/ml ³⁵S-methionine. After a 12 hour incubation, the conditioned medium is collected, concentrated on a spin filter, and loaded onto a 15% SDS gel. The processed gel may be dried and exposed to film for a selected period of time to reveal the presence of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide. The cultures containing transfected cells may undergo further incubation (in serum free medium) and the medium is tested in selected bioassays.

In an alternative technique, PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 DNA may be introduced into 293 cells transiently using the dextran sulfate method described by Sompanyrac *et al.*, Proc. Natl. Acad. Sci., 12:7575 (1981). 293 cells are grown to maximal density in a spinner flask and 700 μ g pRK5-PRO212, pRK5-PRO290, pRK5-PRO341, pRK5-PRO535, pRK5-PRO619, pRK5-PRO717, pRK5-PRO809, pRK5-PRO830, pRK5-PRO848, pRK-PRO943, pRK5-PRO1005, pRK5-PRO1009, pRK5-PRO1025, pRK5-PRO1030, pRK5-PRO1097, pRK5-PRO1107, pRK5-PRO1111, pRK5-PRO1153, pRK5-

PRO1182, pRK5-PRO1184, pRK5-PRO1187, pRK5-PRO1281, pRK5-PRO23, pRK5-PRO39, pRK5-PRO834, pRK5-PRO1317, pRK5-PRO1710, pRK5-PRO2094, pRK5-PRO2145 or pRK5-PRO2198 DNA is added. The cells are first concentrated from the spinner flask by centrifugation and washed with PBS. The DNA-dextran precipitate is incubated on the cell pellet for four hours. The cells are treated with 20% glycerol for 90 seconds, washed with tissue culture medium, and re-introduced into the spinner flask containing tissue culture medium, 5 μ g/ml bovine insulin and 0.1 μ g/ml bovine transferrin. After about four days, the conditioned media is centrifuged and filtered to remove cells and debris. The sample containing expressed PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can then be concentrated and purified by any selected method, such as dialysis and/or column chromatography.

In another embodiment PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can be expressed in CHO cells. The pRK5-PRO212, pRK5-PRO290, pRK5-PRO341, pRK5-PRO535, pRK5-PRO619, pRK5-PRO717, pRK5-PRO809, pRK5-PRO830, pRK5-PRO848, pRK5-PRO943, pRK5-PRO1005, pRK5-PRO1009, pRK5-PRO1025, pRK5-PRO1030, pRK5-PRO1097, pRK5-PRO1107, pRK5-PRO1111, pRK5-PRO1153, pRK5-PRO1182, pRK5-PRO1184, pRK5-PRO1187, pRK5-PRO1281, pRK5-PRO23, pRK5-PRO39, pRK5-PRO834, pRK5-PRO1317, pRK5-PRO1710, pRK5-PRO2094, pRK5-PRO2145 or pRK5-PRO2198 vector can be transfected into CHO cells using known reagents such as CaPO_4 or DEAE-dextran. As described above, the cell cultures can be incubated, and the medium replaced with culture medium (alone) or medium containing a radiolabel such as ^{35}S -methionine. After determining the presence of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, the culture medium may be replaced with serum free medium. Preferably, the cultures are incubated for about 6 days, and then the conditioned medium is harvested. The medium containing the expressed PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can then be concentrated and purified by any selected method.

Epitope-tagged PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 may also be expressed in host CHO cells. The PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 may be subcloned out of the pRK5 vector. The subclone insert can undergo PCR to fuse in frame with a selected epitope tag such as a poly-His tag into a Baculovirus expression vector. The poly-His

tagged PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 insert can then be subcloned into a SV40 driven vector containing a selection marker such as DHFR for selection of stable clones. Finally, the CHO cells can be transfected (as described above) with the SV40 driven vector. Labeling may be performed, as described above, to verify expression. The culture medium containing the expressed poly-His tagged PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can then be concentrated and purified by any selected method, such as by Ni²⁺-chelate affinity chromatography. Expression in CHO and/or COS cells may also be accomplished by a transient expression procedure.

PRO535 and PRO1025 were expressed in CHO cells by a transient expression procedure. In addition, PRO212, PRO535, PRO943, PRO1005, PRO1025 and PRO1182 were stably expressed in CHO cells.

Stable expression in CHO cells was performed using the following procedure. The proteins were expressed as an IgG construct (immunoadhesin), in which the coding sequences for the soluble forms (*e.g.*, extracellular domains) of the respective proteins were fused to an IgG1 constant region sequence containing the hinge, CH2 and CH2 domains and/or is a poly-His tagged form.

Following PCR amplification, the respective DNAs were subcloned in a CHO expression vector using standard techniques as described in Ausubel *et al.*, Current Protocols of Molecular Biology, Unit 3.16, John Wiley and Sons (1997). CHO expression vectors are constructed to have compatible restriction sites 5' and 3' of the DNA of interest to allow the convenient shuttling of cDNA's. The vector used expression in CHO cells is as described in Lucas *et al.*, Nucl. Acids Res., 24:9 (1774-1779 (1996), and uses the SV40 early promoter/enhancer to drive expression of the cDNA of interest and dihydrofolate reductase (DHFR). DHFR expression permits selection for stable maintenance of the plasmid following transfection.

Twelve micrograms of the desired plasmid DNA were introduced into approximately 10 million CHO cells using commercially available transfection reagents Superfect® (Quiagen), Dospet® or Fugene® (Boehringer Mannheim). The cells were grown and described in Lucas *et al.*, supra. Approximately 3 x 10⁷ cells are frozen in an ampule for further growth and production as described below.

The ampules containing the plasmid DNA were thawed by placement into water bath and mixed by vortexing. The contents were pipetted into a centrifuge tube containing 10 mLs of media and centrifuged at 1000 rpm for 5 minutes. The supernatant was aspirated and the cells were resuspended in 10 mL of selective media (0.2 µm filtered PS20 with 5% 0.2 µm diafiltered fetal bovine serum). The cells were then aliquoted into a 100 mL spinner containing 90 mL of selective media. After 1-2 days, the cells were transferred into a 250 mL spinner filled with 150 mL selective growth medium and incubated at 37°C. After another 2-3 days, 250 mL, 500 mL and 2000 mL spinners were seeded with 3 x 10⁵ cells/mL. The cell media was exchanged with fresh media by centrifugation and resuspension in production medium. Although any suitable CHO media may be employed, a production medium described in US Patent No. 5,122,469, issued June 16, 1992 was actually used. 3L production spinner is seeded at 1.2 x 10⁶ cells/mL. On day 0, the cell number pH were determined. On day 1, the spinner was sampled and sparging

with filtered air was commenced. On day 2, the spinner was sampled, the temperature shifted to 33°C, and 30 mL of 500 g/L glucose and 0.6 mL of 10% antifoam (e.g., 35% polydimethylsiloxane emulsion, Dow Corning 365 Medical Grade Emulsion). Throughout the production, pH was adjusted as necessary to keep at around 7.2. After 10 days, or until viability dropped below 70%, the cell culture was harvested by centrifugation and filtering through a 0.22 μ m filter. The filtrate was either stored at 4°C or immediately loaded onto columns for purification.

For the poly-His tagged constructs, the proteins were purified using a Ni-NTA column (Qiagen). Before purification, imidazole was added to the conditioned media to a concentration of 5 mM. The conditioned media was pumped onto a 6 ml Ni-NTA column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 4°C. After loading, the column was washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein was subsequently desalted into a storage buffer containing 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc containing) constructs were purified from the conditioned media as follows. The conditioned medium was pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column was washed extensively with equilibration buffer before elution with 100 mM citric acid, pH 3.5. The eluted protein was immediately neutralized by collecting 1 ml fractions into tubes containing 275 μ L of 1 M Tris buffer, pH 9. The highly purified protein was subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity was assessed by SDS polyacrylamide gels and by N-terminal amino acid sequencing by Edman degradation.

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EXAMPLE 28

Expression of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 in

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Yeast

The following method describes recombinant expression of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 in yeast.

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First, yeast expression vectors are constructed for intracellular production or secretion of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 from the ADH2/GAPDH promoter. DNA encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 and the promoter is inserted into suitable restriction enzyme sites in the selected plasmid to direct intracellular expression of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809,

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PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. For secretion, DNA encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can be cloned into the selected plasmid, together with DNA encoding the ADH2/GAPDH promoter, a native PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 signal peptide or other mammalian signal peptide, or, for example, a yeast alpha-factor or invertase secretory signal/leader sequence, and linker sequences (if needed) for expression of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198.

Yeast cells, such as yeast strain AB110, can then be transformed with the expression plasmids described above and cultured in selected fermentation media. The transformed yeast supernatants can be analyzed by precipitation with 10% trichloroacetic acid and separation by SDS-PAGE, followed by staining of the gels with Coomassie Blue stain.

Recombinant PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can subsequently be isolated and purified by removing the yeast cells from the fermentation medium by centrifugation and then concentrating the medium using selected cartridge filters. The concentrate containing PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 may further be purified using selected column chromatography resins.

EXAMPLE 29

Expression of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 in

Baculovirus-infected Insect Cells

The following method describes recombinant expression in Baculovirus-infected insect cells.

The sequence coding for PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145

or PRO2198 is fused upstream of an epitope tag contained within a baculovirus expression vector. Such epitope tags include poly-His tags and immunoglobulin tags (like Fc regions of IgG). A variety of plasmids may be employed, including plasmids derived from commercially available plasmids such as pVL1393 (Novagen). Briefly, the sequence encoding PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 or the desired portion of the coding sequence of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 such as the sequence encoding the extracellular domain of a transmembrane protein or the sequence encoding the mature protein if the protein is extracellular is amplified by PCR with primers complementary to the 5' and 3' regions. The 5' primer may incorporate flanking (selected) restriction enzyme sites. The product is then digested with those selected restriction enzymes and subcloned into the expression vector.

Recombinant baculovirus is generated by co-transfecting the above plasmid and BaculoGold™ virus DNA (Pharmingen) into *Spodoptera frugiperda* ("Sf9") cells (ATCC CRL 1711) using lipofectin (commercially available from GIBCO-BRL). After 4 - 5 days of incubation at 28°C, the released viruses are harvested and used for further amplifications. Viral infection and protein expression are performed as described by O'Reilley *et al.*, Baculovirus expression vectors: A Laboratory Manual, Oxford: Oxford University Press (1994).

Expressed poly-His tagged PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can then be purified, for example, by Ni²⁺-chelate affinity chromatography as follows. Extracts are prepared from recombinant virus-infected Sf9 cells as described by Rupert *et al.*, Nature, **362**:175-179 (1993). Briefly, Sf9 cells are washed, resuspended in sonication buffer (25 mL Hepes, pH 7.9; 12.5 mM MgCl₂; 0.1 mM EDTA; 10% glycerol; 0.1% NP-40; 0.4 M KCl), and sonicated twice for 20 seconds on ice. The sonicates are cleared by centrifugation, and the supernatant is diluted 50-fold in loading buffer (50 mM phosphate, 300 mM NaCl, 10% glycerol, pH 7.8) and filtered through a 0.45 µm filter. A Ni²⁺-NTA agarose column (commercially available from Qiagen) is prepared with a bed volume of 5 mL, washed with 25 mL of water and equilibrated with 25 mL of loading buffer. The filtered cell extract is loaded onto the column at 0.5 mL per minute. The column is washed to baseline A₂₈₀ with loading buffer, at which point fraction collection is started. Next, the column is washed with a secondary wash buffer (50 mM phosphate; 300 mM NaCl, 10% glycerol, pH 6.0), which elutes nonspecifically bound protein. After reaching A₂₈₀ baseline again, the column is developed with a 0 to 500 mM imidazole gradient in the secondary wash buffer. One mL fractions are collected and analyzed by SDS-PAGE and silver staining or Western blot with Ni²⁺-NTA-conjugated to alkaline phosphatase (Qiagen). Fractions containing the eluted His₁₀-tagged PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198, respectively, are pooled and dialyzed against loading buffer.

Alternatively, purification of the IgG tagged (or Fc tagged) PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 can be performed using known chromatography techniques, including
5 for instance, Protein A or protein G column chromatography.

Following PCR amplification, the respective coding sequences were subcloned into a baculovirus expression vector (pb.PH.IgG for IgG fusions and pb.PH.His.c for poly-His tagged proteins), and the vector and Baculogold® baculovirus DNA (Pharmingen) were co-transfected into 105 *Spodoptera frugiperda* ("Sf9") cells (ATCC CRL 1711), using Lipofectin (Gibco BRL). pb.PH.IgG and pb.PH.His are modifications of the
10 commercially available baculovirus expression vector pVL1393 (Pharmingen), with modified polylinker regions to include the His or Fc tag sequences. The cells were grown in Hink's TNM-FH medium supplemented with 10% FBS (Hyclone). Cells were incubated for 5 days at 28°C. The supernatant was harvested and subsequently used for the first viral amplification by infecting Sf9 cells in Hink's TNM-FH medium supplemented with 10% FBS at an approximate multiplicity of infection (MOI) of 10. Cells were incubated for 3 days at 28°C. The supernatant was
15 harvested and the expression of the constructs in the baculovirus expression vector was determined by batch binding of 1 ml of supernatant to 25 mL of Ni-NTA beads (QIAGEN) for histidine tagged proteins or Protein-A Sepharose CL-4B beads (Pharmacia) for IgG tagged proteins followed by SDS-PAGE analysis comparing to a known concentration of protein standard by Coomassie blue staining.

The first viral amplification supernatant was used to infect a spinner culture (500 ml) of Sf9 cells grown
20 in ESF-921 medium (Expression Systems LLC) at an approximate MOI of 0.1. Cells were incubated for 3 days at 28°C. The supernatant was harvested and filtered. Batch binding and SDS-PAGE analysis was repeated, as necessary, until expression of the spinner culture was confirmed.

The conditioned medium from the transfected cells (0.5 to 3 L) was harvested by centrifugation to remove the cells and filtered through 0.22 micron filters. For the poly-His tagged constructs, the protein construct were
25 purified using a Ni-NTA column (Qiagen). Before purification, imidazole was added to the conditioned media to a concentration of 5 mM. The conditioned media were pumped onto a 6 ml Ni-NTA column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 4°C. After loading, the column was washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein was subsequently desalted into a storage buffer containing
30 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc containing) constructs of proteins were purified from the conditioned media as follows. The conditioned media were pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column was washed extensively with equilibration buffer before
35 elution with 100 mM citric acid, pH 3.5. The eluted protein was immediately neutralized by collecting 1 ml fractions into tubes containing 275 mL of 1 M Tris buffer, pH 9. The highly purified protein was subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity of the proteins was verified by SDS polyacrylamide gel (PAGE) electrophoresis and N-terminal amino acid sequencing by Edman degradation.

Alternatively, a modified baculovirus procedure may be used incorporating high 5 cells. In this procedure, the DNA encoding the desired sequence was amplified with suitable systems, such as Pfu (Stratagene), or fused upstream (5'-of) of an epitope tag contained with a baculovirus expression vector. Such epitope tags include poly-His tags and immunoglobulin tags (like Fc regions of IgG). A variety of plasmids may be employed, including plasmids derived from commercially available plasmids such as pIE1-1 (Novagen). The pIE1-1 and pIE1-2 vectors are designed for constitutive expression of recombinant proteins from the baculovirus ie1 promoter in stably-transformed insect cells (1). The plasmids differ only in the orientation of the multiple cloning sites and contain all promoter sequences known to be important for ie1-mediated gene expression in uninfected insect cells as well as the hr5 enhancer element. pIE1-1 and pIE1-2 include the translation initiation site and can be used to produce fusion proteins. Briefly, the desired sequence or the desired portion of the sequence (such as the sequence encoding the extracellular domain of a transmembrane protein) is amplified by PCR with primers complementary to the 5' and 3' regions. The 5' primer may incorporate flanking (selected) restriction enzyme sites. The product was then digested with those selected restriction enzymes and subcloned into the expression vector. For example, derivatives of pIE1-1 can include the Fc region of human IgG (pb.PH.IgG) or an 8 histidine (pb.PH.His) tag downstream (3'-of) the desired sequence. Preferably, the vector construct is sequenced for confirmation.

Hi5 cells are grown to a confluency of 50% under the conditions of, 27°C, no CO₂, NO pen/strep. For each 150 mm plate, 30 ug of pIE based vector containing the sequence was mixed with 1 ml Ex-Cell medium (Media: Ex-Cell 401 + 1/100 L-Glu JRH Biosciences #14401-78P (note: this media is light sensitive)), and in a separate tube, 100 ul of CellFectin (CellFECTIN (GibcoBRL #10362-010) (vortexed to mix)) was mixed with 1 ml of Ex-Cell medium. The two solutions were combined and allowed to incubate at room temperature for 15 minutes. 8 ml of Ex-Cell media was added to the 2ml of DNA/CellFECTIN mix and this is layered on Hi5 cells that have been washed once with Ex-Cell media. The plate is then incubated in darkness for 1 hour at room temperature. The DNA/CellFECTIN mix is then aspirated, and the cells are washed once with Ex-Cell to remove excess CellFECTIN, 30 ml of fresh Ex-Cell media was added and the cells are incubated for 3 days at 28°C. The supernatant was harvested and the expression of the sequence in the baculovirus expression vector was determined by batch binding of 1 ml of supernatant to 25 mL of Ni-NTA beads (QIAGEN) for histidine tagged proteins or Protein-A Sepharose CL-4B beads (Pharmacia) for IgG tagged proteins followed by SDS-PAGE analysis comparing to a known concentration of protein standard by Coomassie blue staining.

The conditioned media from the transfected cells (0.5 to 3 L) was harvested by centrifugation to remove the cells and filtered through 0.22 micron filters. For the poly-His tagged constructs, the protein comprising the sequence is purified using a Ni-NTA column (Qiagen). Before purification, imidazole is added to the conditioned media to a concentration of 5 mM. The conditioned media was pumped onto a 6 ml Ni-NTA column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 48°C. After loading, the column was washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein was then subsequently desalted into a storage buffer containing 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc containing) constructs of proteins were purified from the conditioned media as follows.

The conditioned media was pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column was washed extensively with equilibration buffer before elution with 100 mM citric acid, pH 3.5. The eluted protein was immediately neutralized by collecting 1 ml fractions into tubes containing 275 mL of 1 M Tris buffer, pH 9. The highly purified protein was subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity of the sequence was assessed by SDS polyacrylamide gels and by N-terminal amino acid sequencing by Edman degradation and other analytical procedures as desired or necessary.

PRO212, PRO535, PRO943, PRO1005, PRO1025, PRO1111, PRO1182, PRO1187 and PRO848 were expressed using the above baculovirus procedure employing high-5 cells.

EXAMPLE 30

Preparation of Antibodies that Bind PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198

This example illustrates preparation of monoclonal antibodies which can specifically bind PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198.

Techniques for producing the monoclonal antibodies are known in the art and are described, for instance, in Goding, *supra*. Immunogens that may be employed include purified PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198, fusion proteins containing PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 and cells expressing recombinant PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 on the cell surface. Selection of the immunogen can be made by the skilled artisan without undue experimentation.

Mice, such as Balb/c, are immunized with the PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 immunogen emulsified in complete Freund's adjuvant and injected subcutaneously or intraperitoneally in an amount from 1-100 micrograms. Alternatively, the immunogen is emulsified in MPL-TDM adjuvant (Ribi Immunochemical Research, Hamilton, MT) and injected into the animal's hind foot pads. The immunized mice are then boosted 10 to 12 days later with additional immunogen emulsified in the selected adjuvant.

Thereafter, for several weeks, the mice may also be boosted with additional immunization injections. Serum samples may be periodically obtained from the mice by retro-orbital bleeding for testing in ELISA assays to detect anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibodies.

After a suitable antibody titer has been detected, the animals "positive" for antibodies can be injected with a final intravenous injection of PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. Three to four days later, the mice are sacrificed and the spleen cells are harvested. The spleen cells are then fused (using 35% polyethylene glycol) to a selected murine myeloma cell line such as P3X63AgU.1, available from ATCC, No. CRL 1597. The fusions generate hybridoma cells which can then be plated in 96 well tissue culture plates containing HAT (hypoxanthine, aminopterin, and thymidine) medium to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.

The hybridoma cells will be screened in an ELISA for reactivity against PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198. Determination of "positive" hybridoma cells secreting the desired monoclonal antibodies against PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 is within the skill in the art.

The positive hybridoma cells can be injected intraperitoneally into syngeneic Balb/c mice to produce ascites containing the anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 monoclonal antibodies. Alternatively, the hybridoma cells can be grown in tissue culture flasks or roller bottles. Purification of the monoclonal antibodies produced in the ascites can be accomplished using ammonium sulfate precipitation, followed by gel exclusion chromatography. Alternatively, affinity chromatography based upon binding of antibody to protein A or protein G can be employed.

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Deposit of Material:

The following materials have been deposited with the American Type Culture Collection, 10801 University Blvd., Manassas, VA 20110-2209, USA (ATCC):

	<u>Material</u>	<u>ATCC Deposit No.:</u>	<u>Deposit Date</u>
5	DNA30942-1134	209254	9/16/97
	DNA35680-1212	209790	4/21/98
	DNA26288-1239	209792	4/21/98
	DNA49143-1429	203013	6/23/98
	DNA49821-1562	209981	6/16/98
10	DNA50988-1326	209814	4/28/98
	DNA57836-1338	203025	6/23/98
	DNA56866-1342	203023	6/23/98
	DNA59839-1461	209988	6/16/98
	DNA52192-1369	203042	7/1/98
15	DNA57708-1411	203021	6/23/98
	DNA57129-1413	209977	6/16/98
	DNA59622-1334	209984	6/16/98
	DNA59485-1336	203015	6/23/98
	DNA59841-1460	203044	7/1/98
20	DNA59606-1471	209945	6/9/98
	DNA58721-1475	203110	8/11/98
	DNA59842-1502	209982	6/16/98
	DNA59848-1512	203088	8/4/98
	DNA59220-1514	209962	6/9/98
25	DNA62876-1517	203095	8/4/98
	DNA59820-1549	203129	8/18/98

These deposits were made under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and the Regulations thereunder (Budapest Treaty). This assures the maintenance of a viable culture of the deposit for 30 years from the date of deposit. The deposit will be made available by the ATCC under the terms of the Budapest Treaty, and subject to an agreement between Genentech, Inc., and the ATCC, which assures permanent and unrestricted availability of the progeny of the culture of the deposit to the public upon issuance of the pertinent U.S. patent or upon laying open to the public of any U.S. or foreign patent application, whichever comes first, and assures availability of the progeny to one determined by the U.S. Commissioner of Patents and Trademarks to be entitled thereto according to 35 U.S.C. § 122 and the Commissioner's rules pursuant thereto (including 37 C.F.R. § 1.14 with particular reference to 886 OG 638).

The assignee of the present application has agreed that if a culture of the materials on deposit should die or be lost or destroyed when cultivated under suitable conditions, the materials will be promptly replaced on

notification with another of the same. Availability of the deposited material is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any government in accordance with its patent laws.

5 The foregoing written specification is considered to be sufficient to enable one skilled in the art to practice the invention. The present inventions is not to be limited in scope by the construct deposited, since the deposited embodiment is intended as a single illustration of certain aspects of the invention and any constructs that are functionally equivalent are within the scope of this invention. The deposit of material herein does not constitute an admission that the written description herein contained is inadequate to enable the practice of any aspect of the invention, including the best mode thereof, nor is it to be construed as limiting the scope of the claims to the specific
10 illustrations that it represents. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and fall within the scope of the appended claims.

WHAT IS CLAIMED IS:

1. An isolated antibody that binds to a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.
2. The antibody of Claim 1 which specifically binds to said polypeptide.
3. The antibody of Claim 1 which induces the death of a cell that expresses said polypeptide.
4. The antibody of Claim 3, wherein said cell is a cancer cell that overexpresses said polypeptide as compared to a normal cell of the same tissue type.
5. The antibody of Claim 1 which is a monoclonal antibody.
6. The antibody of Claim 5 which comprises a non-human complementarity determining region (CDR) or a human framework region (FR).
7. The antibody of Claim 1 which is labeled.
8. The antibody of Claim 1, which is immobilized on a solid support.
9. The antibody of Claim 1 which is an antibody fragment or a single-chain antibody.
10. A composition of matter which comprises an antibody of Claim 1 in admixture with a pharmaceutically acceptable carrier.
11. The composition of matter of Claim 10 which comprises a therapeutically effective amount of said antibody.
12. The composition of matter of Claim 10 which further comprises a cytotoxic or a chemotherapeutic agent.
13. The composition of matter of Claim 10 which is sterile.
14. An isolated nucleic acid molecule that encodes the antibody of Claim 1.

15. A vector comprising the nucleic acid molecule of Claim 14.

16. A host cell comprising the vector of Claim 15.

17. A method for producing an antibody that binds to a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, said method comprising culturing the host cell of Claim 16 under conditions sufficient to allow expression of said antibody and recovering said antibody from the cell culture.

18. An antagonist of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

19. The antagonist of Claim 18 wherein said antagonist inhibits tumor cell growth.

20. An isolated nucleic acid molecule that hybridizes to the complement of a nucleic acid sequence that encodes a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

21. The isolated nucleic acid molecule of Claim 20 wherein said hybridization is under stringent conditions.

22. A method for determining the presence of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in a sample suspected of containing said polypeptide, said method comprising exposing the sample to an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody and determining binding of said antibody to a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in said sample.

23. The method of Claim 22, wherein said sample comprises a cell suspected of comprising a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

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24. A method of diagnosing tumor in a mammal, said method comprising detecting the level of expression of a gene encoding a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide (a) in a test sample of tissue cells obtained from the mammal, and (b) in a control sample of known normal tissue cells of the same cell type, wherein a higher expression level in the test sample, as compared to the control sample, is indicative of the presence of tumor in the mammal from which the test tissue cells were obtained.

25. A method of diagnosing tumor in a mammal, said method comprising (a) contacting an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody with a test sample of tissue cells obtained from the mammal, and (b) detecting the formation of a complex between the anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody and a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in the test sample, wherein the formation of a complex is indicative of the presence of a tumor in said mammal.

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26. The method of Claim 25, wherein said antibody is detectably labeled.

27. The method of Claim 25, wherein said test sample of tissue cells is obtained from an individual suspected of having neoplastic cell growth or proliferation.

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28. A cancer diagnostic kit comprising an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-

PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody and a carrier in suitable packaging.

29. The kit of Claim 28 which further comprises instructions for using said antibody to detect the presence of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in a sample suspected of containing the same.

30. A method for inhibiting the growth of tumor cells, said method comprising exposing tumor cells that express a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide to an effective amount of an agent that inhibits an activity of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, wherein growth of said tumor cells is thereby inhibited.

31. The method of Claim 30, wherein said tumor cells overexpress said polypeptide as compared to normal cells of the same tissue type.

32. The method of Claim 30, wherein said agent is an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody.

33. The method of Claim 32, wherein said anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody induces cell death.

34. The method of Claim 30, wherein said tumor cells are further exposed to radiation treatment, a cytotoxic agent or a chemotherapeutic agent.

35. A method for inhibiting the growth of tumor cells, said method comprising exposing tumor cells that express a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943,

PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide to an effective amount of an agent that inhibits the expression of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, wherein growth of said tumor cells is thereby inhibited.

36. The method of Claim 35, wherein said tumor cells overexpress said polypeptide as compared to normal cells of the same tissue type.

37. The method of Claim 35, wherein said agent is an antisense oligonucleotide.

38. The method of Claim 37, wherein said tumor cells are further exposed to radiation treatment, a cytotoxic agent or a chemotherapeutic agent.

39. An article of manufacture, comprising:
a container;
a label on the container; and
a composition comprising an active agent contained within the container, wherein the composition is effective for inhibiting the growth of tumor cells and wherein the label on the container indicates that the composition is effective for treating conditions characterized by overexpression of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in said tumor cells as compared to in normal cells of the same tissue type.

40. The article of manufacture of Claim 39, wherein said active agent inhibits an activity of and/or the expression of said PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide.

41. The article of manufacture of Claim 40, wherein said active agent is an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody.

42. The article of manufacture of Claim 40, wherein said active agent is an antisense oligonucleotide.

43. A method of identifying a compound that inhibits an activity of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, said method comprising contacting a candidate compound with a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide under conditions and for a time sufficient to allow the two components to interact and determining whether an activity of said PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is inhibited.

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44. The method of Claim 43, wherein said candidate compound is an anti-PRO212, anti-PRO290, anti-PRO341, anti-PRO535, anti-PRO619, anti-PRO717, anti-PRO809, anti-PRO830, anti-PRO848, anti-PRO943, anti-PRO1005, anti-PRO1009, anti-PRO1025, anti-PRO1030, anti-PRO1097, anti-PRO1107, anti-PRO1111, anti-PRO1153, anti-PRO1182, anti-PRO1184, anti-PRO1187, anti-PRO1281, anti-PRO23, anti-PRO39, anti-PRO834, anti-PRO1317, anti-PRO1710, anti-PRO2094, anti-PRO2145 or anti-PRO2198 antibody.

45. The method of Claim 43, wherein said candidate compound or said PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is immobilized on a solid support.

46. The method of Claim 45, wherein the non-immobilized component is detectably labeled.

47. A method of identifying a compound that inhibits an activity of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide, said method comprising the steps of (a) contacting cells and a candidate compound to be screened in the presence of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide under conditions suitable for the induction of a cellular response normally induced by a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111,

PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide and (b) determining the induction of said cellular response to determine if the test compound is an effective antagonist.

5 48. A method for identifying a compound that inhibits the expression of a PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide in cells that express said polypeptide, wherein said method comprises contacting said cells with a candidate compound and determining whether expression of said
10 PRO212, PRO290, PRO341, PRO535, PRO619, PRO717, PRO809, PRO830, PRO848, PRO943, PRO1005, PRO1009, PRO1025, PRO1030, PRO1097, PRO1107, PRO1111, PRO1153, PRO1182, PRO1184, PRO1187, PRO1281, PRO23, PRO39, PRO834, PRO1317, PRO1710, PRO2094, PRO2145 or PRO2198 polypeptide is inhibited.

15 49. The method of Claim 48, wherein said candidate compound is an antisense oligonucleotide.

COMPOSITIONS AND METHODS FOR THE TREATMENT OF TUMOR

The invention concerns compositions and methods for the diagnosis and treatment of neoplastic cell growth
5 and proliferation in mammals, including humans. The invention is based upon the identification of genes that are
amplified in the genome of tumor cells. Such gene amplification is expected to be associated with the overexpression
of the gene product as compared to normal cells of the same tissue type and contribute to tumorigenesis.
Accordingly, the proteins encoded by the amplified genes are believed to be useful targets for the diagnosis and/or
treatment (including prevention) of certain cancers, and may act as predictors of the prognosis of tumor treatment.

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FIGURE 1

TCCGCAGGCGGACCGGGGGCAAAGGAGGTGGCATGTTCGGTCAGGCACAGCAGGGTCCTGTGT
5 CCGCGCTGAGCCGCGCTCTCCCTGCTCCAGCAAGGACC**ATG**AGGGCGCTGGAGGGGCCAGGC
CTGTTCGCTGCTGTGCCTGGTGTGGCGCTGCCTGCCCTGCTGCCGGTGCCGGCTGTACGCGG
AGTGGCAGAAACACCCACCTACCCCTGGCGGGACGCAGAGACAGGGGAGCGGCTGGTGTGCG
CCCAGTGCCCCCAGGCACCTTTGTGTCAGCGGCCGTGCCGCCGAGACAGCCCCACGACGTGT
GGCCCCGTGTCCACCGCGCCACTACACGCAGTTCTGGAACCTACCTGGAGCGCTGCCGCTACTG
10 CAACGTCCTCTGCGGGGAGCGTGAGGAGGAGGCACGGGCTTGCCACGCCACCCACAACCGTG
CCTGCCGCTGCCGCACCGGCTTCTTCGCGCACGCTGGTTTCTGCTTGAGACGCATCGTGT
CCACCTGGTGCCGGCGTGATTGCCCCGGGCACCCCCAGCCAGAACACGCAGTGCCAGCCGTG
CCCCCAGGCACCTTCTCAGCCAGCAGCTCCAGCTCAGAGCAGTGCCAGCCCCACCGCAACT
GCACGGCCCTGGGCCTGGCCCTCAATGTGCCAGGCTCTTCCTCCCATGACACCCTGTGCACC
15 AGCTGCACTGGCTTCCCCCTCAGCACCAGGGTACCAGGAGCTGAGGAGTGTGAGCGTGCCGT
CATCGACTTTGTGGCTTTCCAGGACATCTCCATCAAGAGGCTGCAGCGGCTGCTGCAGGCCC
TCGAGGCCCCGGAGGGCTGGGGTCCGACACCAAGGGCGGGCCGCGCGGCCTTGCAGCTGAAG
CTGCGTCGGCGGCTCACGGAGCTCCTGGGGGCGCAGGACGGGGCGCTGCTGGTGCAGGCTGCT
GCAGGCGCTGCGCGTGCCAGGATGCCCGGGCTGGAGCGGAGCGTCCGTGAGCGCTTCCTCC
20 CTGTGCAC**TGA**TCCTGGCCCCCTCTTATTTATTCTACATCCTTGGCACCCCACTTGCACTGA
AAGAGGCTTTTTTTTAAATAGAAGAAATGAGGTTTCTTAAAAAAAAAAAAAAAAAAAAA

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FIGURE 2

5 Secretion Signal Sequence: amino acids 1-23
Extracellular Domain: amino acids 1-215
N-glycosylation Site: amino acid 173

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MRALEGPGLSLLCLVLALPALLPVPAVRGVAETPTY PWRDAETGERLVCAQC PPGTFVQRPC
RRDSPTTCGPCPPRHYTQFWNYLERCRYCNVLCGERÉÉEARACHATHNRACRCRTGFFAHAG
FCLEHASCPPGAGVIAPGTPSQNTQCQPCPPGTFSASSSSSEQCQPHRNCTALGLALNVPGS
15 SSHDTLCTSCTGFPLSTRVPGAEECERAVIDFVAFQDISIKRLQRL LQALEAPEGWGPTPRA
GRAALQLKLRRRLTELLGAQDGALLVRL LQALRVARMPGLERSVRERFLPVH

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FIGURE 3A

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 CCCACGCGTCCGCCCCACGCGTCCGGCTGAACACCTCTTCTTTGGAGTCAGCCACTGATGAGG
 CAGGGTCCCCACTTGCAGCTGCAGCAGCTGCAGCAGCTGCAGAGCGCTGCTCCTGGCTGGTG
 CCACTGGTGCGCACGCTGCTAGACCGTGCCTATGAGCCGCTGGGGCTGCAGTGGGGACTGCC
 CTCCCTGCCACCCACCAATGGCAGCCCCACCTTCTTTGAAGACTTCCAGGCTTTTTTGTGCCA
 10 CACCCGAATGGCGCCACTTTCATCGACAAACAGGTACAGCCAACCATGTCCCAGTTTCGAAATG
 GACACGTATGCTAAGAGCCACGACCTTATGTCAGGTTTCTGGAATGCCTGCTATGACATGCT
 TATGAGCAGTGGGCAGCGGCGCCAGTGGGAGCGCGCCAGAGTCGTCGGGCCTTCCAGGAGC
 TGGTGCTGGAACCTGCGCAGAGGCGGGCGCGCCTGGAGGGGCTACGCTACACGGCAGTGCTG
 AAGCAGCAGGCAACGCAGCACTCCATGGCCCTGCTGCACTGGGGGGCGCTGTGGCGCCAGCT
 15 CGCCAGCCCATGTGGGGCCTGGGCGCTGAGGGACACTCCCATCCCCCGCTGGAAACTGTCCA
 GCGCCGAGACATATTCACGCATGCGTCTGAAGCTGGTGCCCAACCATCACTTCGACCCTCAC
 CTGGAAGCCAGCGCTCTCCGAGACAATCTGGGTGAGGTTCCCTTGACACCCACCGAGGAGGC
 CTCCTGCTTCTGGCAGTGACCAAAGAGGGCCAAAGTGAGCACCCACCCGAGTTGCTGCAGG
 AGGACCAGCTCGGCGAGGACGAGCTGGCTGAGCTGGAGACCCCGATGGAGGCAGCAGAACTG
 20 GATGAGCAGCGTGAGAAGCTGGTGCTGTCGGCCGAGTGCCAGCTGGTGACGGTAGTGCCCGT
 GGTCCCAGGGCTGCTGGAGGTCACCACACAGAATGTATACTTCTACGATGGCAGCACTGAGC
 GCGTGGAACCCGAGGAGGGCATCGGCTATGATTTCCGGCGCCCACTGGCCCAGCTGCGTGAG
 GTCCACCTGCGGCGTTTCAACCTGCGCCGTTTCAGCACTTGAGCTCTTCTTTATCGATCAGGC
 CAACTACTTCTCAACTTCCCATGCAAGGTGGGCACGACCCAGTCTCATCTCCTAGCCAGA
 25 CTCCGAGACCCAGCCTGGCCCCATCCCACCCCATACCCAGGTACGGAACCAGGTGTACTCG
 TGGCTCCTGCGCCTACGGCCCCCTCTCAAGGCTACCTAAGCAGCCGCTCCCCCAGGAGAT
 GCTGCGTGCCCTCAGGCCTTACCCAGAAATGGGTACAGCGTGAGATATCCAACCTTCGAGTACT
 TGATGCAACTCAACACCATTGCGGGGCGGACCTACAATGACCTGTCTCAGTACCCTGTGTTT
 CCCTGGGTCTGTCAGGACTACGTGTCCCAACCCCTGGACCTCAGCAACCCAGCCGTCTTCCG
 30 GGACCTGTCTAAGCCCATCGGTGTGGTGAACCCCAAGCATGCCAGCTCGTGAGGGAGAAGT
 ATGAAAGCTTTGAGGACCCAGCAGGGACCATTGACAAGTTCCACTATGGCACCCACTACTCC
 AATGCAGCAGGCGTGATGCACTACCTCATCCGCGTGGAGCCCTTCACCTCCCTGCACGTCCA
 GCTGCAAAGTGGCCGCTTTGACTGCTCCGACCGGCAGTTCCACTCGGTGGCGGCAGCCTGGC
 AGGCACGCCTGGAGAGCCCTGCCGATGTGAAGGAGCTCATCCCGGAATTCTTCTACTTTCT
 35 GACTTCCTGGAGAACCAGAACGGTTTTGGACCTGGGCTGTCTCCAGCTGACCAACGAGAAGGT
 AGGCGATGTGGTGCTACCCCCGTGGGCCAGCTCTCCTGAGGACTTCATCCAGCAGCACCGCC
 AGGCTCTGGAGTCGGAGTATGTGTCTGCACACCTACACGAGTGGATCGACCTCATCTTTGGC
 TACAAGCAGCGGGGGCCAGCCGCCGAGGAGGCCCTCAATGTCTTCTATTACTGCACCTATGA
 GGGGGCTGTAGACCTGGACCATGTGACAGATGAGCGGGAACGGAAGGCTCTGGAGGGCATTA
 40 TCAGCAACTTTGGGCAGACTCCCTGTCAGCTGCTGAAGGAGCCACATCCAACCTCGGCTCTCA
 GCTGAGGAAGCAGCCCATCGCCTTGCACGCCTGGACACTAACTCACCTAGCATCTTCCAGCA
 CCTGGACGAACTCAAGGCATTCTTCGAGAGGTGACTGTGAGTGCCAGTGGGCTGCTGGGCA
 CCCACAGCTGGTTGCCCTATGACCGCAACATAAGCAACTACTTCAGCTTCAGCAAAGACCCC
 ACCATGGGCAGCCACAAGACGCAGCGACTGCTGAGTGGCCCGTGGGTGCCAGGCAGTGGTGT
 45 GAGTGGACAAGCACTGGCAGTGGCCCCGGATGGAAAGCTGCTATTAGCGGTGGCCACTGGG
 ATGGCAGCCTGCGGGTGACTGCACTACCCCGTGGCAAGCTGTTGAGCCAGCTCAGCTGCCAC
 CTTGATGTAGTAACCTGCCTTGCCTTGGACACCTGTGGCATCTACCTCATCTCAGGCTCCCC

FIGURE 3B

5 GGACACCACGTGCATGGTGTGGCGGCTCCTGCATCAGGGTGGTCTGTCAGTAGGCCTGGCAC
CAAAGCCTGTGCAGGTCCTGTATGGGCATGGGGCTGCAGTGAGCTGTGTGGCCATCAGCACT
GAACTTGACATGGCTGTGTCTGGATCTGAGGATGGAAGTGTGATCATACACACTGTACGCCG
CGGACAGTTTGTAGCGGCACTACGGCCTCTGGGTGCCACATTCCCTGGACCTATTTTCCACC
TGGCATTGGGGTCCGAAGGCCAGATTGTGGTACAGAGCTCAGCGTGGGAACGTCCTGGGGCC
10 CAGGTACCTACTCCTTGCACCTGTATTTCAGTCAATGGGAAGTTGCGGGCTTCACTGCCCCCT
GGCAGAGCAGCCTACAGCCCTGACGGTGACAGAGGACTTTGTGTTGCTGGGCACCGCCCAGT
GCGCCCTGCACATCCTCCAACATAACACACTGCTCCCGGCCGCGCCTCCCTTGCCCATGAAG
GTGGCCATCCGCAGCGTGGCCGTGACCAAGGAGCGCAGCCACGTGCTGGTGGGCCTGGAGGA
TGGCAAGCTCATCGTGGTGGTTCGCGGGGCAGCCCTCTGAGGTGCGCAGCAGCCAGTTCGCGC
15 GGAAGCTGTGGCGGTCCTCGCGGCGCATCTCCCAGGTGTCCTCGGGAGAGACGGAATACAAC
CCTACTGAGGCGCGCT**TGA**ACCTGGCCAGTCCGGCTGCTCGGGCCCCGCCCCGGCAGGCCTG
GCCCCGGGAGGCCCCGCCCAGAAGTCGGCGGGAACACCCCGGGGTGGGCAGCCAGGGGGTGA
GCGGGGCCCCACCCTGCCCAGCTCAGGGATTGGCGGGCGATGTTACCCCCCTCAGGGATTGGCG
GGCGGAAGTCCCGCCCCCTCGCCGGCTGAGGGGCCGCCCTGAGGGCCAGCACTGGCGTCT

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FIGURE 4

5 MSQFEMDTYAKSHDLMSGFWNACYDMLMSSGQRRQWERAQSRRAFQELVLEPAQRRARLEGL
RYTAVLKQQATQHSMALLHWGALWRQLASPCGAWALRDTPIPRWKLSSAETYSRMRLKLVPN
HHFDPHLEASALRDNLGEVPLTPTEEASLPLAVTKEAKVSTPPELLQEDQLGEDELAELETP
MEAAELDEQREKLVLSAECQLVTVVAVVPGLLEVTTQNVYFYDGSTERVETEEGIGYDFRRP
LAQLREVHLRRFNLRRSALELFFIDQANYFLNFPCKVGTTPVSSPSQTPRPQPGPIPPHTQV
10 RNQVYSWLLRLRPPSQGYLSSRSPQEMLRASGLTQKWVQREISNFEYLMQLNTIAGRITYNDL
SQYPVFPWVLQDYVSPITLDLSNPAVFRDLSKPIGVVNPKHAQLVREKYESFEDPAGTIDKFH
YGTHYSNAAGVMHYLIRVEPFTSLHVQLQSGRFDCSDRQFHSVAAAWQARLESPADVKEIP
EFFYFPDFLENQNGFDLGCLQLTNEKVGDVVLPWASSPEDFIQQHRQALESEYVSAHLHEW
IDLIFGYKQRGPAEEALNVFYYCTYEGAVDLDHVTDERERKALEGII SNFGQTPCQLLKEP
15 HPTRLSAEEAAHRLARLDTNSPSIFQHLDELKAFFAEVTVSASGLLGTHSWLPYDRNISNYF
SFSKDPTMGSHKTQRLLSGPWVPGSGVSGQALAVAPDGKLLFSGGHWDGSLRVTALPRGKLL
SQLSCHLDVVTCLALDTCGIY LISGRDTCMVWRLLHQGGLSVGLAPKPVQVLYGHGA AVS
CVAISTELDMAVSGSEDGTVIIHTVRRGQFVAALRPLGATFPGPFIHFLALGSEGQIVVQSSA
WERPGAQVTYSLHLYSVNGKLRASLPLAEQPTALTVTEDFVLLGTAQCALHILQLNTLLPAA
20 PPLPMKVAIRSVAVTKERSHVLVGLEDGKLIVVVAGQPSEVRSSQFARKLWRSSRRISQVSS
GETEYNPTAR

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FIGURE 5

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CGGACGCGTGGGCGGACGCGTGGGCGGACGCGTGGGGCCGGCTTGGCTAGCGCGCGGCGGCC
GTGGCTAAGGCTGCTACGAAGCGAGCTTGGGAGGAGCAGCGGCCTGCGGGGCAGAGGAGCAT
CCCGTCTACCAGGTCCCAAGCGGCGTGGCCCGCGGGTCATGGCCAAAGGAGAAGGCGCCGAG
AGCGGCTCCGCGGCGGGGCTGCTACCCACCAGCATCCTCCAAAGCACTGAACGCCCCGGCCCA
10 GGTGAAGAAAGAACCGAAAAAGAAACAACAGTTGTCTGTTTGCAACAAGCTTTGCTATG
CACTTGGGGGAGCCCCCTACCAGGTGACGGGCTGTGCCCTGGGTTTCTTCCTTCAGATCTAC
CTATTGG**ATGT**GCGCTCAGGTGGGCCCTTTCTCTGCCTCCATCATCCTGTTTGTGGGCCGAGC
CTGGGATGCCATCACAGACCCCCCTGGTGGGCCCTCTGCATCAGCAAATCCCCCTGGACCTGCC
TGGGTGCGCTTATGCCCTGGATCATCTTCTCCACGCCCTGGCCGTCATTGCCCTACTTCCTC
15 ATCTGGTTCGTGCCCCGACTTCCACACGGCCAGACCTATTGGTACCTGCTTTTCTATTGCCCT
CTTTGAAACAATGGTCACGTGTTTCCATGTTCCCTACTCGGCTCTCACCATGTTTCATCAGCA
ACCGAGCAGACTGAGCGGGATTCTGCCACCGCCTATCGGATGACTGTGGAAGTGCTGGGCAC
AGTGCTGGGCACGGCGATCCAGGGACAAATCGTGGGCCAAGCAGACACGCCTTGTTTCCAGG
ACTTCAATAGCTCTACAGTAGCTTCACAAAGTGCCAACCATAACATGGCACCACTTCACAC
20 AGGGAAACGCAAAAGGCATACCTGCTGGCAGCGGGGGTCATTGTCTGTATCTATATAATCTG
TGCTGTCATCCTGATCCTGGGCGTGCGGGAGCAGAGAGAACCCTATGAAGCCCAGCAGTCTG
AGCCAATCGCCTACTTCCGGGGCCCTACGGCTGGTCATGAGCCACGGCCCATACATCAAACCTT
ATTACTGGCTTCTCTTCACCTCCTTGGCTTTTCATGCTGGTGGAGGGGAACCTTTGTCTTGTT
TTGCACCTACACCTTGGGCTTCCGCAATGAATTCCAGAATCTACTCCTGGCCATCATGCTCT
25 CGGCCACTTTAACCATTCCCATCTGGCAGTGGTTCTTGACCCGGTTTGGCAAGAAGACAGCT
GTATATGTTGGGATCTCATCAGCAGTGCCATTTCTCATCTTGGTGGCCCTCATGGAGAGTAA
CCTCATCATTAACATATGCGGTAGCTGTGGCAGCTGGCATCAGTGTGGCAGCTGCCTTCTTAC
TACCCTGGTCCATGCTGCCTGATGTCATTGACGACTTCCATCTGAAGCAGCCCCACTTCCAT
GGAACCGAGCCCATCTTCTTCTCCTTCTATGTCTTCTTCACCAAGTTTGCCTCTGGAGTGTC
30 ACTGGGCATTTCTACCCCTCAGTCTGGACTTTGCAGGGTACCAGACCCGTGGCTGCTCGCAGC
CGGAACGTGTCAAGTTTACACTGAACATGCTCGTGACCATGGCTCCCATAGTTCTCATCCTG
CTGGGCCTGCTGCTCTTCAAAATGTACCCCATTGATGAGGAGAGGCGGCGGCAGAAATAAGAA
GGCCCTGCAGGCACTGAGGGACGAGGCCAGCAGCTCTGGCTGCTCAGAAACAGACTCCACAG
AGCTGGCTAGCATCCTC**TAG**GGCCCGCCACGTTGCCCGAAGCCACCATGCAGAAGGCCACAG
35 AAGGGATCAGGACCTGTCTGCCGGCTTGCTGAGCAGCTGGACTGCAGGTGCTAGGAAGGGAA
CTGAAGACTCAAGGAGGTGGCCCAGGACACTTGCTGTGCTCACTGTGGGGCCGGCTGCTCTG
TGGCCTCCTGCCTCCCCCTCTGCCTGCCTGTGGGGCCAAGCCCTGGGGCTGCCACTGTGAATA
TGCCAAGGACTGATCGGGCCTAGCCCGGAACACTAATGTAGAAACCTTTTTTTTACAGAGCC
TAATTAATAACTTAATGACTGTGTACATAGCAATGTGTGTGTATGTATATGTCTGTGAGCTA
40 TTAATGTTATTAATTTTCATAAAAGCTGGAAAGC

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FIGURE 6

5	Signal Peptide:	amino acids 1-17
	Transmembrane Domains:	amino acids 171-190
		amino acids 220-239
		amino acids 259-275
		amino acids 286-305
10		amino acids 316-335
		amino acids 353-378
		amino acids 396-417
	N-glycosylation Sites:	amino acids 145-147
		amino acids 155-158

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MWLRWALSLPPSSCLWAEPMPSQTPWWASASANPPGPAWVALCPGSSSPRPWPSLPTSSSG
 SCPTSHTARPIGTCTFSIASLKQWSRVSMFPTRLSPCSSATEQTERDSATAYRMTVEVLGTVL
 20 GTAIQGQIVGQADTPCFQDFNSSTVASQSANTHGTTSHTRETQKAYLLAAGVIVCIYIICAV
 ILILGVREQREPYEAQQSEPIAYFRGLRLVM SHGPYIKLITGFLFTSLAFMLVEGNFVLFCT
 YTLGFRNEFQNLLLAIMLSATLTPIWQWFLTRFGKKTAVYVGISSAVPFLILVALMESNLI
 ITYAVAVAAGISVAAAFLLPWSMLPDVIDDFHLKQPHFHGTETPIFFSFYVFFTKFASGVSLG
 ISTLSLDFAGYQTRGCSQPERVKFTLNMLVTMAPIVLILLGLLLFKMYPIDEERRRQNKAL
 25 QALRDEASSSGCSETDSTELASIL

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FIGURE 7

5 CCGCGGGAACGCTGTCCTGGCTGCCGCCACCCGAACAGCCTGTCCTGGTGCCCCGGCTCCCT
GCCCCGCGCCCAGTCA**ATG**ACCCTGCGCCCCCTCACTCCTCCCGCTCCATCTGCTGCTGCTGCT
GCTGCTCAGTGCGGCGGTGTGCCGGGCTGAGGCTGGGCTCGAAACCGAAAGTCCCGTCCGGA
CCCTCCAAGTGGAGACCCTGGTGGAGCCCCCAGAACCATGTGCCGAGCCCGCTGCTTTTGGA
10 GACACGCTTCACATACTACACGGGAAGCTTGGTAGATGGACGTATTATTGACACCTCCCT
GACCAGAGACCCTCTGGTTATAGAACTTGGCCAAAAGCAGGTGATTCCAGGTCTGGAGCAGA
GTCTTCTCGACATGTGTGTGGGAGAGAAGCGAAGGGCAATCATTCTTCTCACTTGGCCTAT
GGAAAACGGGGATTTCCACCATCTGTCCCAGCGGATGCAGTGGTGCAGTATGACGTGGAGCT
GATTGCACTAATCCGAGCCAACTACTGGCTAAAGCTGGTGAAGGGCATTTTGCCTCTGGTAG
15 GGATGGCCATGGTGGCCAGCCCTCCTGGGCCTCATTGGGTATCACCTATACAGAAAGGCCAAT
AGACCCAAAGTCTCCAAAAAGAAGCTCAAGGAAGAGAAACGAAACAAGAGCAAAAAGAA**TA**
ATAAATAATAAATTTTAAAAAACTTAAAAAAAAAAAAAAAAAAAA

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FIGURE 8

5	Signal Peptide:	amino acids 1-25
	Transmembrane Domain	amino acids 155-174
	N-glycosylation Site:	amino acids 196-199
	FKBP-type peptidyl-prolyl	
	cis-trans isomer Signature Sequences:	amino acids 62-77
10		amino acids 87-123
		amino acids 128-141

MTLRPSLLPLHLLLLLLLLSAAVCRAEAGLETESPVRTLQVETLVEPPEPCAEPAAFGDTLHI
15 HYTGSLVDGRIIDTSLTRDPLVIELGQKQVIPGLEQSLDMCVGEKRRAIIPSHLAYGKRGF
PPSVPADAVVQYDVELIALIRANYWLKLVKGILPLVGMAMVPALLGLIGYHLYRKANRPKVS
KKKLKEEKRNSKKK

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FIGURE 9

5 CTTCCCAGCCCTGTGCCCCAAAGCACCTGGAGCATATAGCCTTGCAGAACTTCTACTTGCCT
GCCTCCCTGCCTCTGGCC**ATG**GCCTGCCGGTGCCTCAGCTTCCTTCTGATGGGGACCTTCCT
GTCAGTTTCCCAGACAGTCCTGGCCCAGCTGGATGCACTGCTGGTCTTCCCAGGCCAAGTGG
CTCAACTCTCCTGCACGCTCAGCCCCCAGCACGTCACCATCAGGGACTACGGTGTGTCCTGG
10 TACCAGCAGCGGGCAGGCAGTGCCCCCTCGATATCTCCTCTACTACCGCTCGGAGGAGGATCA
CCACCGGCCTGCTGACATCCCCGATCGATTCTCGGCAGCCAAGGATGAGGCCACAAATGCCT
GTGTCCTCACCATTAGTCCCGTGCAGCCTGAAGACGACGCGGATTACTACTGCTCTGTTGGC
TACGGCTTTAGTCCC**TAG**GGGTGGGGTGTGAGATGGGTGCCTCCCCTCTGCCTCCCATTCT
GCCCCTGACCTTGGGTCCCTTTTAAACTTTCTCTGAGCCTTGCTTCCCCTCTGTAAAATGGG
15 TTAATAATATTCAACATGTCAACAAC

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FIGURE 10

5 Signal Peptide: amino acids 1-20

MACRCLSFLLMGTFLSVSQTVLAQLDALLVFPQVAQLSCTLSPQHVTIRDYGVSWYQQRAG
SAPRYLLYYRSEEDHHRPADIPDRFSAKDEAHNACVLTISPVQPEDDADYYCSVGYGFS

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FIGURE 11

5 GTGGAGGCCGCCGACG**ATG**GCGGGGCCGACGGAGGCCGAGACGGGGTTGGCCGAGCCCCGGG
CCCTGTGCGCGCAGCGGGGCCACCGCACCTACGCGCGCCGCTGGGTGTTCTGCTCGCGATC
AGCCTGCTCAACTGCTCCAACGCCACGCTGTGGCTCAGCTTTGCACCTGTGGCTGACGTCAT
TGCTGAGGACTTGGTCCTGTCCATGGAGCAGATCAACTGGCTGTCACTGGTCTACCTCGTGG
10 TATCCACCCCATTGTCGTGGCGGCCATCTGGATCCTGGACTCCGTCGGGCTCCGTGCGGCG
ACCATCCTGGGTGCGTGGCTGAACTTTGCCGGGAGTGTGCTACGCATGGTGCCCTGCATGGT
TGTTGGGACCCAAAACCCATTGTCCTTCCTCATGGGTGGCCAGAGCCTCTGTGCCCTTGCCC
AGAGCCTGGTCATCTTCTCTCCAGCCAAGCTGGCTGCCTTGTGGTTCCCAGAGCACCAGCGA
GCCACGGCCAACATGCTCGCCACCATGTCGAACCCTCTGGGCGTCCTTGTGGCCAATGTGCT
15 GTCCCCCTGTGCTGGTCAAGAAGGGTGAGGACATTCCGTTAATGCTCGGTGTCTATAACCATCC
CTGCTGGCGTCGTCTGCCTGCTGTCCACCATCTGCCTGTGGGAGAGTGTGCCCCCACC
CCCTCTGCCGGGGCTGCCAGCTCCACCTCAGAGAAGTTCTTGGATGGGCTCAAGCTGCAGCT
CATGTGGAACAAGGCCATATGTCATCCTGGCTGTGTGCTTGGGGGGAATGATCGGGATCTCTG
CCAGCTTCTCAGCCCTCCTGGAGCAGATCCTCTGTGCAAGCGGCCACTCCAGTGGGTTTTCC
20 GGCCTCTGTGGCGCTCTCTTCATCACGTTTGGGATCCTGGGGGCACTGGCTCTCGGCCCTTA
TGTGGACCGGACCAAGCACTTCACTGAGGCCACCAAGATTGGCCTGTGCCTGTTCTCTCTGG
CCTGCGTGCCCTTTGCCCTGGTGTCCCAGCTGCAGGGACAGACCCTTGCCCTGGCTGCCACC
TGCTCGCTGCTCGGGCTGTTTGGCTTCTCGGTGGGCCCCGTGGCCATGGAGTTGGCGGTGCA
GTGTTCTTCCCCGTGGGGGAGGGGGCTGCCACAGGCATGATCTTTGTGCTGGGGCAGGCCG
25 AGGGAATACTCATCATGCTGGCAATGACGGCACTGACTGTGCGACGCTCGGAGCCGTCCTTG
TCCACCTGCCAGCAGGGGGAGGATCCACTTGACTGGACAGTGTCTCTGCTGCTGATGGCCGG
CCTGTGCACCTTCTTCAGCTGCATCCTGGCGGTCTTCTTCCACACCCCATACCGGCGCCTGC
AGGCCGAGTCTGGGGAGCCCCCTCCACCCGTAACGCCGTGGGCGGCGCAGACTCAGGGCCG
GGTGTGGACCGAGGGGGAGCAGGAAGGGCTGGGGTCCTGGGGCCCAGCACGGCGACTCCGGA
30 GTGCACGGCGAGGGGGGCTCGCTAGAGGACCCAGAGGGCCCCGGGAGCCCCACCCAGCCT
GCCACCGAGCGACTCCCCGTGCGCAAGGCCAGCAGCCACCGACGCGCCCTCCCGCCCCGGC
AGACTCGCAGGCAGGGTCCAAGCGTCCAGGTTTATTGACCCGGCTGGGTCTCACTCCTCCTT
CTCCTCCCCGTGGGTGATCACG**TAG**CTGAGCGCCTTGTTAGTCCAGGTTGCCCGCCACATCGA
TGGAGGCGAACTGGAACATCTGGTCCACCTGCGGGCGGGGGCGAAAGGGCTCCTTGCGGGCT
35 CCGGGAGCGAATTACAAGCGCGCACCTGAAAA

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FIGURE 12

5 Transmembrane Domains: amino acids 30-50
amino acids 61-79
amino acids 98-112
amino acids 126-146
amino acids 169-182
10 amino acids 201-215
amino acids 248-268
amino acids 280-300
amino acids 318-337
amino acids 341-357
15 amino acids 375-387
amino acids 420-441
N-glycosylation Sites: amino acids 40-43
amino acids 43-46
Glycosaminoglycan Attachment Site: amino acids 468-471
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MAGPTEAETGLAEPRALCAQRGHRITYARRWVFLLAISLLNCSNATLWLSFAPVADVIAEDLV
LSMEQINWLSLVYLVVSTPFGVAAIWILDSVGLRAATILGAWLNFAGSVLRMVPCMVVGTON
PFAFLMGGQSLCALAQSLVIFSPAKLAALWFPEHQ RATANMLATMSNPLGVLVANVLSPLV
25 KKGEDIPLMLGVYTIPAGVVCLLSTICLWESVPPTPPSAGAASSTSEKFLDGLKLQLMWNKA
YVILAVCLGGMIGISASFSALLEQILCASGHSSGFSGLCGALFITFGILGALALGPYVDR TK
HFTEATKIGLCLFSLACVPFALVSQ LQGQTLALAATCSLLGLFGFSVGPVAMELAVECSFPV
GEGAATGMIFVLGQAEGILIMLMTALT VRRSEPSLSTCQQGEDPLDWTVSLLLMAGLCTFF
SCILAVFFHTPYRRLQAESGEPPSTRNAVGGADSGPGVDRGGAGRAGVLGPSTATPECTARG
30 ASLEDPRGPGSPHPACHRATPRAQGP AATDAPSRPGRLAGRVQASRFIDPAGSHSSFSSPWVIT

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FIGURE 13

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GGCACGAGCCAGGAACTAGGAGGTTCTCACTGCCCCGAGCAGAGGCCCTACACCCACCGAGGC
ATGGGGCTCCCTGGGCTGTTCTGCTTGGCCGTGCTGGCTGCCAGCAGCTTCTCCAAGGCACG
GGAGGAAGAAATTACCCCTGTGGTCTCCATTGCCTACAAAGTCCTGGAAGTTTCCCCAAAG
GCCGCTGGGTGCTCATAACCTGCTGTGCACCCAGCCACCACCGCCCATCACCTATTCCCTC
10 TGTGGAACCAAGAACATCAAGGTGGCCAAGAAGGTGGTGAAGACCCACGAGCCGGCCTCCTT
CAACCTCAACGTCACACTCAAGTCCAGTCCAGACCTGCTCACCTACTTCTGCCGGGCGTCCT
CCACCTCAGGTGCCCATGTGGACAGTGCCAGGCTACAGATGCACTGGGAGCTGTGGTCCAAG
CCAGTGTCTGAGCTGCGGGCCAACTTCACTCTGCAGGACAGAGGGGCAGGCCCCAGGGTGGA
GATGATCTGCCAGGCGTCCTCGGGCAGCCCACCTATCACCAACAGCCTGATCGGGAAGGATG
15 GGCAGGTCCACCTGCAGCAGAGACCATGCCACAGGCAGCCTGCCAACTTCTCCTTCCCTGCCG
AGCCAGACATCGGACTGGTTCTGGTGCCAGGCTGCAAACAACGCCAATGTCCAGCACAGCGC
CCTCACAGTGGTGCCCCCAGGTGGTGACCAGAAGATGGAGGACTGGCAGGGTCCCCTGGAGA
GCCCCATCCTTGCCCTTGCCGCTCTACAGGAGCACCCGCCGTCTGAGTGAAGAGGAGTTTGGG
GGGTTCAGGATAGGGAATGGGGAGGTCAGAGGACGCAAAGCAGCAGCCATG**TAGA**ATGAACC
20 GTCCAGAGAGCCAAGCACGGCAGAGGACTGCAGGCCATCAGCGTGCACTGTTTCGTATTTGGA
GTTTCATGCAAAATGAGTGTGTTTTAGCTGCTCTTGCCACAAAAAAAAAAAAAAAAAAAAA

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FIGURE 14

5 Signal Peptide: amino acids 1-18
N-glycosylation Sites: amino acids 86-89
amino acids 132-135
amino acids 181-184

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MGLPGLFCLAVLAASSFSKAREEEITPVVSIAYKVLEVFPKGRWVLITCCAPQPPPPITYSL
CGTKNIKVAKKVVKTHEPASFNLNVTLKSSPDLLTYFCRASSTSGAHVDSARLQMHWELWSK
PVSELNANFTLQDRGAGPRVEMICQASSGSPITNSLIGKDGQVHLQQRPCRPANFSFLP
15 SQTSDWFWCQAANNANVQHSALTVPVPPGGDQKMEDWQGPLESFILALPLYRSTRRLSEEEFG
GFRIGNGEVRGRKAAAM

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FIGURE 15

5 CTGCAGGTAGACATCTCCACTGCCCAGGAATCACTGAGCGTGCAGACAGCACAGCCTCCTCT
GAAGGCCGGCCATAACCAGAGTCCTGCCTCGGCATGGGCCTCACCATTGAGGCAGCTCCACTG
TCTGTGCTGGTCTGAGGGTGCTGCCTGTC**ATG**GGGGGCAGCCATCTCCCAGGGGGGCCCTCATC
GCCATCGTCTGCAACGGTCTCGTGGGCTTCTTGCTGCTGCTGCTCTGGGTGCATCCTCTGCTG
10 GGCCTGCCATTCTCGTCTGCCGACGTTGACTCTCTCTCTGAATCCAGTCCCAACTCCAGCCC
TGGCCCCTGTCCTGAGAAGGCCCCACCACCCCAGAAGCCCAGCCATGAAGGCAGCTACCTGC
TGCAGCCCCTGAAGGCCCTGGCCTAGCCTGGAGCCCAGGACC**TAA**GTCCACCTCACCTAGAG
CCTGGAATTAGGATCCCAGAGTTCAGCCAGCCTGGGGTCCAGAACTCAAGAGTCCGCCTGCT
TGGAGCTGGACCCAGCGGCCAGAGTCTAGCCAGCTTGGCTCCAATAGGAGCTCAGTGGCCC
15 TAAGGAGATGGGCCTGGGGTGGGGGCTTATGAGTTGGTGCTAGAGCCAGGGCCATCTGGACT
ATGCTCCATCCAAGGGCCAAGGGTCAGGGGCCGGGTCCACTCTTTCCCTAGGCTGAGCACC
TCTAGGCCCTCTAGGTGGGGAAGCAAACCTGGAACCCATGGCAATAATAGGAGGGTGTCCAG
GCTGGGGCCCCTCCCCTGGTCCTCCAGTGTTTGCTGGATAATAAATGGAACCTATGGCTCTAA
AAAAAAAAAAAAAAAAAAAA

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FIGURE 16

5 Signal Peptide: amino acids 1-33
N-myristoylation Sites: amino acids 2-7
amino acids 8-13
Thioredoxin family of
proteins homology block: amino acids 23-39

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MGA AISQ GALIA IVCNGLVGFL LLL L WILCWACHSRLPTLTLSLNPVPTPALAPVLRRPHH
PRSPAMKAATCCSPEGPWPSLEPRT

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FIGURE 17

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AAACTCAGCACTTGCCGGAGTGGCTCATTGTTAAGACAAAGGGTGTGCACTTCCTGGCCAGG
AAACCTGAGCGGTGAGACTCCCAGCTGCCTACATCAAGGCCCCAGGACATGCAGAACCTTCC
TCTAGAACCCGACCCACCACC**ATG**AGGTCTGCCTGTGGAGATGCAGGCACCTGAGCCAAGG
CGTCCAGTGGTTCCTTGCTTCTGGCTGTCTTGGTCTTCTTTCTCTTCGCCTTGCCCTCTTTTA
10 TTAAGGAGCCTCAAACAAAGCCTTCCAGGCATCAACGCACAGAGAACATTAAAGAAAGGTCT
CTACAGTCCCTGGCAAAGCCTAAGTCCCAGGCACCCACAAGGGCGAGGAGGACAACCATCTA
TGCAGAGCCAGCGCCAGAGAACAATGCCCTCAACACACAAACCCAGCCCAAGGCCACACCA
CCGGAGACAGAGGAAAGGAGGCCAACCCAGGCACCGCCGGAGGAGCAGGACAAGGTGCCCCAC
ACAGCACAGAGGGCAGCATGGAAGAGCCAGAAAAAGAGAAAACCATGGTGAACACACTGTC
15 ACCCAGAGGGCAAGATGCAGGGATGGCCTCTGGCAGGACAGAGGCACAATCATGGAAGAGCC
AGGACACAAAGACGACCCAAGGAAATGGGGGCCAGACCAGGAAGCTGACGGCCTCCAGGACG
GTGTCAGAGAAGCACCAGGGCAAAGCGGCAACCACAGCCAAGACGCTCATTTCCAAAAGTCA
GCACAGAATGCTGGCTCCCACAGGAGCAGTGTCAACAAGGACGAGACAGAAAGGAGTGACCA
CAGCAGTCATCCACCTAAGGAGAAGAAACCTCAGGCCACCCACCCCCCTGCCCTTTCCAG
20 AGCCCCACGACGCAGAGAAACCAAAGACTGAAGGCCGCCAACTTCAAATCTGAGCCTCGGTG
GGATTTTGAGGAAAAATACAGCTTCGAAATAGGAGGCCTTCAGACGACTTGCCCTGACTCTG
TGAAGATCAAAGCCTCCAAGTCGCTGTGGCTCCAGAACTCTTTCTGCCCAACCTCACTCTC
TTCTTGACTCCAGACACTTCAACCAGAGTGAGTGGGACCGCCTGGAACACTTTGCACCACC
CTTTGGCTTCATGGAGCTCAACTACTCCTTGGTGCAGAAGGTCGTGACACGCTTCCCTCCAG
25 TGCCCCAGCAGCAGCTGCTCCTGGCCAGCCTCCCCGCTGGGAGCCTCCGGTGCATCACCTGT
GCCGTGGTGGGCAACGGGGGCATCCTGAACAACTCCCACATGGGCCAGGAGATAGACAGTCA
CGACTACGTGTTCCGATTGAGCGGAGCTCTCATTAAGGCTACGAACAGGATGTGGGGACTC
GGACATCCTTCTACGGCTTTACCGCCTTCTCCCTGACCCAGTCACTCCTTATATTGGGCAAT
CGGGGTTTCAAGAACGTGCCTCTTGGGAAGGACGTCCGCTACTTGCACCTTCTGGAAGGCAC
30 CCGGGACTATGAGTGGCTGGAAGCACTGCTTATGAATCAGACGGTGATGTCAAAAAACCTTT
TCTGGTTTCAGGCACAGACCCCAGGAAGCTTTTCGGGAAGCCCTGCACATGGACAGGTACCTG
TTGCTGCACCCAGACTTTCTCCGATACATGAAGAACAGGTTTCTGAGGTCTAAGACCCTGGA
TGGTGCCCACTGGAGGATATACCGCCCCACCACTGGGGCCCTCCTGCTGCTCACTGCCCTTC
AGCTCTGTGACCAGGTGAGTGCTTATGGCTTCATCACTGAGGGCCATGAGCGCTTTTCTGAT
35 CACTACTATGATACATCATGGAAGCGGCTGATCTTTTACATAAACCATGACTTCAAGCTGGA
GAGAGAAGTCTGGAAGCGGCTACACGATGAAGGGATAATCCGGCTGTACCAGCGTCCTGGTC
CCGGAAGTGCCAAAGCCAAGAACT**TGA**CCGGGGCCAGGGCTGCCATGGTCTCCTTGCCTGCTC
CAAGGCACAGGATACAGTGGGAATCTTGAGACTCTTTGGCCATTTCCCATGGCTCAGACTAA
GCTCCAAGCCCTTCAGGAGTTCCAAGGGAACACTTGAACCATGGACAAGACTCTCTCAAGAT
40 GGCAAATGGCTAATTGAGGTTCTGAAGTTCTTCAGTACATTGCTGTAGGTCTTGAGGCCAGG
GATTTTTTAATTAAATGGGGTGATGGGTGGCCAATACCACAATTCCTGCTGAAAAACACTCTT
CCAGTCCAAAAGCTTCTTGATACAGAAAAAGAGCCTGGATTACAGAAACATATAGATCTG
GTTTGAATTCCAGATCGAGTTTACAGTTGTGAAATCTTGAAGGTATTACTTAACTTCACTAC
AGATTGTCTAGAAGACCTTTCTAGGAGTTATCTGATTCTAGAAGGGTCTATACTTGTCTTGG
45 TCTTTAAGCTATTTGACAACCTCTACGTGTTGTAGAAAACCTGATAATAATACAAATGATTGTT
GTCCATGGAAAGGCCAAATAAATTTCTACAGTGAAAAA

FIGURE 18

5	Cytoplasmic Domain:	amino acids 1-10
	Type II Transmembrane Domain:	amino acids 11-35
	Lumenal catalytic Domain:	amino acids 36-600
	Ribonucleotide Reductase	
	small subunit Signature:	amino acids 481-496
10	N-glycosylation Sites:	amino acids 300-303
		amino acids 311-314
		amino acids 331-334
		amino acids 375-378
		amino acids 460-463

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MRSCLWRCRHL SQGVQWSLLLAVLVFFLFALPSFIKEPQTKPSRHQRTENIKERSLQSLAKP
 KSQAPTRARRTTIYAEPAPENNALNTQTQPKAHTTGDRGKEANQAPPEEQDKVPHTAQRAAW
 20 KSPEKEKTMVNTLSPRGQDAGMASGRTEAQSWSQDTKTTQGNGGQTRKLTASRTVSEKHQG
 KAATTAKTLIPKSQHRMLAPTGA VSTRTRQKGVTTAVIPPKEKKPQATPPPAPFQSPTTQRN
 QRLKAANFKSEPRWDFEEKYSFEIGGLQTTCPDSVKIKASKSLWLQKLFLPNLTLFLDSRHF
 NQSEWDRLEHFAPPPFGFMELNYSLVQKV VTRFPVPVQQQLLLASLPAGSLRCITCAVVGNGG
 ILNNSHMGQEIDSHDYVFRLSGALIKGYEQDVGTRTSFYGF TAFSLTQSL LILGNRGFKNVP
 25 LGKDVRYLHFLEGTRDYEWLEALLMNQTVMSKNLFWFRHRPQEAFREALHMDRYLLLHPDFL
 RYMKNRFLRSKTL DGAHWRIYRPTTGALLLLTALQLCDQVSAYGFITEGHERFSDHYDTSW
 KRLIFYINHDFKLEREVWKRLHDEGIIRLYQRPGPGTAKAKN

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FIGURE 19A

5
GCCGCCCCGCCCCGAGACCGGGCCCCGGGGCGCGGGGCGGCGGGATGCGGCGCCCCGGGGCGG
CGATGACCGCGGAGCGCACGCCGCGGGCCCCGGCCCTGACCCCGCCGCCCCGCTGAGCCC
CCCGCCGAGGTCCGGACAGGCCGAGATGACGCCGAGCCCCCTGTTGCTGCTCCTGCTGCCGC
CGCTGCTGCTGGGGGCCTTCCCACCGGCCGCGCCGCCCCGAGGCCCCCAAAGATGGCGGAC
10 AAGGTGGTCCACGGCAGGTGGCCCCGCTGGGCGCAGTGTGCGGCTGCAGTGCCAGTGGA
GGGGGACCCGCGCCGCTGACCATGTGGACCAAGGATGGCCGCACCATCCACAGCGGCTGGA
GCCGCTTCCGCGTGCTGCCGCAGGGGCTGAAGGTGAAGCAGGTGGAGCGGGAGGATGCCGGC
GTGTACGTGTGCAAGGCCACCAACGGCTTCGGCAGCCTGAGCGTCAACTACACCCTCGTCGT
GCTGGATGACATTAGCCCAGGGAAGGAGAGCCTGGGGCCCCGACAGCTCCTCTGGGGGTCAAG
15 AGGACCCCGCCAGCCAGCAGTGGGCACGACCGCGCTTCACACAGCCCTCCAAGATGAGGCGC
CGGGTGATCGCACGGCCCGTGGGTAGCTCCGTGCGGCTCAAGTGCGTGGCCAGCGGGCACCC
TCGGCCCCGACATCACGTGGATGAAGGACGACCAGGCCCTTGACGCGCCCAGAGGCCGCTGAGC
CCAGGAAGAAGAAGTGGACACTGAGCCTGAAGAACCTGCGGCCGGAGGACAGCGGCAAATAC
ACCTGCCGCGTGTGCAACCGCGCGGGGCGCCATCAACGCCACCTACAAGGTGGATGTGATCCA
20 GCGGACCCGTTCCAAGCCCGTGCTCACAGGCACGCACCCCGTGAACACGACGGTGGACTTCG
GGGGGACCACGTCTTCCAGTGCAAGGTGCGCAGCGACGTGAAGCCGGTGATCCAGTGGCTG
AAGCGCGTGGAGTACGGCGCCGAGGGCCGCCACAACCTCCACCATCGATGTGGGCGGCCAGAA
GTTTGTGGTGCTGCCACGGGTGACGTGTGGTTCGCGGCCCGACGGCTCCTACCTCAATAAGC
TGCTCATCACCCGTGCCCCGCCAGGACGATGCGGGCATGTACATCTGCCTTGGCGCCAACACC
25 ATGGGCTACAGCTTCCGCAGCGCCTTCCCTCACCGTGCTGCCAGACCCAAAACCGCCAGGGCC
ACCTGTGGCCTCCTCGTCCTCGGCCACTAGCCTGCCGTGGCCCGTGGTCATCGGCATCCCAG
CCGGCGCTGTCTTCATCCTGGGCACCCTGCTCCTGTGGCTTTGCCAGGCCCAAGAAGCCG
TGACCCCCCGCGCCTGCCCTTCCCTGCCTGGGCACCGCCCGCCGGGGACGGCCCCGCGACCG
CAGCGGAGACAAGGACCTTCCCTCGTTGGCCGCCCTCAGCGCTGGCCCTGGTGTGGGGCTGT
30 GTGAGGAGCATGGGTCTCCGGCAGCCCCCAGCACTTACTGGGCCCAGGCCCAGTTGCTGGC
CCTAAGTTGTACCCCCAACTCTACACAGACATCCACACACACACACACACTCTCACAC
ACACTCACACGTGGAGGGCAAGGTCCACCAGCACATCCACTATCAGTGCTAGACGGCACCGT
ATCTGCAGTGGGCACGGGGGGGCGGCCAGACAGGCAGACTGGGAGGATGGAGGACGGAGCT
GCAGACGAAGGCAGGGGACCCATGGCGAGGAGGAATGGCCAGCACCCCAGGCAGTCTGTGTG
35 TGAGGCATAGCCCCCTGGACACACACACACAGACACACACACTACCTGGATGCATGTATGCAC
ACACATGCGCGCACACGTGCTCCCTGAAGGCACACGTACGCACACGCACATGCACAGATATG
CCGCTGGGCACACAGATAAGCTGCCCAAATGCACGCACACGCACAGAGACATGCCAGAACA
TACAAGGACATGCTGCCTGAACATACACACGCACACCCATGCGCAGATGTGCTGCCTGGACA
CACACACACACACGGATATGCTGTCTGGACGCACACACGTGCAGATATGGTATCCGGACACA
40 CACGTGCACAGATATGCTGCCTGGACACACAGATAATGCTGCCTTGACACACACATGCACGG
ATATTGCCTGGACACACACACACACACACACGCGTGACAGATATGCTGTCTGGACACGCACAC
ACATGCAGATATGCTGCCTGGACACACACTTCCAGACACACGTGCACAGGCGCAGATATGCT
GCCTGGACACACGCAGATATGCTGTCTAGTCACACACACACGCAGACATGCTGTCCGGACAC
ACACACGCATGCACAGATATGCTGTCCGGACACACACACGCACGCAGATATGCTGCCTGGAC
45 ACACACACAGATAATGCTGCCTCAACACTCACACACGTGCAGATATTGCCTGGACACACACA
TGTGCACAGATATGCTGTCTGGACATGCACACACGTGCAGATATGCTGTCCGGATACACACG
CACGCACACATGCAGATATGCTGCCTGGGCACACACTTCCGGACACACATGCACACACAGGT

FIGURE 19B

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GCAGATATGCTGCCTGGACACACACACAGATAATGCTGCCTCAACACTCACACACGTGCAGA
TATTGCCTGGACACACACATGTGCACAGATATGCTGTCTGGACATGCACACACGTGCAGATA
TGCTGTCCGGATACACACGCACGCACACATGCAGATATGCTGCCTGGGCACACACTTCCGGA
10 CACACATGCACACACAGGTGCAGATATGCTGCCTGGACACACGCAGACTGACGTGCTTTTGG
GAGGGTGTGCCGTGAAGCCTGCAGTACGTGTGCCGTGAGGCTCATAGTTGATGAGGGACTTT
CCCTGCTCCACCGTCACTCCCCCAACTCTGCCCGCCTCTGTCCCCGCCTCAGTCCCCGCCTC
CATCCCCGCCTCTGTCCCCTGGCCTTGGCGGCTATTTTTTGCCACCTGCCTTGGGTGCCCAGG
AGTCCCCTACTGCTGTGGGCTGGGGTTGGGGGCACAGCAGCCCCAAGCCTGAGAGGCTGGAG
15 CCCATGGCTAGTGGCTCATCCCCAGTGCATTCTCCCCCTGACACAGAGAAGGGGCCTTGGTA
TTTATATTTAAGAAATGAAGATAATATTAATAATGATGGAAGGAAGACTGGGTTCAGGGAC
TGTGGTCTCTCCTGGGGCCCCGGACCCGCCTGGTCTTTCAGCCATGCTGATGACCACACCCC
GTCCAGGCCAGACACCACCCCCCACCCTGTCGTGGTGGCCCCAGATCTCTGTAATTTTA
TGTAAGTTTGAGCTGAAGCCCCGTATATTTAATTTATTTTGTAAACACAAAA

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FIGURE 20

5	Signal Peptide:	amino acids 1-17
	Transmembrane Domain:	amino acids 376-396
	Tyrosine Kinase Phosphorylation Sites:	amino acids 212-219
		amino acids 329-336
	N-glycosylation Sites:	amino acids 111-114
10		amino acids 231-234
		amino acids 255-258
		amino acids 293-296
	Immunoglobulin and MHC protein sequence homology block	amino acids 219-136
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MTPSPLLLLLLPPLLLGAFPPAAAARGPPKMADKVVPVPRQVARLGRTVRLQCPVEGDPPPLTM
 WTKDGRTIHSGWSRFRVLPQGLKVKQVEREDAGVYVCKATNGFGSLSVNYTLVVLDDISPGK
 20 ESLGPDSSSGGQEDPASQQWARPRFTQPSKMRRRVVIARPVGSSVRLKCVASGHPRPDITWMK
 DDQALTRPEAAEPRKKKWTLSLKNLRPEDSGKYTCRVSNRAGAINATYKVDVIQRTSRKPV
 TGTHPVNTTVDFGGTTSFQCKVRSVDVKPVIQWLKRVEYGAEGRHNSTIDVGGQKFVVLPTGD
 VWSRPDGSYLNKLLITRARQDDAGMYICLGANTMGYSFRSAFLTVLPDPKPPGPPVASSSSA
 TSLPWPVVIGIPAGAVFILGTLLLWLCQAQKKPCTPAPAPPLPGHRPPGTARDRSGDKDLPS
 25 LAALSAGPGVGLCEEHGSPAAPQHLLGPGPVAGPKLYPKLYTDIHTHTHTSHTHSHVEGKV
 HQHIHYQC

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FIGURE 21

5 CCTCTGTCCACTGCTTTCGTGAAGACAAG**ATGA**AGTTCACAATTGTCTTTGCTGGACTTCTT
GGAGTCTTTCTAGCTCCTGCCCTAGCTAACTATAATATCAACGTCAATGATGACAACAACAA
TGCTGGAAGTGGGCAGCAGTCAGTGAGTGTCAACAATGAACACAATGTGGCCAATGTTGACA
ATAACAACGGATGGGACTCCTGGAATTCCATCTGGGATTATGGAAATGGCTTTGCTGCAACC
10 AGACTCTTTCAAAGAAGACATGCATTGTGCACAAAATGAACAAGGAAGTCATGCCCTCCAT
TCAATCCCTTGATGCACTGGTCAAGGAAAAGAAGCTTCAGGGTAAGGGACCAGGAGGACCAC
CTCCAAGGGCCTGATGTACTCAGTCAACCCAAACAAAGTCGATGACCTGAGCAAGTTCGGA
AAAAACATTGCAAACATGTGTTCGTGGGATTCCAACATACATGGCTGAGGAGATGCAAGAGGC
AAGCCTGTTTTTTTTTACTCAGGAACGTGCTACACGACCAGTGTACTATGGATTGTGGACATTT
15 CCTTCTGTGGAGACACGGTGGAGAAC**TAA**ACAATTTTTTAAAGCCACTATGGATTTAGTCAT
CTGAATATGCTGTGCAGAAAAAATATGGGCTCCAGTGGTTTTTACCATGTCATTCTGAAATT
TTTCTCTACTAGTTATGTTTGATTTCTTTAAGTTTCAATAAAATCATTTAGCATTGAAAAAAA

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FIGURE 22

5	Signal Peptide:	amino acids 1-20
	N-myristoylation Sites:	amino acids 67-72
		amino acids 118-123
		amino acids 163-168
	Flavodoxin protein homology:	amino acids 156-174

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MKFTIVFAGLLGVFLAPALANYNINVNDDNNNAGSGQQSVSVNNEHNVANVDNNGWDSWNS
IWDYGNGFAATRLFQKKTCIVHKMNKEVMPISIQSLDALVKEKKLQKGPGGPPPKGLMYSVN
15 PNKVDDLKFGKNIANMCRGIPTYMAEEMQEASLFFYSGTCYTTSVLWIVDISFCGDTVEN

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FIGURE 23

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CGGACGCGTGGGCGGGCCGGGACGCAGGGCAAAGCGAGCC**ATGG**CTGTCTACGTCTGGGATGC
TGCGCCTGGGGAGGCTGTGCGCCGGGAGCTCGGGGGTGCTGGGGGCCCCGGGCCGCCCTCTCT
CGGAGTTGGCAGGAAGCCAGGTTGCAGGGTGTCCGCTTCCTCAGTTCCAGAGAGGTGGATCG
10 CATGGTCTCCACGCCCATCGGAGGCCTCAGCTACGTTTCAGGGGTGCACCAAAAAGCATCTTA
ACAGCAAGACTGTGGGCCAGTGCCCTGGAGACCACAGCACAGAGGGTCCCAGAACGAGAGGCC
TTGGTCGTCTCTCCATGAAGACGTCAGGTTGACCTTTGCCCAACTCAAGGAGGAGGTGGACAA
AGCTGCTTCTGGCCTCCTGAGCATTGGCCTCTGCAAAGGTGACCGGCTGGGCATGTGGGGAC
CTAACTCCTATGCATGGGTGCTCATGCAGTTGGCCACCGCCAGGCGGGCATCATTCTGGTG
15 TCTGTGAACCCAGCCTACCAGGCTATGGAAGTGGAGTATGTCTCAAGAAGGTGGGCTGCAA
GGCCCTTGTTGTTCCCCAAGCAATTCAAGACCCAGCAATACTACAACGTCTGAAGCAGATCT
GTCCAGAAGTGGAGAATGCCCAGCCAGGGGCCCTGAAGAGTCAGAGGCTCCCAGATCTGACC
ACAGTCATCTCGGTGGATGCCCCCTTTGCCGGGGACCCCTGCTCCTGGATGAAGTGGTGGCGGC
TGGCAGCACACGGCAGCATCTGGACCAGCTCCAATACAACCAGCAGTTCCTGTCTGCCATG
20 ACCCCATCAACATCCAGTTCACCTCGGGGACAACAGGCAGCCCCAAGGGGGCCACCCTCTCC
CACTACAACATTGTCAACAACTCCAACATTTTAGGAGAGCGCCTGAAACTGCATGAGAAGAC
ACCAGAGCAGTTGCGGATGATCCTGCCCAACCCCCCTGTACCATTGCCTGGGTTCCTGGCAG
GCACAATGATGTGTCTGATGTACGGTGCCACCCTCATCCTGGCCTCTCCCATCTTCAATGGC
AAGAAGGCACTGGAGGCCATCAGCAGAGAGAGAGGCACCTTCCTGTATGGTACCCCCACGAT
25 GTTCGTGGACATTCTGAACCAGCCAGACTTCTCCAGTTATGACATCTCGACCATGTGTGGAG
GTGTCAATTGCTGGGTCCCCTGCACCTCCAGAGTTGATCCGAGCCATCATCAACAAGATAAAT
ATGAAGGACCTGGTGGTTGCTTATGGAACCACAGAGAACAGTCCCGTGACATTCGCGCACTT
CCCTGAGGACACTGTGGAGCAGAAGGCAGAAAGCGTGGGCAGAATTATGCCTCACACGGAGG
CCCGGATCATGAACATGGAGGCAGGGACGCTGGCAAAGCTGAACACGCCCCGGGGAGCTGTGC
30 ATCCGAGGGTACTGCGTCATGCTGGGCTACTGGGGTGAGCCTCAGAAGACAGAGGAAGCAGT
GGATCAGGACAAGTGGTATTGGACAGGAGATGTCGCCACAATGAATGAGCAGGGCTTCTGCA
AGATCGTGGGCCGCTCTAAGGATATGATCATCCGGGGTGGTGAGAACATCTACCCCGCAGAG
CTCGAGGACTTCTTTTACACACACCCGAAGGTGCAGGAAGTGCAGGTGGTGGGAGTGAAGGA
CGATCGGATGGGGGAAGAGATTTGTGCCTGCATTTCGGCTGAAGGACGGGGAGGAGACCACGG
35 TGGAGGAGATAAAAGCTTTCTGCAAAGGGAAGATCTCTCACTTCAAGATTCCGAAGTACATC
GTGTTTGTACAAACTACCCCTCACCATTTTCAGGAAAGATCCAGAAATTCAAACCTTCGAGA
GCAGATGGAACGACATCTAAATCTGTGAATAAAAGCAGCAGGCCTGTCTTGGCCGGTTGGCTT
GACTCTCTCCTGTGAGAAATGCAACCTGGCTTTATGCACCTAGATGTCCCCAGCACCCAGTTC
TGAGCCAGGCACATCAAATGTCAAGGAATTGACTGAACGAATAAGAGCTCCTGGATGGGTC
40 CGGGAACCTCGCCTGGGCACAAGGTGCCAAAAGGCAGGCAGCCTGCCAGGCCCTCCCTCCTG
TCCATCCCCACATTCCCCTGTCTGTCTTGTGATTTGGCATAAAGAGCTTCTGTTTTCTTT
GAAAAAAAAAAAAAAAAA

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FIGURE 24

5	Signal Peptide:	amino acids 1-22
	Transmembrane Domains:	amino acids 140-161
		amino acids 213-229
		amino acids 312-334
	Putative AMP-binding Domain Signature:	amino acids 260-271
10	N-myristoylation Sites:	amino acids 19-24
		amino acids 22-27
		amino acids 120-125
		amino acids 203-208
		amino acids 268-273
15		amino acids 272-277
		amino acids 314-319
		amino acids 318-323
		amino acids 379-384
		amino acids 380-385
20		amino acids 409-413
	N-glycosylation Site:	amino acids 282-285
25	MAVYVGMLRLGRLCAGSSGVLGARAALSRWQEARLQGVRFLSSREVD RMVSTPIGGLSYVQ	
	GCTKKHLNSKTVGQCLETTAQRVPEREALVVLHEDVRLTFAQLKEEVDKAASGLLSIGLCKG	
	DRLGMWGPNSYAWVLMQLATAQAGIILVSVNPAYQAMELEYVLKKVGCKALVFPKQFKTQQY	
	YNVLKQICPEVENAQP GALKSQRLPDLTTVISVDAPLP GTLLLDEVVAAGSTRQHLDQLQYN	
30	QQFLSCHDPINIQFTSGTTGSPKGATLSHYNIVNNSNILGERLKLHEKTPEQLRMILPNPLY	
	HCLGSVAGTMMCLMYGATLILASPIFNGKKALEAISRRERGTFLYGTPTMFVDILNQPDFSSY	
	DISTMCGGVIAGSPAPPELIRAIINKINMKDLVVAYGTTENSPVTFAHFPEDTVEQKAESVG	
	RIMPHTEARIMNMEAGTLAKLNTPGELCIRGYCVMLGYWGEPQKTEEAVDQDKWYWTGDVAT	
	MNEQGFCKIVGRSKDMIIRGGENIYPAELEDFHTHPKVQEVQVVGVKDDRMGEEICACIRL	
35	KDGEETTVEEIKAFCKGKISHFKIPKYIVFVTNYPLTISGKIQKFKLREQMERHLNL	
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FIGURE 25

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CAGAGAGGAGGCTTTGGGAATTGTCCAGCAGAAACAGAGAAGTCTGAGGTGGTGTCAAGACA
AAAG**ATG**CTTCAGCTTTGGAAACTTGTTCCTGTGCGGCGTGCTCACTGGGACCTCAGAGT
CTCTTCTTGACAATCTTGGCAATGACCTAAGCAATGTCGTGGATAAGCTGGAACCTGTTCTT
CACGAGGGACTTGAGACAGTTGACAATACTCTTAAAGGCATCCTTGAGAAACTGAAGGTCGA
10 CCTAGGAGTGCTTCAGAAATCCAGTGCTTGGCAACTGGCCAAGCAGAAGGCCACAGGAAGCTG
AGAAATTGCTGAACAATGTCATTTCTAAGCTGCTTCCAACATAACACGGACATTTTGGGGTTG
AAAATCAGCAACTCCCTCATCCTGGATGTCAAAGCTGAACCGATCGATGATGGCAAAGGCCT
TAACCTGAGCTTCCCTGTCACCGCGAATGTCACTGTGGCCGGGCCCATCATTTGGCCAGATTA
TCAACCTGAAAGCCTCCTTGGACCTCCTGACCGCAGTCACAATTGAAACTGATCCCCAGACA
15 CACCAGCCTGTTGCCGTCCTGGGAGAATGCGCCAGTGACCCAACCAGCATCTCACTTTCCTT
GCTGGACAAACACAGCCAAATCATCAACAAGTTCGTGAATAGCGTGATCAACACGCTGAAAA
GCACTGTATCCTCCCTGCTGCAGAAGGAGATATGTCCACTGATCCGCATCTTCATCCACTCC
CTGGATGTGAATGTCATTCAGCAGGTCGTCGATAATCCTCAGCACAAAACCCAGCTGCAAAC
CCTCATC**TGA**AGAGGACGAATGAGGAGGACCACTGTGGTGCATGCTGATTGGTTCCCAGTGG
20 CTTGCCCCACCCCCTTATAGCATCTCCCTCCAGGAAGCTGCTGCCACCACCTAACCAGCGTG
AAAGCCTGAGTCCCACCAGAAGGACCTTCCCAGATACCCCTTCTCCTCACAGTCAGAACAGC
AGCCTCTACACATGTTGTCCTGCCCCTGGCAATAAAGGCCCATTTCTGCACCCTTAA

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FIGURE 26

5 Signal Peptide: amino acids 1-14
N-glycosylation Sites: amino acids 124-127
amino acids 132-135
Leucine zipper Pattern: amino acids 44-65

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MLQLWKLVLCCGVLTGTSESLLDNLGNDLSNVVDKLEPVLHEGLETVDNTLKGILEKLKVDL
GVLQKSSAWQLAKQKAQEAELNNVISKLLPTNTDIFGLKISNSLILDVKAEPIDDGKGLN
LSFPVTANVTVAGPIIGQIINLKASLDLLTAVTIETDPQTHQPVAVLGECASDPTSISLSLL
DKHSQIINKFVNSVINTLKSTVSSLLQKEICPLIRIFIHSLDVNVIQQVVDNPQHKTQLQTLI

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FIGURE 27

5 CAAGTAAATGCAGCACTAGTGGGTGGGATTGAGGTATGCCCTGGTGCATAAATAGAGACTCA
GCTGTGCTGGCACACTCAGAAGCTTGGACCGCATCCTAGCCGCCGACTCACACAAGGCAGGT
GGGTGAGGAAATCCAGAGTTGCC**ATG**GAGAAAATTCCAGTGTGAGCATTCCTTGCTCCTTG
GCCCTCTCCTACACTCTGGCCAGAGATACCACAGTCAAACCTGGAGCCAAAAAGGACACAAA
10 GGACTCTCGACCCAACTGCCCCAGACCCCTCTCCAGAGGTTGGGGTGACCAACTCATCTGGA
CTCAGACATATGAAGAAGCTCTATATAAATCCAAGACAAGCAACAAACCCTTGATGATTATT
CATCACTTGATGAGTGGCCACACAGTCAAGCTTTAAAGAAAGTGTTTGCTGAAAATAAAGA
AATCCAGAAATTGGCAGAGCAGTTTGTCTCCTCAATCTGGTTTATGAAACAACTGACAAAC
ACCTTTCTCCTGATGGCCAGTATGTCCCCAGGATTATGTTTGTTGACCCATCTCTGACAGTT
15 AGAGCCGATATCACTGGAAGATATTCAAATCGTCTCTATGCTTACGAACCTGCAGATACAGC
TCTGTGCTTGACAACATGAAGAAAGCTCTCAAGTTGCTGAAGACTGAATTG**TAA**AGAAAAA
AAATCTCCAAGCCCTTCTGTCTGTCAGGCCTTGAGACTTGAAACCAGAAGAAGTGTGAGAAG
ACTGGCTAGTGTGGAAGCATAGTGAACACACTGATTAGGTTATGGTTTAAATGTTACAACAAC
TATTTTTTAAAGAAAAACAAGTTTTAGAAATTTGGTTTCAAGTGTACATGTGTGAAAACAATA
20 TTGTATACTACCATAGTGAGCCATGATTTTCTAAAAAAAATAAATGTTA

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FIGURE 28

5 Signal Peptide: amino acids 1-20

MEKIPVSAFLLLVALSYTLARDTTVKPGAKKDTKDSRPKLPQTL SRGWGDQLIWTQTYEEAL
YKSKTSNKPLMIIHHLDEC PHSQALKKVFAENKEIQKLAEQFVLLNLVYETTDKHLSPDGQY
10 VPRIMFVDPSLTVRADITGRYSNRLYAYEPADTALLLDNMKKALKLLKTEL

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FIGURE 29

5 CGATGCGCGGACCCGGGCACCCCCTCCTCCTGGGGCTGCTGCTGGTGCTGGGGCCTTCGCCG
GAGCAGCGAGTGGAATTGTTTCCTCGAGATCTGAGGATGAAGGACAAGTTTCTAAAACACCT
TACAGGCCCTCTTTATTTTAGTCCAAAGTGCAGCAAACACTTCCATAGACTTTATCACAACA
CCAGAGACTGCACCATTCCTGCATACTATAAAAGATGCGCCAGGCTTCTTACCCGGCTGGCT
10 GTCAGTCCAGTGTGCATGGAGGATAAG**TGAG**CAGACCGTACAGGAGCAGCACACCAGGAGCC
ATGAGAAGTGCCTTGGAAACCAACAGGGAAACAGAACTATCTTTATACACATCCCCTCATGG
ACAAGAGATTTATTTTTTGCAGACAGACTCTTCCATAAGTCCTTTGAGTTTGTATGTTGTTG
ACAGTTTGCAGATATATATTTCGATAAATCAGTGTACTTGACAGTGTTATCTGTCACTTATTT

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FIGURE 30

5 Signal Peptide: amino acids 1-20
Glycoprotease family proteins: amino acids 56-80
Acyltransferases ChoActase/COT/CPT Family: amino acids 49-70

10 MRGPGHPLLLGLLLVLGPSPEQRVEIVPRDLRMKDKFLKHLTGPLYFSPKCSKHFHRLYHNT
RDCTIPAYYKRCARLLTRLAVSPVCMEDK

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FIGURE 31

TTGAGCGCAGGTGAGCTCCTGCGCGTTCCGGGGGCGTTCCTCCAGTCACCCTCCCGCCGTTA
5 CCCGCGGCGCGCCCCGAGGGAGTCTCCTCCAGACCCTCCCTCCCGTTGCTCCAACTAATACG
GACTGAACGGATCGCTGCGAGGGTGGGAGAGAAAATTAGGGGGAGAAAGGACAGAGAGAGCA
ACTACCATCCATAGCCAGATAGATTATCTTACACTGAACTGATCAAGTACTTTGAAAATGAC
TTCGAAATTTATCTTGGTGTCTTCATACTTGCTGCACTGAGTCTTTCAACCACCTTTTCTC
TCCAAC TAGACCAGCAAAAGGTTCTACTAGTTTCTTTTGATGGATTCCGTTGGGATTACTTA
10 TATAAAGTTCCAACGCCCCATTTTCATTATATTATGAAATATGGTGTTCACGTGAAGCAAGT
TACTAATGTTTTTATTACAAAAACCTACCCTAACCATTTACTTTGGTAACTGGCCTCTTTG
CAGAGAATCATGGGATTGTTGCAAATGATATGTTTGATCCTATTTCGGAACAAATCTTTCTCC
TTGGATCAGATGAATATTTATGATTCCAAGTTTGGGAAGAAGCGACACCAATATGGATCAC
AAACCAGAGGGCAGGACATACTAGTGGTGCAGCCATGTGGCCCGGAACAGATGTAAAAATAC
15 ATAAGCGCTTTCTACTCATTTACATGCCTTACAATGAGTCAGTTTCATTTGAAGATAGAGTT
GCCAAAATTGTTGAATGGTTTACGTCAAAAGAGCCATAAATCTTGGTCTTCTCTATTGGGA
AGACCCTGATGACATGGGCCACCATTGTTGGGACCTGACAGTCCGCTCATGGGGCCTGTCAATTT
CAGATATTGACAAGAAGTTAGGATATCTCATACAAATGCTGAAAAAGGCAAAGTTGTGGAAC
ACTCTGAACCTAATCATCACAAGTGATCATGGAATGACGCAGTGCTCTGAGGAAAGGTTAAT
20 AGAAGTTGACCAGTACCTGGATAAAGACCCTATACCCTGATTGATCAATCTCCAGTAGCAG
CCATCTTGCCAAAAGAAGGTAAATTTGATGAAGTCTATGAAGCACTAACTCACGCTCATCCT
AATCTTACTGTTTACAAAAAGAAGACGTTCCAGAAAGGTGGCATTACAAATACAACAGTCG
AATTCAACCAATCATAGCAGTGGCTGATGAAGGGTGGCACATTTTACAGAATAAGTCAGATG
ACTTTCTGTTAGGCAACCACGGTTACGATAATGCGTTAGCAGATATGCATCCAATATTTTTA
25 GCCCATGGTCCTGCCTTCAGAAAGAATTTCTCAAAAGAAGCCATGAACTCCACAGATTTGTA
CCCCTACTATGCCACCTCCTCAATATCACTGCCATGCCACACAATGGATCATTTCTGGAATG
TCCAGGATCTGCTCAATTCAGCAATGCCAAGGGTGGTCCCTTATACACAGAGTACTATACTC
CTCCCTGGTAGTGTTAAACCAGCAGAATATGACCAAGAGGGGTCATACCCTTATTTTCATAGG
GGTCTCTCTTGGCAGCATTATAGTGATTGTATTTTTTTGTAATTTTCATTAAGCATTTAATTC
30 ACAGTCAAATACCTGCCTTACAAGATATGCATGCTGAAATAGCTCAACCATTATTACAAGCC
TAATGTTACTTTGAAGTGGATTTGCATATTGAAGTGGAGATTCCATAATTATGTCAGTGTTT
AAAGGTTTCAAATTTCTGGGAAACCAGTTCCAAACATCTGCAGAAACCATTAAGCAGTTACAT
ATTTAGGTATACACACACACACACACACACATACACACACACGGACCAAATACTTACAC
CTGCAAAGGAATAAAGATGTGAGAGTATGTCTCCATTGTTCACTGTAGCATAGGGATAGATA
35 AGATCCTGCTTTATTTGGACTTGGCGCAGATAATGTATATATTTAGCAACTTTGCACTATGT
AAAGTACCTTATATATTGCACTTTAAATTTCTCTCCTGATGGGTACTTTAAATTTGAAATGCA
CTTTATGGACAGTTATGTCTTATACTTGATTGAAAATGACAACCTTTTTGCAACCCATGTCAC
AGAATACTTGTTACGCATTGTTCAAACCTGAAGGAAATTTCTAATAATCCCGAATAATGAACA
TAGAAATCTATCTCCATAAATTGAGAGAAGAAGGTGATAAGTGTGAAAATTAAATGTG
40 ATAACCTTTGAACCTTGAATTTTGGAGATGTATTCCCAACAGCAGAATGCAACTGTGGGCAT
TTCTTGCTTATTTCTTTCCAGAGAACGTGGTTTTTCATTTATTTTTCCCTCAAAAGAGAGTC
AAATACTGACAGATTCGTTCTAAATATATTGTTTCTGTCATAAAATTAATTGTGATTTCCTGA
TGAGTCATATTACTGTGATTTTCATAATAATGAAGACACCATGAATATACTTTTCTTCTATA
TAGTTCAGCAATGGCCTGAATAGAAGCAACCAGGCACCATCTCAGCAATGTTTTCTCTTGTT
45 TGTAATTATTTGCTCCTTTGAAAATTAAATCACTATTAATTACATTAAAAATCAAATTGGAT
AAAAAAAAAAAAAAAAAAAA

FIGURE 32

5	Signal Peptide:	amino acids 1-22
	Transmembrane Doman:	amino acids 429-452
	N-glycosylation Domains:	amino acids 101-104
		amino acids 158-161
		amino acids 292-295
10		amino acids 329-332
		amino acids 362-365
		amino acids 369-372
		amino acids 382-385
		amino acids 389-392
15	Somatomedin B Domain:	amino acids 69-85
	Sulfatase protein Region:	amino acids 212-241

MTSKFILVSFILAAALSLSTTFSLQLDQQKVLLVSFDGFRWDYLYKVPTPHFHYIMKYGVHVK
 20 QVTNVFITKTYPNHYTLVTGLFAENHGIVANDMFDPINRKSFSLDHMNIYDSKFWEEATPIW
 ITNQ RAGHTSGAAMWPGTDVKIHKRFPTHYMPYNESVSFEDRVAKIVEWFTSKEPINLGLLY
 WEDPDDMGHHLGPDSPLMGPVISDIDKKLGYLIQMLKKAKLWNTLNLIITSDHGMTQCSEER
 LIELDQYLDKDHYTLIDQSPVAAILPKEGKFDEVYEALTHAHPNLTVYKKEDVPERWHYKYN
 SRIQPIIAVADEGWHILQNKSDDFLLGNHGYDNALADMHPIFLAHGPAFRKNFSKEAMNSTD
 25 LYPLLCHLLNITAMPHNGSFWNVQDLLNSAMPRVVPYTQSTILLPGSVKPAEYDQEGSYPYF
 IGVSLGSIIVIVFFVIFIKHLIHSQIPALQDMHAEIAQPLLQA

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FIGURE 33

5 GTTCTCCTTTTCCGAGCCAAAATCCCAGGCGATGGTGAATTATGAACGTGCCACACC**ATGAAG**
CTCTTGTGGCAGGTAACCTGTGCACCACCACACCTGGAATGCCATCCTGCTCCCGTTCGTCTA
CCTCACGGCGCAAGTGTGGATTCTGTGTGCAGCCATCGCTGCTGCCGCCTCAGCCGGGCCCC
AGAACTGCCCCCTCCGTTTGGCTCGTGCAGTAACCAGTTCAGCAAGGTGGTGTGCACGCGCCGG
10 GGCCCTCTCCGAGGTCCCGCAGGGTATTCCCTCGAACACCCGGTACCTCAACCTCATGGAGAA
CAACATCCAGATGATCCAGGCCGACACCTTCCGCCACCTCCACCACCTGGAGGTCTCTGCAGT
TGGGCAGGAACCTCCATCCGGCAGATTGAGGTGGGGGCCTTCAACGGCCTGGCCAGCCTCAAC
ACCCTGGAGCTGTTTCGACAACCTGGCTGACAGTCATCCCTAGCGGGGCCTTTGAATACCTGTC
CAAGCTGCGGGAGCTCTGGCTTCGCAACAACCCCATCGAAAGCATCCCCTCTTACGCCTTCA
15 ACCGGGTGCCCTCCCTCATGCGCCTGGACTTGGGGGAGCTCAAGAAGCTGGAGTATATCTCT
GAGGGAGCTTTTGAAGGGCTGTTCAACCTCAAGTATCTGAACTTGGGCATGTGCAACATTAA
AGACATGCCCAATCTCACCCCCCTGGTGGGGCTGGAGGAGCTGGAGATGTCAGGGAACCACT
TCCCTGAGATCAGGCCTGGCTCCTTCCATGGCCTGAGCTCCCTCAAGAAGCTCTGGGTCATG
AACTCACAGGTCAGCCTGATTGAGCGGAATGCTTTTGACGGGCTGGCTTCACTTGTGGAACCT
20 CAACCTTGGCCCCACAATAACCTCTCTTCTTTGCCCCATGACCTCTTTACCCCGCTGAGGTACC
TGGTGGAGTTGCATCTACACCACAACCCCTTGGAACTGTGATTGTGACATTCTGTGGCTAGCC
TGGTGGCTTCGAGAGTATATACCCACCAATTCCACCTGCTGTGGCCGCTGTGATGCTCCCAT
GCACATGCGAGGCCGCTACCTCGTGGAGGTGGACCAGGCCTCCTTCCAGTGCTCTGCCCCCT
TCATCATGGACGCACCTCGAGACCTCAACATTTCTGAGGGTCGGATGGCAGAACTTAAGTGT
25 CGGACTCCCCCTATGTCCTCCGTGAAGTGGTTGCTGCCCAATGGGACAGTGCTCAGCCACGC
CTCCCCGCCACCCAAGGATCTCTGTCTCAACGACGGCACCTTGAACTTTTCCACAGTGCTGC
TTTCAGACACTGGGGTGTACACATGCATGGTGACCAATGTTGCAGGCAACTCCAACGCCTCG
GCCTACCTCAATGTGAGCACGGCTGAGCTTAACACCTCCAACCTACAGCTTCTTACCCACAGT
AACAGTGGAGACCACGGAGATCTCGCCTGAGGACACAACGCGAAAGTACAAGCCTGTTCCCTA
30 CCACGTCCACTGGTTACCAGCCGGCATATACCACCTCTACCACGGTGCTCATTCAGACTACC
CGTGTGCCCAAGCAGGTGGCAGTACCCGCGACAGACACCACTGACAAGATGCAGACCAGCCT
GGATGAAGTCATGAAGACCACCAAGATCATCATTGGCTGCTTTGTGGCAGTGACTCTGCTAG
CTGCCGCCATGTTGATTGTCTTCTATAAACTTCGTAAGCGGCACCAGCAGCGGAGTACAGTC
ACAGCCGCCCCGACTGTTGAGATAATCCAGGTGGACGAAGACATCCCAGCAGCAACATCCGC
35 AGCAGCAACAGCAGCTCCGTCCGGTGTATCAGGTGAGGGGGCAGTAGTGCTGCCCACAATTC
ATGACCATATTAACCTACAACACCTACAAACCAGCACATGGGGCCCACTGGACAGAAAACAGC
CTGGGGAACCTCTCTGCACCCACAGTCAACCTATCTCTGAACCTTATATAATTCAGACCCA
TACCAAGGACAAGGTACAGGAACTCAAATAT**TGACT**CCCCCTCCCCCAAAAACTTATAAAAT
GCAATAGAATGCACACAAAGACAGCAACTTTTGTACAGAGTGGGGAGAGACTTTTTCTTGTA
40 TATGCTTATATATTAAGTCTATGGGCTGGTTAAAAAAAACAGATTATATTAATAATTAAAGA
CAAAAAGTCAAAACA

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FIGURE 34

5 Transmembrane Domains: amino acids 21-40
amino acids 528-548
N-glycosylation Sites: amino acids 277-280
amino acids 322-325
amino acids 363-366
10 amino acids 388-391
amino acids 410-413
amino acids 434-437
amino acids 440-443
amino acids 447-450
15 amino acids 450-453
Glycosaminoglycan attachment Site: amino acids 591-594
Leucine zipper Pattern: amino acids 271-292

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MKLLWQVTVHHHTWNAILLPFVYLTAQVWILCAAIAAAASAGPQNCPSVCSCSNQFSKVVCT
RRGLSEVPQGIPSNTRYLNLMENNIQMIQADTFRHLHHLEVLQLGRNSIRQIEVGAFNGLAS
LNTLELFDNWLTVIPSGAFEYLSKLRELWLRNNPIESIPSYAFNRVP SLMRLDLGELKKLEY
ISEGAFEGFLFNLYLNLGMCNIKDMPNLTPLVGLEELEM SGNHFPEIRPGSFHGLSSLKKLW
25 VMNSQVSLIERNAFDGLASLVELNLAHNNLSSLPHDLFTPLRYLVELHLHHPWNCDCDILW
LAWWLREYIPTNSTCCGRCHAPMHMRGRYLVEVDQASFC SAFPIMDAPRDLNISEGRMAEL
KCRTPPMSSVKWLLPNGTVLSHASRHPRI SVLNDGTLNFSHVLLSDTGVYTCMVTNVAGNSN
ASAYLNVSTAE LNTSNYSFFTTVTVETTEISPEDTTRKYKPVPTTSTGYQPAYTTSTTVLIQ
TTRVPKQVAVPATD TTDKMQTSLEVMKTTKIIIGCFVAVTLLAAAMLIVFYKLKRHQQRS
30 TVTAARTVEIIQVDEDIPAATSAAATAAPSGVSGEGAVVLPTIHDHINYNTYKPAHGAHWTE
NSLGNSLHPTVTTISEPYIIQTHTKDKVQETQI

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FIGURE 35

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GGGCTGGGCCCCGCCGCAGCTCCAGCTGGCCGGCTTGGTCCTGCGGGTCCCTTCTCTGGGAGG
CCCGACCCCGGCCGCGCCCAGCCCCACCA**ATG**CCACCCGCGGGGCTCCGCCGGGCGCGCCG
CTCACCGCAATCGCTCTGTTGGTGCTGGGGGTCCCCCTGGTGCTGGCCGGCGAGGACTGCCT
GTGGTACCTGGACCGGAATGGCTCCTGGCATCCGGGGTTTAACTGCGAGTTCTTCACCTTCT
10 GCTGCGGGACCTGCTACCATCGGTACTGCTGCAGGGACCTGACCTTGCTTATCACCGAGAGG
CAGCAGAAGCACTGCCTGGCCTTCAGCCCCAAGACCATAGCAGGCATCGCCTCAGCTGTGAT
CCTCTTTGTTGCTGTGGTTGCCACCACCATCTGCTGCTTCCTCTGTTCTCTGTTGCTACCTGT
ACCGCCGGCGCCAGCAGCTCCAGAGCCCATTTGAAGGCCAGGAGATTCCAATGACAGGCATC
CCAGTGCAGCCAGTATACCCATACCCCCAGGACCCCAAAGCTGGCCCTGCACCCCCACAGCC
15 TGGCTTCATGTACCCACCTAGTGGTCCTGCTCCCCAATATCCACTCTACCCAGCTGGGCCCC
CAGTCTACAACCCTGCAGCTCCTCCTCCCTATATGCCACCACAGCCCTCTTACCCGGGAGCC
TGAGGAACCAGCCATGTCTCTGCTGCCCTTCAGTGATGCCAACCTTGGGAGATGCCCTCAT
CCTGTACCTGCATCTGGTCCTGGGGGTGGCAGGAGTCCTCCAGCCACCAGGCCCCAGACCAA
GCCAAGCCCTGGGCCCTACTGGGGACAGAGCCCCAGGGAAGTGGAACAGGAGCTGAACTAGA
20 ACTATGAGGGGTTGGGGGGAGGGCTTGGAATTATGGGCTATTTTTACTGGGGGCAAGGGAGG
GAGATGACAGCCTGGGTCACAGTGCCCTGTTTTCAAATAGTCCCTCTGCTCCCAAGATCCCAG
CCAGGAAGGCTGGGGCCCTACTGTTTGTCCCCTCTGGGCTGGGGTGGGGGGAGGGAGGAGGT
TCCGTCAGCAGCTGGCAGTAGCCCTCCTCTCTGGCTGCCCCACTGGCCACATCTCTGGCCTG
CTAGATTAAAGCTGTAAAGACAAAA

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FIGURE 36

5 Transmembrane Domains:	amino acids 10-28
	amino acids 85-110
N-glycosylation Site:	amino acids 38-41
N-myristoylation Sites:	amino acids 5-10
	amino acids 88-93

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MPPAGLRRRAAPLTAIALLVLGAPLVLAGEDCLWYLDRNGSWHPGFNCEFFTFCCGTCYHRYC
CRDLTLLITERQQKHCLAFSPKTIAGIASAVILFVAVVATTICCFLCSCCYLYRRRQQQLQSP
FEGQEIPMTGIPVQPVYPYPQDPKAGPAPPQPGFMYPSPGPAPQYPLYPAGPPVYNPAAPP

15 YMPPQPSYFGA

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FIGURE 37

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GCGACGGGCAGGACGCCCCGTTGCGCTAGCGCGTGCTCAGGAGTTGGTGTCTGCCTGCGCT
CAGG**ATG**AGGGGGGAATCTGGCCCTGGTGGGCGTTCTAATCAGCCTGGCCTTCCTGTCACTGC
TGCCATCTGGACATCCTCAGCCGGCTGGCGATGACGCCTGCTCTGTGCAGATCCTCGTCCCT
10 GGCCTCAAAGGGGATGCGGGAGAGAAGGGAGACAAAGGCGCCCCCGGACGGCCTGGAAGAGT
CGGCCCCACGGGAGAAAAAGGAGACATGGGGGACAAAGGACAGAAAGGCAGTGTGGGTCTGTC
ATGGAAAAATTGGTCCCATTGGCTCTAAAGGTGAGAAAGGAGATTCCGGTGACATAGGACCC
CCTGGTCCCTAATGGAGAACCAGGCCTCCCATGTGAGTGCAGCCAGCTGCGCAAGGCCATCGG
GGAGATGGACAACCAGGTCTCTCAGCTGACCAGCGAGCTCAAGTTTCATCAAGAATGCTGTCTG
15 CCGGTGTGCGCGAGACGGAGAGCAAGATCTACCTGCTGGTGAAGGAGGAGAAGCGCTACGCG
GACGCCCAGCTGTCTTGCCAGGGCCGCGGGGGCACGCTGAGCATGCCCAAGGACGAGGCTGC
CAATGGCCTGATGGCCGCATACCTGGCGCAAGCCGGCCTGGCCCGTGTCTTCATCGGCATCA
ACGACCTGGAGAAGGAGGGCGCCTTCGTGTACTCTGACCACTCCCCCATGCGGACCTTCAAC
AAGTGGCGCAGCGGTGAGCCCAACAATGCCTACGACGAGGAGGACTGCGTGGAGATGGTGGC
20 CTCGGGCGGCTGGAACGACGTGGCCTGCCACACCACCATGTACTTCATGTGTGAGTTTGACA
AGGAGAACATG**TGA**GCCTCAGGCTGGGGCTGCCCATTGGGGGCCCCACATGTCCCTGCAGGG
TTGGCAGGGACAGAGCCCAGACCATGGTGCCAGCCAGGGAGCTGTCCCTCTGTGAAGGGTGG
AGGCTCACTGAGTAGAGGGCTGTTGTCTAAACTGAGAAAATGGCCTATGCTTAAGAGGAAAA
TGAAAGTGTTCTGGGGTGCTGTCTCTGAAGAAGCAGAGTTTCATTACCTGTATTGTAGCCC
25 CAATGTCATTATGTAATTATTACCCAGAATTGCTCTTCCATAAAGCTTGTGCCTTTGTCCAA
GCTATACAATAAAATCTTTAAGTAGTGCAGTAGTTAAGTCCAAAAAAAAAAAAAAAAAAAAA

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FIGURE 38

5 Signal Peptide: amino acids 1-25
C-type lectin Domain: amino acids 247-256
Clq Domain: amino acids 44-77

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MRGNLALVGVLIISLAFLLPSGHPQPAGDDACSVQILVPGLKGDAGEKGDKGAPGRPGRVG
PTGEKGMGDKGQKGSVGRHGKIGPIGSKGEKGDSDIGPPGPNGEPGLPCECSQLRKAIGE
MDNQVSQLTSELKFIKNAVAGVRETESKIYLLVKEEKRYADAQLSCQGRGGTLSMPKDEAAN
15 GLMAAYLAQAGLARVFIGINDLEKEGAFVYSDHSPMRTFNKWRSGEPNNAYDEEDCVEMVAS
GGWNDVACHTTMYFMCEFDKENM

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FIGURE 39

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CGGCCACAGCTGGCATGCTCTGCCTGATCGCCATCCTGCTGTATGTCCTCGTCCAGTACCTC
GTGAACCCCGGGGTGCTCCGCACGGACCCAGATGTCAAGAATATGAACACGTGGCTGCTGT
TCCTCCCCCTGTTCCCGGTGCAGGTGCAGACCCTGATAGTCGTGATCATCGGGATGCTCGTG
10 CTCCTGCTGGACTTTCTTGGCTTGGTGCACCTGGGCCAGCTGCTCATCTTCCACATCTACCT
GAGTATGTCCCCCACCCTAAGCCCCCGATCCCCCAAGGCTGGGTGGTTCAGAGCTGCTCATC
TTACACCTCTACTTGAGTATGTCCCTAACCCTGAGCCCCCACGCCTGGGGCCAGAGTCTTT
GTCCCCCGTGTGCGCATGTGTTTCAGGGTCAGCCTCTCCCAGAAGTGAGATCATGGACAAAAA
GGGCAAATCACAGGAAGAAATTAAATCCATGAGGACCCAGCAGGCCCCAGCAAGAAGCTGAAC
15 TCACGCCGAGACCTGCAGGAGTGGTGCCAGGTGCTTGAAGTAACAAGTTTAAAATGTTTCAGA
GACAATGGAATGGAATCTATTAGGCAAGAACAGGACATTATGAAATAAGGACAGGTGGACTT
CCAAAAACACAAGTAGAAATTCTAACAATGAAATATATTACAGGCAGGTCACCCACTAACCA
AACAACTGAAGCGAGAGCTGGTGGTCTTGCTTGGTCTCACAGTGGGCACAGCGGTAGGCGGT
CAGTCATGTTGCTGAACGACGGAGGGTAAACTCCCCAGCCCCAAGAAAACCTGTGTTGGAAG
20 TAACAACAACCTCCCTGCTCCTGGCACCAGCCGTTTTTGGTTCATGGTGGGCCAGCTGCAAAGC
GTCTTCCATTCTCTGGGCAGTGGTGGCCCCGAGGCTGTGGCCTCTCAGGGGGTTTTCTGTGGA
CACGGGCAGCAGAGTGTTGTCCAGGCCAGCCCCAAGAATGCCCTGCTCCTGACAGCTTGGCC
AACCCTGGTTCAGGGCAGAGGGAGTTGGGTGGGTTCAGGCTCTGGGCTCACCTCCATCTCCAG
AGCATCCCCTGCCTGCAGTTGTGGCAAGAACGCCCAGCTCAGAATGAACACACCCCACCAAG
25 AGCCTCCTTGTTTCATAACCACAGGTTACCCTACAAACCACTGTCCCCACACAACCCTGGGGA
TGTTTTTAAACACACACCTCTAACGCATATCTTACAGTCACTGTTGTCTTGCCTGAGGGTTG
AATTTTTTTTAAATGAAAGTGCAATGAAAATCACTGGATTAAATCCTACGGACACAGAGCTGA
AAAAAAAAAAAAAAAAAAAAAAAAAAAAA

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FIGURE 40

5 Signal Peptide: amino acids 1-38

MNTWLLFLPLFPVQVQTLIVVIIGMLVLLLDLGLVHLGQLLIFHIYLSMSPTLSPRSPQGW
10 VVRAAHLTPLLEYVPNPEPPTPGARVFVPRVRMCSGSASPRSEIMDKKGKSQEEIKSMRTQQ
AQQEAELTPRPAGVVPGA

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FIGURE 41

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GGACTCTGAAGGTCCCAAGCAGCTGCTGAGGCCCCCAAGGAAGTGGTTCCAACCTTGGACCC
CTAGGGGTCTGGATTTGCTGGTTAACAAGATAACCTGAGGGCAGGACCCCATAGGGGAATGC
TACCTCCTGCCCTTCCACCTGCCCTGGTGTTCACGGTGGCCTGGTCCCTCCTTGCCGAGAGA
10 GTGTCCTGGGTCAGGGACGCAGAGGACGCTCACAGACTCCAGCCCTTTGTTACCGAGAGGAC
ACTTGGCAAGGTCCAGCGATGGTCCGGAGTCCACACACAGACTGGCGGCAGGGCAGGAGGGG
GACAGTTCTGTTGTGCTTGGTTGGACAGTAAGAGGGTCTTGGCCAGTCCAGGGTGGGGGGCG
GCAAACCTCCATAAAGAACCAGAGGGTCTGGGCCCCGGCCACAGAGTCATCTGCCCAGCTCCT
CTGCTGCTGGCCAGTGGGAGTGGCACGAGGTGGGGCTTTGTGCCAGTAAAAACACAGGCTGG
15 ATTTGCCTGCGGGCCATGGTCCCTGTCTAGGGCAGCAATTCTCAACCTTCTTGCTCTCAGGA
CCCCAAAGAGCTTTCATTGTATCTATTGATTTTTTACCACATTAGCAATTAAACTGAGAAAT
GGGCCGGGCACGGTGGCTCACGCCTGTAATCCCAGCACTTTGGGAGGCCGAGGCGGGTGGAT
CACCTGAGATCAGGAGTTCAAGACCAGCCTGGCCAACATGGTGAAACCTTGTCTACTAAAAA
TACAAAAAATTAGCCAGGCACAGTGGTGTGCACTGGTAGTCCCAGTTACTCGGGAGGCTGAG
20 GCAGGAAAATCGCTTGAACCCAGGAGGCGGACGTTGCGGTGAGCCGAGATCGCGCCGCTGAT
TCCAGCCTGGGCGACAAGAGTGAGACTCCATCTCACACA

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FIGURE 42

5 Signal Peptide: amino acids 1-17
N-myristoylation Sites: amino acids 58-63
amino acids 63-68
amino acids 64-69
amino acids 83-88
10 amino acids 111-116
amino acids 115-120

15 MLPPALPPALVFTVAWSLLAERVSWVRDAEDAHRLQPFVTERTLGKVQRWSGVHTQTGGRAG
GGQFCCAWLDSKRVLASPGWGAANSIKNRVWAPATESSAQLLCCWPVGVARGGALCQ

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FIGURE 43A

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 GCCGAGCGCAAGAACCCTGCGCAGCCCAGAGCAGCTGCTGGAGGGGAATCGAGGCGCGGCTC
 CGGGGATTCGGCTCGGGCCGCTGGCTCTGCTCTGCGGGGAGGGAGCGGGCCCGCCCGCGGGG
 CCCGAGCCCTCCGGATCCGCCCCCTCCCCGGTCCCGCCCCCTCGGAGACTCCTCTGGCTGCT
 CTGGGGGTTCGCCGGGGCCGGGGACCCGCGGTCCGGGGCGCC**ATG**CGGGCATCGCTGCTGCTG
 10 TCGGTGCTGCGGCCCCGAGGGCCCCGTGGCCGTGGGCATCTCCCTGGGCTTCACCCTGAGCCT
 GCTCAGCGTCACCTGGGTGGAGGAGCCGTGCGGCCAGGCCCGCCCCAACCTGGAGACTCTG
 AGCTGCCGCCGCGCGGCAACACCAACGCGGCGCGCCGGCCCAACTCGGTGCAGCCCCGGAGCG
 GAGCGCGAGAAGCCCCGGGGCCGGCGAAGGCGCCGGGGAGAATTGGGAGCCGCGCGTCTTGCC
 CTACCACCCTGCACAGCCCCGGCCAGGCCGCCAAAAAGGCCGTCAGGACCCGCTACATCAGCA
 15 CGGAGCTGGGCATCAGGCAGAGGCTGCTGGTGGCGGTGCTGACCTCTCAGACCACGCTGCC
 ACGCTGGGCGTGGCCGTGAACCGCACGCTGGGGCACCGGCTGGAGCGTGTGGTGTTCCTGAC
 GGGCGCACGGGGCCCGCCGGGCCCCACCTGGCATGGCAGTGGTGACGCTGGGCGAGGAGCGAC
 CCATTGGACACCTGCACCTGGCGCTGCGCCACCTGCTGGAGCAGCACGGCGACGACTTTGAC
 TGGTTCTTCCTGGTGCCTGACACCACCTACACCGAGGCGCACGGCCTGGCACGCCTAACTGG
 20 CCACCTCAGCCTGGCCTCCGCCGCCACCTGTACCTGGGCCGGCCCCAGGACTTCATCGGCG
 GAGAGCCCACCCCCGGCCGCTACTGCCACGGAGGCTTTGGGGTGCTGCTGTGCGCGCATGCTG
 CTGCAACAACCTGCGCCCCCACCTGGAAGGCTGCCGCAACGACATCGTCAGTGCGCGCCCTGA
 CGAGTGGCTGGGTGCTGCTGCATTCTCGATGCCACCGGGGTGGGCTGCACTGGTGACCACGAGG
 GGGTGCACTATAGCCATCTGGAGCTGAGCCCTGGGGAGCCAGTGCAGGAGGGGGACCCTCAT
 25 TTCCGAAGTGCCCTGACAGCCCACCCTGTGCGTGACCCTGTGCACATGTACCAGCTGCACAA
 AGCTTTTCGCCCGAGCTGAACTGGAACGCACGTACCAGGAGATCCAGGAGTTACAGTGGGAGA
 TCCAGAATACCAGCCATCTGGCCGTTGATGGGGACCGGGCAGCTGCTTGGCCCCGTGGGTATT
 CCAGCACCATCCCCGCCCGCCTCCCGCTTTGAGGTGCTGCGCTGGGACTACTTCACGGAGCA
 GCACGCTTTCTCCTGCGCCGATGGCTCACCCCGCTGCCCACTGCGTGGGGCTGACCGGGCTG
 30 ATGTGGCCGATGTTCTGGGGACAGCTCTAGAGGAGCTGAACCGCCGCTACCACCCGGCCTTG
 CGGCTCCAGAAGCAGCAGCTGGTGAATGGCTACCGACGCTTTGATCCGGCCCCGGGGTATGGA
 ATACACGCTGGACTTGCAGCTGGAGGCACTGACCCCCCAGGGAGGCCGCCGGCCCCCTCACTC
 GCCGAGTGCAGCTGCTCCGGCCGCTGAGCCGCGTGAGATCTTGCCCTGTGCCCTATGTCACT
 GAGGCCTCACGTCTCACTGTGCTGCTGCCTCTAGCTGCGGCTGAGCGTGACCTGGCCCCCTGG
 35 CTTCTTGAGGGCCTTTGCCACTGCAGCACTGGAGCCTGGTGATGCTGCGGCAGCCCTGACCC
 TGCTGCTACTGTATGAGCCGCGCCAGGCCCAGCGCGTGGCCCATGCAGATGTCTTCGCACCT
 GTCAAGGCCCCACGTGGCAGAGCTGGAGCGGCGTTTCCCCGGTGCCCGGGTGCCATGGCTCAG
 TGTGCAGACAGCCGCACCCTCACCACTGCGCCTCATGGATCTACTCTCCAAGAAGCACCCGC
 TGGACACACTGTTCTTGCTGGCCGGGCCAGACACGGTGCTCACGCCTGACTTCCTGAACCGC
 40 TGCCGCATGCATGCCATCTCCGGCTGGCAGGCCTTCTTTCCCATGCATTTCCAAGCCTTCCA
 CCCAGGTGTGGCCCCACCACAAGGGCCTGGGCCCCCAGAGCTGGGCCGTGACACTGGCCGCT
 TTGATCGCCAGGCAGCCAGCGAGGCCTGCTTCTACAACCTCCGACTACGTGGCAGCCCCGTGGG
 CGCCTGGCGGCAGCCTCAGAACAAGAAGAGAGCTGCTGGAGAGCCTGGATGTGTACGAGCT
 GTTCCTCCACTTCTCCAGTCTGCATGTGCTGCGGGCGGTGGAGCCGGCGCTGCTGCAGCGCT
 45 ACCGGGCCCAGACGTGCAGCGCGAGGCTCAGTGAGGACCTGTACCACCGCTGCCTCCAGAGC
 GTGCTTGAGGGCCTCGGCTCCCGAACCCAGCTGGCCATGCTACTCTTTGAACAGGAGCAGGC
 AACAGCACCT**TGA**CCCCACCCTGTCCCCGTGGGCCGTGGCATGGCCACACCCACCCCACTTC

FIGURE 43B

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TCCCCAAAACCAGAGCCACCTGCCAGCCTCGCTGGGCAGGGCTGGCCGTAGCCAGACCCCA
AGCTGGCCCCTGGTCCCCTCTCTGGCTCTGTGGGTCCCTGGGCTCTGGACAAGCACTGGGG
GACGTGCCCCCAGAGCCACCCACTTCTCATCCCAAACCCAGTTTCCCTGCCCCCTGACGCTG
CTGATTCGGGCTGTGGCCTCCACGTATTTATGCAGTACAGTCTGCCTGACGCCAGCCCTGCC
10 TCTGGGCCCTGGGGGCTGGGCTGTAGAAGAGTTGTTGGGGAAGGAGGGAGCTGAGGAGGGGG
CATCTCCCAACTTCTCCCTTTTGGACCCTGCCGAAGCTCCCTGCCTTTAATAAACTGGCCAA
GTGTGGAAAAA

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FIGURE 44

5 Signal Peptide: amino acids 1-15
N-glycosylation Sites: amino acids 138-141
amino acids 361-364

10 MRASLLLSVLRPAGPVAVGISLGFTLSLLSVTWVEEPCGPGPPQPGDSELP PRGNTNAARRP
NSVQPGAEREKPGAGEGAGENWEPRVLPYHPAQPGQAAKKAVRTRYISTELGIRQRLLVAVL
TSQTTLPTLGVAVNRTLGHRLERVVFLTGARGRRAPPGMAVVTLGEERPIGHLHLALRHLE
QHGDDEFDWFFLVPDTTYTEAHGLARLTGHLSLASAAHLYLGRPQDFIGGEPTPGRYCHGGFG
VLLSRMLLQQLRPHLEGCRNDIVSARPDEWLGRCIL DATGVGCTGDHEGVHYSHLELSPGEP
15 VQEGDPHFERSALTAHPVRDPVHMYQLHKAFARAELE RTYQEIQELQWEIQNTSHLAVDGDRA
AAWPVGIPAPSRPASRFEVLRWDYFTEQHAFSCADGSPRCPLRGADRADVADVLGTALEELN
RRYHPALRLQKQQLVNGYRRFDPARGMEYTL DLQLEALTPQGRRPLTRRVQLLRPLSRVEI
LPVPYVTEASRLTVLLPLAAAERDLAPGFLEAFATAALEPGDAAAALTLLLLYEPRQAQRVA
HADVFAPVKAHVAELERRFPGARVPWLSVQTAAPSPLRLMDLLSKKHPLDTLFLLAGPDTVL
20 TPDFLNRCRMHAISGWQAFFPMHFQAFHPGVAPPQGPPELGRDTGRFDRQAASEACFYNS
DYVAARGRLAAASEQEEELLES LDVYELFLHFSSSLHVLRAVEPALLQRYRAQTCSARLSEDL
YHRCLQSVLEGLGSRTQLAMLLFEQE QGNST

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FIGURE 45

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TGCAGCCGCTCGGGGGCCCTGTTGCTGGCCTTGCTGCTTCAGGCCTCCATGGAAGTGCGTGG
CTGGTGCCTGGAGAGCAGCCAGTGTGTCAGGACCTCACCACGGAAAGCAACCTGCTGGAGTGCA
TCCGGGCCTGCAAGCCCCGACCTCTCGGCCGAGACTCCCATGTTCCCGGGAAATGGCGACGAG
CAGCCTCTGACCGAGAACCCCCGGAAGTACGTCATGGGCCACTTCCGCTGGGACCGATTTCGG
10 CCGCCGCAACAGCAGCAGCAGCGGCAGCAGCGGCGCAGGGCAGAAGCGCGAGGACGTCTCAG
CGGGCGAAGACTGCGGGCCCGCTGCCTGAGGGCGGGCCCCGAGCCCCGCAGCGATGGTGCCAAG
CCGGGCCCCGCGCGAGGGCAAGCGCTCCTACTCCATGGAGCACTTCCGCTGGGGCAAGCCGGT
GGGCAAGAAGCGGCGCCCCAGTGAAGGTGTACCCTAACGGCGCCGAGGACGAGTCGGCCGAGG
CCTTCCCCCTGGAGTTCAAGAGGGAGCTGACTGGCCAGCGACTCCGGGAGGGAGATGGCCCC
15 GACGGCCCTGCCGATGACGGCGCAGGGGCCCAGGCCGACCTGGAGCACAGCCTGCTGGTGGC
GGCCGAGAAGAAGGACGAGGGCCCCCTACAGGATGGAGCACTTCCGCTGGGGCAGCCCCGCCCA
AGGACAAGCGCTACGGCGGTTTCATGACCTCCGAGAAGAGCCAGACGCCCCCTGGTGACGCTG
TTCAAAAACGCCATCATCAAGAACGCCTACAAGAAGGGCGAGTGAGGGCACAGCGGGCCCCA
GGGCTACCCTCCCCCAGGAGGTCGACCCCAAAGCCCCTTGCTCTCCCCTGCCCTGCTGCCGC
20 CTCCCAGCCTGGGGGGTCGTGGCAGATAATCAGCCTCTTA

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FIGURE 46

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Mature acth:

SYSMEHFRWGKPVGKKRRPVKVYPNGAEDESAEAFPLEFKREL TGQRLREGDGPDPGADDGA
GAQADLEHSLLVAAEKKDEGPYRMEHFRWGSPPKDKRYGGFMTSEKSQTPLVTLFKNAIIKN

10 AYKKGE

Truncated form:

ELTGQRLREGDGPDPGADDGAGA QADLEHSLLVAAEKKDEGPYRMEHFRWGSPPKDKRYGGF
15 MTSEKSQTPLVTLFKNAIIKNAYKKGE

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FIGURE 47

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ATGCATCTCTCCCAGCTGCTGGCCTGCGCCCTGCTGCTCACGCTGCTCTCCCTCCGGCCCTC
CGAAGCCAAGCCCGGGGCGCCGCCGAAGGTCCCGCGAACCCCGCCGGCAGAGGAGCTGGCCG
AGCCGCAGGCTGCGGGCGGCGGTGAGAAGAAGGGCGACAAGGCTCCCGGGGGCGGGGGCGCC
AATCTCAAGGGCGACCGGTGCGGACTGCTCCGGGACCTGCGCGTGGACACCAAGTCGCGGGC
10 AGCGTGGGCTCGCCTTCTGCAAGAGCACCCCAACGCGCGCAAATACAAAGGAGCCAACAAGA
AGGGCTTGTCCAAGGGCTGCTTCGGCCTCAAGCTGGACCGAATCGGCTCCATGAGCGGCCTG
GGATGT

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FIGURE 48

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GLSKGCFGLKLDRIGMSGGLGC

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FIGURE 49

5 ACC**ATG**AGCCCTCTGCTCCGCCGCCTGCTGCTCGCCGCACTCCTGCAGCTGGCCCCGCCCA
GGCCCCTGTCTCCCAGCCTGATGCCCCCTGGCCACCAGAGGAAAGTGGTGTTCATGGATAGATG
TGTATACTCGCGCTACCTGCCAGCCCCGGGAGGTGGTGGTGGCCCTTGACTGTGGAGCTCATG
GGCACCGTGGCCAAACAGCTGGTGCCCAGCTGCGTGACTGTGCAGCGCTGTGGTGGCTGCTG
10 CCCTGACGATGGCCTGGAGTGTGTGCCCCACTGGGCAGCACCAAGTCCGGATGCAGATCCTCA
TGATCCGGTACCCGAGCAGTCAGCTGGGGGAGATGTCCCTGGAAGAACACAGCCAGTGTGAA
TGCAGACCTAAAAAAAAGGACAGTGCTGTGAAGCCAGACAGCCCCAGGCCCTCTGCCCACG
CTGCACCCAGCACCAACAGCGCCCTGACCCCCGGACCTGCCGCTGCCGCTGCCGACGCCGCA
GCTTCCTCCGTTGCCAAGGGCGGGGCTTAGAGCTCAACCCAGACACCTGCAGGTGCCGGAAG
15 CTGCGAAGG**TGA**

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FIGURE 50

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Mature VEGF-b:

PVSQPDAPGHQRKVVS
WIDVYTRATCQPREVVVPLTVELMGTVAKQLVPSCVTVQRCGGCCP
DDGLECVPTGQHQVRMQILMIRYPSSQLGEMSLEEHSQCECRPKKKDSAVKPDSPRPLCPRC
10 TQHHQRPDPRTCRCRCRRRSFLRCQGRGLELNPDTCRCRKLRR

proVEGF-b:

15 MSPLLRLLLLAALLQLAPAQAPVSQPDAPGHQRKVVS
WIDVYTRATCQPREVVVPLTVELMGTVAKQLVPSCVTVQRCGGCCP
DDGLECVPTGQHQVRMQILMIRYPSSQLGEMSLEEHSQCECRPKKKDSAVKPDSPRPLCPRCTQHHQRPDPRTCRCRCRRRSFLRCQGRGLELNPDTCRCRKLRR

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FIGURE 51A

5
 GGGCTGAGGCACTGAGAGACCGGAAAGCCTGGCATTCCAGAGGGAGGGAAACGCAGCGGCAT
 CCCCAGGCTCCAGAGCTCCCTGGTGACAGTCTGTGGCTGAGC**ATGG**CCCTCCCAGCCCTGGG
 CCTGGACCCCTGGAGCCTCCTGGGCCTTTTCTCTTCCAAGTCTTCACTGCTTCAGCTGCTGCTGCCGA
 CGACGACCGCGGGGGGAGGCGGGCAGGGGCCCATGCCCAGGGTCAGATACTATGCAGGGGAT
 10 GAACGTAGGGCACTTAGCTTCTTCCACCAGAAGGGCCTCCAGGATTTTGACACTCTGCTCCT
 GAGTGGTGATGGAAATACTCTCTACGTGGGGGCTCGAGAAGCCATTCTGGCCTTGATATCC
 AGGATCCAGGGGTCCCCAGGCTAAAGAACATGATACCGTGGCCAGCCAGTGACAGAAAAAAG
 AGTGAATGTGCCTTTAAGAAGAAGAGCAATGAGACACAGTGTTCAACTTCATCCGTGTCCT
 GGTTTCTTACAATGTCACCCATCTCTACACCTGCGGCACCTTCGCCTTCAGCCCTGCTTGTA
 15 CCTTCATTGAACTTCAAGATTCCCTACCTGTTGCCCATCTCGGAGGACAAGGTCATGGAGGGA
 AAAGGCCAAAGCCCCTTTGACCCCGCTCACAAGCATACGGCTGTCTTGGTGGATGGGATGCT
 CTATTCTGGTACTATGAACAACTTCCTGGGCAGTGAGCCCATCCTGATGCGCACACTGGGAT
 CCCAGCCTGTCTTCAAGACCGACAACCTTCCTCCGCTGGCTGCATCATGACGCCTCCTTTGTG
 GCAGCCATCCCTTCGACCCAGGTCGTCTACTTCTTCTTCGAGGAGACAGCCAGCGAGTTTGA
 20 CTTCTTTGAGAGGCTCCACACATCGCGGGTGGCTAGAGTCTGCAAGAATGACGTGGGCGGCG
 AAAAGCTGCTGCAGAAGAAGTGGACCACCTTCCTGAAGGCCAGCTGCTCTGCACCCAGCCG
 GGGCAGCTGCCCTTCAACGTCATCCGCCACGCGGTCTGCTCCCCGCCGATTCTCCCACAGC
 TCCCCACATCTACGCAGTCTTCACCTCCCAGTGGCAGGTTGGCGGGACCAGGAGCTCTGCGG
 TTTGTGCCTTCTCTCTCTTGGACATTGAACGTGTCTTTAAGGGGAAATACAAAGAGTTGAAC
 25 AAAGAACTTCACGCTGGACTACTTATAGGGGCCCTGAGACCAACCCCCGCCAGGCAGTTG
 CTCAGTGGGCCCCCTCCTCTGATAAGGCCCTGACCTTCATGAAGGACCATTTCCTGATGGATG
 AGCAAGTGGTGGGGACGCCCCCTGCTGGTGAAATCTGGCGTGGAGTATACACGGCTTGCACTG
 GAGACAGCCCAGGGCCTTGATGGGCACAGCCATCTTGTATGTACCTGGGAACCACCACAGG
 GTCGCTCCACAAGGCTGTGGTAAGTGGGGACAGCAGTGTCTATCTGGTGGGAAGAGATTGAGC
 30 TGTTCCCTGACCCTGAACCTGTTTCGCAACCTGCAGCTGGCCCCCACCAGGGTGCAGTGTTT
 GTAGGCTTCTCAGGAGGTGTCTGGAGGGTGCCCCGAGCCAAGTGTAGTGTCTATGAGAGCTG
 TGTGGACTGTGTCTTGGCCGGGACCCCCACTGTGCCTGGGACCCTGAGTCCCGAACCTGTT
 GCCTCCTGTCTGCCCCCAACCTGAACTCCTGGAAGCAGGACATGGAGCGGGGGAACCCAGAG
 TGGGCATGTGCCAGTGGCCCCATGAGCAGGAGCCTTCGGCCTCAGAGCCGCCCGCAAATCAT
 35 TAAAGAAGTCTGGCTGTCCCCAACTCCATCCTGGAGCTCCCCCTGCCCCACCTGTCAGCCT
 TGGCCTCTTATTATTGGAGTCATGGCCCAGCAGCAGTCCCAGAAGCCTCTTCCACTGTCTAC
 AATGGCTCCCTCTTGCTGATAGTGCAGGATGGAGTTGGGGGTCTCTACCAGTGTGGGCAAC
 TGAGAATGGCTTTTCATACCCTGTGATCTCCTACTGGGTGGACAGCCAGGACCAGACCTGG
 CCTGGATCCTGAACTGGCAGGCATCCCCCGGGAGCATGTGAAGGTCCCGTTGACCAGGGTC
 40 AGTGGTGGGGCCGCCCTGGCTGCCCAGCAGTCCTACTGGCCCCACTTTGTCACTGTCACTGT
 CCTCTTTGCCTTAGTGCTTTCAGGAGCCCTCATCATCCTCGTGGCCTCCCCATTGAGAGCAC
 TCCGGGCTCGGGGCAAGGTTTCAGGGCTGTGAGACCCTGCGCCCTGGGGAGAAGGCCCCGTTA
 AGCAGAGAGCAACACCTCCAGTCTCCCAAGGAATGCAGGACCTCTGCCAGTGATGTGGACGC
 TGACAACAACCTGCCTAGGCCTGAGGTAGCT**TAA**ACTCTAGGCACAGGCCGGGGCTGCGGTG
 45 CAGGCACCTGGCCATGCTGGCTGGGCGGCCCAAGCACAGCCCTGACTAGGATGACAGCAGCA
 CAAAAGACCACCTTTCTCCCTGAGAGGAGCTTCTGCTACTCTGCATCACTGATGACACTCA
 GCAGGGTGATGCACAGCAGTCTGCCTCCCCTATGGGACTCCCTTCTACCAAGCACATGAGCT

FIGURE 51B

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CTCTAACAGGGTGGGGGCTACCCCCAGACCTGCTCCTACACTGATATTGAAGAACCTGGAGA
GGATCCTTCAGTTCTGGCCATTCCAGGGACCCTCCAGAAACACAGTGTTTCAAGAGACCCTA
AAAAACCTGCCTGTCCCAGGACCCTATGGTAATGAACACCAAACATCTAAACAATCATATGC
10 TAACATGCCACTCCTGGAAACTCCACTCTGAAGCTGCCGCTTTGGACACCAACACTCCCTTC
TCCCAGGGTCATGCAGGGATCTGCTCCCTCCTGCTTCCCTTACCAGTCGTGCACCGCTGACT
CCCAGGAAGTCTTTCCTGAAGTCTGACCACCTTTCTTCTTGCTTCAGTTGGGGCAGACTCTG
ATCCCTTCTGCCCTGGCAGAATGGCAGGGGTAATCTGAGCCTTCTTCACTCCTTTACCCTAG
CTGACCCCTTCACCTCTCCCCCTCCCTTTTCCTTTGTTTTGGGATTCAGAAAACCTGCTTGTC
15 AGAGACTGTTTATTTTTTTATTAAAAATATAAGGCTTAAAAAAA

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FIGURE 52

5 MALPALGLDPWSLLGLFLFQLLQLLLPTTTAGGGGQGPMPRVRYYAGDERRALSFFHQKGLQ
DFDTLLLSGDGNTLYVGAREAILALDIQDPGVPRLKNMIPWPASDRKKSECAFKKKSNETQC
FNFIRVLVSYNVTHLYTCGTFAFSPACTFIELQDSYLLPISEDKVMEGKGQSPFDPAHKHTA
VLVDGMLYSGTMNNFLGSEPILMRTLGSQPVLKTDNFLRWLHHDASFVAAIPSTQVVYFFFE
10 ETASEFDFFERLHTSRVARVCKNDVGGEKLLQKKWTTFLKAQLLCTQPGQLPFNVIRHAVLL
PADSPTAPHIYAVFTSQWQVGGTRSSAVCAFSLLDIERVFKGKYKELNKETSRWTTYRGPET
NPRPGSCSVGPSSDKALTFMKDHFMLDEQVVGTPLLVKSGVEYTRLAVETAQGLDGHSHLVM
YLGTTTGSLSHKAVVSGDSSAHLVEEIQLPDPEPVRNLQLAPTQGAVFVGFSGGVWRVPRAN
CSVYESCVDCVLARDPHCAWDPESTRCCLLSAPNLNSWKQDMERGNPEWACASGPMRSRSLRP
15 QSRPQIIKEVLAVPNSILELPCPHLSALASYWSHGPAAVPEASSTVYNGSLLLIVQDGVGG
LYQCWATENGFSYPVISYWVDSQDQTLALDPELAGIPREHVKVPLTRVSGGAALAAQQSYWP
HFVTVTVLFAVLVLSGALIILVASPLRALRARGKVQGCETLRPGEKAPLSREQHLQSPKECRT
SASDVDADNNCLGTEVA

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FIGURE 53

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CCCAGCGCACCCGCACC**ATG**GCCGGCCCCAGCCTCGCTTGCTGTCTGCTCGGCCTCCTGGCG
CTGACCTCCGCCTGCTACATCCAGAACTGCCCCCTGGGAGGCAAGAGGGCCGCGCCGGACCT
CGACGTGCGCAAGTGCCCTCCCCTGCGGCCCCGGGGGCAAAGGCCGCTGCTTCGGGCCCCAATA
TCTGCTGCGCGGAAGAGCTGGGCTGCTTCGTGGGCACCGCCGAAGCGCTGCGCTGCCAGGAG
10 GAGAACTACCTGCCGTCGCCCTGCCAGTCCGGCCAGAAGGCGTGCGGGAGCGGGGGCCGCTG
CGCGGTCTTGGGCCTCTGCTGCAGCCCGGACGGCTGCCACGCCGACCCTGCCTGCGACGCGG
AAGCCACCTTCTCCCAGCGCT**TGA**AACTTGATGGCTCCGAACACCCTCGAAGCGCGCCACTCG
CTTCCCCCATAGCCACCCCAGAAATGGTGAAAATAAAATAAAGCAGGTTTTTCTCCTCT

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FIGURE 54

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Mature oxytocin:

CYIQNCPLGGKRAAPDL DVRKCLPCGPGGKGRCFGPNICCAEELGCFVGTAELRCQEENYL
PSPCQSGQKACGSGGRC AVLGLCCSPDGCHADPACDAEATFSQR

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Pro-oxytocin:

MAGPSLACCLLGLLALTSACYIQNCPLGGKRAAPDL DVRKCLPCGPGGKGRCFGPNICCAEE
LGCFVGTAELRCQEENYLPSPCQSGQKACGSGGRC AVLGLCCSPDGCHADPACDAEATFSQR

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FIGURE 55A

5 CTTGCAGCA**ATGG**CTTGGATTAGATCCACTTGCATTCTCTTTTTTACCTTGCTTTTTTGCCCA
CATAGCAGCTGTACCGATTAAGCATCTTCCTGAAGAAAATGTACATGATGCAGATTTTGGTG
AACAGAAGGATATTTTCAGAAATCAATTTAGCTGCAGGCTTGGACCTCTTTCAAGGGGACATC
CTCTTGAGAAATCCAGAAATGGCCTGAGAGACCCAAACACCAGGTGGACGTTCCCCATTCC
10 TTACATCTTGGCTGATAATTTGGGGCTGAATGCTAAAGGAGCCATTCTGTATGCCTTTGAGA
TGTTCCGTCTCAAGTCCTGTGTGGATTTCAAGCCCTATGAAGGAGAGAGCTCATATATCATA
TTTCAACAGTTTGATGGGTGCTGGTCTGAGGTTGGTGACCAACATGTGGGACAGAACATTTTC
CATTGGCCAAGGATGTGCCTATAAGGCCATCATAGAACACGAGATCCTGCATGCTTTGGGAT
TTTACCACGAGCAGTCAAGGACGGACCGGGATGATTATGTGAACATCTGGTGGGACCAAATT
15 CTTTCAGGTTACCAGCACAACCTTTGACACCTATGATGATAGCTTAATCACAGACCTCAATAC
ACCCTATGATTATGAGTCTTTGATGCACTACCAGCCTTTCTCATTTAACAAGAATGCAAGTG
TTCCCAACCATCACAGCCAAGATCCCTGAGTTTAACTCCATTATCGGACAACGCCTGGATTTTC
AGTGCCATTGATTTAGAGAGGGCTGAACCGAATGTACAATTGCACCACAACCTCACACTCTTTT
GGACCACTGTACTTTTGAGAAGGCAAACATCTGTGGAATGATTCAGGGCACCAGAGATGACA
20 CTGACTGGGCCCATCAGGACAGTGCTCAGGCTGGAGAAGTGGATCACACCTTGTTGGGACAA
TGCACAGGTGCCGGCTACTTCATGCAGTTCAGCACCAGCTCGGGGTCCGCGGAAGAGGCAGC
CCTACTGGAGTCTCGGATTCTTTACCCAAAGAGGAAGCAGCAGTGCCTGCAATTTTCTATA
AAATGACGGGAAGTCCTTCAGACAGACTCGTTGTCTGGGTGAGGAGGGATGACAGCACAGGC
AATGTTTCGCAAGTTGGTGAAGGTGCAGACTTTTCAAGGAGATGATGACCACAATTGGAAAAT
25 TGCCCATGTGGTGCTCAAAGAGGAACAGAAGTTTCGCTACCTTTTCCAGGGCACAAAAGGCG
ACCCTCAGAACTCAACTGGGGGAATTTACCTAGATGACATCACTCTGACAGAAACCCCTGC
CCCACAGGGGTCTGGACAGTCCGGAATTTCTCCCAAGTCCTTGAGAACACCAGCAAAGGGGA
CAAGCTTCAGAGCCCTCGATTCTACAATTTCGAGAGGGATATGGTTTTTGGGGTAACTTTATACC
CAAATAGCAGAGAAAGCTCTGGTTACTTGAGACTTGCTTTTTCATGTGTGCAGTGGGGAGAAC
30 GATGCTATCCTGGAGTGGCCGGTAGAAAACAGACAGGTGATAATTACCATCCTTGACCAGGA
GCCTGATGTCCGGAACAGGATGTCCTCAAGCATGGTGTTCCTACTACCTCGAAGTCGCACACAT
CTCCAGCGATAAATGACACTGTCATCTGGGACAGGCCGTCCAGGGTGGGAACCTATCATACA
GACTGTAATTGTTTTAGAACATCGACTTGGGCTGGAGTGGTTTCATTTCCCAACAAATGCT
GAAAAGGAGGAGTTTCTGAAAAATGATGACCTCATCATATTTGTGGACTTTGAAGATATCA
35 CCCACCTCAGCCAGACTGAAGTTCCTCTAAAGGCAAAGACTGAGCCCCAAGGCCTCATT
CTCCAAGGCCAGGAGCAGCAGGTCTCCGAAGAAGGTTTCGGGAAAGGCCATGTTAGAGGAAGC
CCTACCTGTCAGCCTGAGCCAGGGGCAGCCAGCCGACAGAAGCGGTTCGGTGGAGAACACAG
GCCCCCTGGAGGACCATAACTGGCCACAGTACTTCAGAGACCCATGTGACCCAAACCTTGC
CAAAATGACGGCATCTGTGTGAACGTGAAGGGGATGGCGAGCTGCAGGTGCATCTCTGGACA
40 TGCTTTCTTCTACACGGGGGAGCGCTGTCAGTCGGCCGAGGTGCACGGCAGTGTCTGGGCA
TGGTGATCGGAGGCACGGCTGGCGTGATCTTCTTGACCTTCTCCATCATCGCCATCCTTTCC
CAAAGGCCAAGGAAG**TGA**CCTGCCTGCTGGCATTGGCCAGACCACAGCAGCACCTCCTCCAT
GCAGGCCTTAACCTTTCCCATGTTCAATGCAGTTTGGGGCAGCTTTTTTATCAGCCTTGCTTT
GGATAGGACCTCCAAGGACTAAGCCTCCAGCCCCATGTGTGACCCTTGTCTATCTCTCTGCCC
45 CACATAATTATGTTACTTTGCTATGTGCTCCTAATGTATCTAGTGTGTCTGTGACAACACT
CATCACACTTCATTGTAAATCACTTGTTTTATTGACTGTCTTTCTTATAGACTGTAAGCTCC
ATGAGGGCAGGCACATGTTGTTCTCATTGACCGTGCTGGCCCCAGTGCCTAGATGCATGGCT

FIGURE 55B

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GGCACATTGTTGGCACTCAACAATGGTTGAATGAATAAAACAATAAATGAATGAATAACTAA
GATATAGAACTCTCATTTATATTGCAGATTGAATATATATGATGAAATTCTTATGTTGAAT
ATGTTAGAATCAAATACTCATTTTTCATTAGATACAGTAGTGTTCATCACTCTTTTAAGATCT
10 TGTAAAGATTTCAAATAAAGGTACTTCTGGCGAGCCAGGCTGCACAGCATTTGCTTTCCTC
TGAGATTCTAAGAGAAGGCCTTTAATAAATTTAATAAATATTGAGTTAGCAAAAAAAAAAAAA
AAAAAA

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FIGURE 56

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MAWIRSTCILFFTLLFAHIAAVPIKHLPEENVHDADFGEQKDI SEINLAAGLDL FQGDILLQ
KSRNGLRDPNTRWTFPIPYILADNLGLNAKGAILYAFEMFRLKSCVDFKPYEGESSYIIFQQ
FDGCWSEVGDQHVGQNISIGQGCA YKAIIEHEILHALGFYHEQSRTDRDDYVNIWWDQILSG
YQHNFD TYDDSLITDLNTPYDYESLMHYQPFSFNKNASVPTITAKIPEFNSIIGQRLDFSAI
10 DLERLNRMYNCTTTHTLLDHCTFEKANICGMIQGTRDDTDWAHQDSAQAGEVDHTLLGQCTG
AGYFMQFSTSSGSAEEAALLESRI LYPKRKQQCLQFFYKMTGSPSDRLVWVRRDDSTGNVR
KLVKVQTFQGD DDHNWKIAHVVLKEEQKFRYLFQGTKGDPQNSTGGIYLDDITLTETPCPTG
VWTVRNFSQVLENTSKGDKLQSPRFYNSEGYGFGVTLYPNSRESSGYLR LAFHVCSGENDAI
LEWPVENRQVIITILDQEPDVRNRMSSSMVF TTSKSHTSPAINDTVIWDRPSRVGTYHTDCN
15 CFRSIDLGWSGFISHQMLKRRSFLKNDDL IIFVDFEDITHLSQTEVPSK GKRLSPQGLILQG
QEQQVSEEGSGKAMLEEALPVSLSQGQPSRQKRSVENTGPLEDHNWPQYFRDPCDPNPCQND
GICVNVKGMASCRCISGHAF FYTGERCQSAEVHGSVLGMVIGGTAGVIFLTFSIIAILSQRPRK

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FIGURE 57

5 **ATG**CGCTGCTCGCCGGGAGGCGTCTGGCTGGCGCTGGCCGCGTCGCTCCTGCACGTGTCCCT
GCAAGGCGAGTTCCAGAGGAAGCTTTACAAGGAGCTGGTCAAGAACTACAATCCCTTGGAGA
GGCCCGTGGCCAATGACTCGCAACCACTCACCGTCTACTTCTCCCTGAGCCTCCTGCAGATC
ATGGACGTGGATGAGAAGAACCAAGTTTAAACCACCAACATTTGGCTGCAAATGTCTTGGAC
10 AGATCACTATTTACAGTGGAATGTGTCAGAATATCCAGGGGTGAAGACTGTTTCGTTTCCCAG
ATGGCCAGATTTGGAAACCAGACATTCTTCTCTATAACAGTGCTGATGAGCGCTTTGACGCC
ACATTCCACACTAACGTGTTGGTGAATTCTTCTGGGCATTGCCAGTACCTGCCTCCAGGCAT
ATTCAAGAGTTCCCTGCTACATCGATGTACGCTGGTTTTCCCTTTGATGTGCAGCACTGCAAAC
TGAAGTTTGGGTCTCCTGGTCTTACGGAGGCTGGTTCCTTGGATCTGCAGATGCAGGAGGCAGAT
15 ATCAGTGGCTATATCCCCAATGGAGAATGGGACCTAGTGGGAATCCCCGGCAAGAGGAGTGA
AAGGTTCTATGAGTGCTGCAAAGAGCCCTACCCCGATGTCACCTTCACAGTGACCATGCGCC
GCAGGACGCTCTACTATGGCCTCAACCTGCTGATCCCCCTGTGTGCTCATCTCCGCCCTCGCC
CTGCTGGTGTTCCCTGCTTCCTGCAGATTCCGGGGAGAAGATTTCCCTGGGGATAACAGTCTT
ACTCTCTCTTACCGTCTTCATGCTGCTCGTGGCTGAGATCATGCCCCGAACATCCGATTTCGG
20 TACCATTGATAGCCCACTACTTCGCCAGCACCATGATCATCGTGGGCCTCTCGGTGGTGGTG
ACGGTGATCGTGCTGCAGTACCACCACCACGACCCCGACGGGGGCAAGATGCCCAAGTGGAC
CAGAGTCATCCTTCTGAACTGGTGCGCGTGGTTCCTGCGAATGAAGAGGCCCGGGAGGACA
AGGTGCGCCCCGGCCTGCCAGCACAAAGCAGCGGCGCTGCAGCCTGGCCAGTGTGGAGATGAGC
GCCGTGGCGCCCGCCGCCCGCCAGCAACGGGAACCTGCTGTACATCGGCTTCCGCGGCCTGGA
25 CGGCGTGCACTGTGTCCCGACCCCCGACTCTGGGGTAGTGTGTGGCCGCATGGCCTGCTCCC
CCACGCACGATGAGCACCTCCTGCACGGTGGGCAACCCCCGAGGGGGACCCGACTTGGCC
AAGATCCTGGAGGAGGTCCGCTACATTGCCAACCGCTTCCGCTGCCAGGACGAAAGCGAGGC
GGTCTGCAGCGAGTGGAAGTTCGCCGCCTGTGTGGTGGACCGCCTGTGCCTCATGGCCTTCT
CGGTCTTCACCATCATCTGCACCATCGGCATCCTGATGTCCGGCTCCCAACTTCGTGGAGGCC
30 GTGTCCAAAGACTTTGCG**TAA**

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FIGURE 58

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MRCSPGGVWLALAASLLHVSLQGEFQRKLYKELVKNYNPLERPVANDSQPLTVYFSLSLLQI
MDVDEKNQVLTTNIWLQMSWTDHYLQWNVSEYPGVKTVRFPDGQIWKPDILLYNSADERFDA
TFHTNVLVNSSGHCQYLPPGIFKSSCYIDVRWFPFDVQHCKLKFGSWSYGGWSLDLQMQEAD
10 ISGYIPNGEWDLVGIPGKRSERFYECCKEPYPDVTFVTMRRRTLYYGLNLLIPCVLISALA
LLVFLLPADSGEKISLGITVLLSLTVFMLLVAEIMPATSDSVPLIAQYFASTMIIVGLSVVV
TVIVLQYHHHDPDGGKMPKWTRVILLNWCAWFLRMKRPGEDKVRPACQHKQRRCSLASVEMS
AVAPPPASNGNLLYIGFRGLDGVHCVPTPDSGVVCGRMACSPTHDEHLLHGGQPPEGDPDLA
KILEEVRYIANRFRCQDESEAVCSEWKFAACVVDRLCLMAFSVFTIICTIGILMSAPNFVEA
15 VSKDFA

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FIGURE 59A

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ACTTGCCTGTCACTCAGCCTGGACGCGCTTCTTCGGGTCGCGGGTGCACTCCGGCCCCGGCT
 CCCGCCTCGGCCCCG**ATGG**ACGCCGCGTTCTCTCGTCCTCGGGCTGTTGGCCAGAGCCT
 CTGCCTGTCTTTGGGGGTTCTTGATGGAGGAGGCCACCACCCTGTACCCCTGGCGCCGGG
 10 CGCCTGCCCTGAGCCGCGTGCGGAGGGCCTGGGTCATCCCCCGATCAGCGTATCCGAGAAC
 CACAAGCGTCTCCCCTACCCCTGGTTCAAGTCGGACAAGCAGCAGCTGGGCAGCGT
 CATCTACAGCATCCAGGGACCCGGCGTGATGAGGAGCCCCGGGGCGTCTTCTCTATCGACA
 AGTTCACAGGGAAGGTCTTCTCAATGCCATGCTGGACCGCGAGAAGACTGATCGCTTCAGG
 CTAAGAGCGTTTGCCCTGGACCTGGGAGGATCCACCCTGGAGGACCCACGGACCTGGAGAT
 15 TGTAAGTTGTGGATCAGAATGACAACCGGCCAGCCTTCTTGACAGGAGCGTTCACTGGCCGCG
 TGCTGGAGGGTGAGTCCCAGGCACCTATGTGACCAGGGCAGAGGCCACAGATGCCGACGAC
 CCCGAGACGGACAACGCAGCGCTGCGGTTCTCCATCCTGCAGCAGGGCAGCCCCGAGCTCTT
 CAGCATCGACGAGCTCACAGGAGAGATCCGCACAGTGCAAGTGGGGCTGGACCGCGAGGTGG
 TCGCGGTGTACAATCTGACCCTGCAGGTGGCGGACATGTCTGGAGACGGCCTCACAGCCACT
 20 GCCTCAGCCATCATCACCTTGATGACATCAATGACAATGCCCCGAGTTCACCAGGGATGA
 GTTCTTCATGGAGGCCATAGAGGCCGTACGCGGAGTGATGTGGGACGCCTGGAAGTGGAGG
 ACAGGGACCTGCCAGGCTCCCCAACTGGGTGGCCAGGTTACCATCCTGGAAGGCGACCCC
 GATGGGCAGTTCACCATCCGCACGGACCCCAAGACCAACGAGGGTGTCTGTCCATTGTGAA
 GGCCCTGGACTATGAGAGCTGTGAACACTACGAACCTCAAAGTGTGGTGCAGAATGAGGCCC
 25 CGCTGCAGGCGGCTGCCCTTAGGGCTGAGCGGGGCCAGGCCAAGGTCCGCGTGCATGTGCAG
 GACACCAACGAGCCCCCGTGTTCAGGAGAACCCACTTCGGACAGCCTAGCAGAGGGGGC
 ACCCCCAGGCACTCTGGTGGCCACCTTCTCTGCCCCGGGACCTTGACACAGAGCAGCTGCAGA
 GGCTCAGTACTCCAAGGACTACGACCCGGAAGACTGGCTGCAAGTGGACGCAGCCACTGGC
 CGGATCCAGACCCAGCACGTGCTCAGCCCGGCGTCCCCCTTCTCAAGGGCGGCTGGTACAG
 30 AGCCATCGTCTTGCCAGGATGACGCCTCCAGCCCCGCACCGCCACCGGCACCCTGTCCA
 TCGAGATCCTGGAGGTGAACGACCATGCACCTGTGCTGGCCCCGCCGCCCGGGCAGCCTG
 TGCAGCGAGCCACACCAAGGCCAGGCCTCCTCCTGGGCGCCACGGATGAGGACCTGCCCCC
 CCACGGGGCCCCCTTCCACTTCCAGCTGAGCCCCAGGCTCCAGAGCTCGGCGGAACTGGA
 GCCTCAGCCAGGTCAACGTGAGCCACGCGCGCCTGCGGGCGCGACACCAGGTCCCCGAAGGC
 35 CTGACCGCCTCAGCCTGCTGCTCCGGGACTCGGGGCAGCCGCCCCAGCAGCGCGAGCAGCC
 TCTGAACGTGACCGTGTGCCGCTGCGGCAAGGACGGCGTCTGCCTGCCGGGGGGCCGCAGCGC
 TGCTGGCGGGGGGCACAGGCCTCAGCCTGGGCGCACTGGTTCATCGTGCTGGCCAGCGCCCTC
 CTGCTGCTGGTGCTGGTCTGCTCGTGGAAGTGGTCTCGGGG
 CAAGGGGCTGCTGCACGGCCCCCAGGACGACCTTCGAGACAATGTCCTCAACTACGATGAGC
 40 AAGGAGGCGGGGAGGAGGACCAGGACGCCTACGACATCAGCCAGCTGCGTCACCCGACAGCG
 CTGAGCCTGCCTCTGGGACCGCCGCCACTTCGCAGAGATGCCCCGAGGGCCGCCTGCACCC
 CCAGCCACCCCGAGTGCTGCCCACCAGCCCCCTGGACATCGCCGACTTCATCAATGATGGCT
 TGGAGGCTGCAGATAGTGACCCAGTGTGCCGCTTACGACACAGCCCTCATCTATGACTAC
 GAGGGTGACGGCTCGGTGGCGGGGACGCTGAGCTCCATCCTGTCCAGCCAGGGCGATGAGGA
 45 CCAGGACTACGACTACCTCAGAGACTGGGGGCCCCGCTTCGCCCGGCTGGCAGACATGTATG
 GGCACCCGCTGCGGGTTGGAGTACGGGGCCAGATGGGACCACCAGGCCAGGGAGGGTCTTTCT
 CCTGGGGCACTGCTACCCAGACACAGAGGCCGGACAGCC**TGA**CCCTGGGGCGCAACTGGACA

FIGURE 59B

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TGCCACTCCCCGGCCTCGTGGCAGTGATGGCCCCTGCAGAGGCAGCCTGAGGTCACCGGGCC
CGACCCCCCTGGGCCTGGGGCAGCCTCCTTCCTGTAGGCGAGGGCCCAAGTCTGGGGGCAGA
ACCTGAGTGTGGATGGGGCGGCCAGGAAGAGGCCCTTCCTGCCGGGGTGGGAAGAGTTTCT
10 CTCCATCGGCCCCATGCGGGTCACCTCCCTAGTCCCACCTTTGCCTCCTACCAGTGAACCTC
ATCTTTGTATGAAAGACAGCAACCTCCTGGGTAAATCTGAATG

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FIGURE 60

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MDAAFLLVLGLLAQSLCLSLGVPGWRRPTTLYPWRRAPALSRVRRRAWVIPPISVSENHKRLP
YPLVQIKSDKQQLGSGVIYSIQGPGVDEEPRGVFSIDKFTGKVFLNAMLDREKTDREKTLRAFA
LDLGGSTLEDPTDLEIVVVDQNDNRPAFLQEAFTGRVLEGAVPGTYVTRAEATDADDPETDN
10 AALRFSILQQGSPELFSIDELTGEIRTVQVGLDREVVAVYNLTQVADMSGDGLTATASAI I
TLDDINDNAPEFTRDEFFMEAIEAVSGVDVGRLEVEDRDLPGSPNWWARFTILEGDPDGQFT
IRTDPKTNEGVLSIVKALDYESCEHYELKVSQNEAPLQAAALRAERGQAKVRVHVQDTNEP
PVFQENPLRTSLAEGAPPGTLVATFSARDPDTEQLQRLSYSKYDPEDWLQVDAATGRIQTQ
HVLSPASPFLKGGWYRAIVLAQDDASQPRATATGTLSEIELEVNDHAPVLAPPPPGSLCSEPH
15 QGPGLLLGATDEDLPPHGAPFHFQLSPRLPELGRNWSLSQVNVSHARLRPRHQVPEGLHRLS
LLLRDSGQPPQREQPLNVTVCRCGKDGVCCLPGAAALLAGGTGLSLGALVIVLASALLLV
VLLVALRARFWKQSRGKGLLHGPQDDLRLDNVLYDEQGGGEEDQDAYDISQLRHPTALSLPL
GPPPLRRDAPQGRLLHPQPPRVLPTSPLDIADFINDGLEAADS DPSVPPYDTALIYDYEGDGS
VAGTLSSILSSQGEDQDYDYLRDWGPRFARLADMYGHPCGLEYGARWDHQAREGLSPGALL
20 PRHRGRTA

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```
PRO                XXXXXXXXXXXXXXXXX      (Length = 15 amino acids)
Comparison Protein  XXXXXYYYYYYYYY       (Length = 12 amino acids)
```

% amino acid sequence identity =

10 (the number of identically matching amino acid residues between
the two polypeptide sequences as determined by ALIGN-2) divided
by (the total number of amino acid residues of the PRO
polypeptide) =

15 5 divided by 15 = 33.3%

•

% amino acid sequence identity =

(the number of identically matching amino acid residues between
10 the two polypeptide sequences as determined by ALIGN-2) divided
by (the total number of amino acid residues of the PRO
polypeptide) =

5 divided by 10 = 50%

FIGURE 61C

PRO-DNA	NNNNNNNNNNNNNNNN	(Length = 14 nucleotides)
Comparison DNA	NNNNNNLLLLLLLLLLLL	(Length = 16
5 nucleotides)		

% nucleic acid sequence identity =

(the number of identically matching nucleotides between the two
10 nucleic acid sequences as determined by ALIGN-2) divided by (the
total number of nucleotides of the PRO-DNA nucleic acid sequence)
=

6 divided by 14 = 42.9%

FIGURE 61D

PRO-DNA NNNNNNNNNNNNNN (Length = 12 nucleotides)
Comparison DNA NNNNLLLTVV (Length = 9 nucleotides)

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% nucleic acid sequence identity =

(the number of identically matching nucleotides between the two
nucleic acid sequences as determined by ALIGN-2) divided by (the
10 total number of nucleotides of the PRO-DNA nucleic acid sequence)
=

4 divided by 12 = 33.3%

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FIGURE 62A

day.h

day.h

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/* C-C increased from 12 to 15
 * Z is average of EQ
 * B is average of ND
 * match with stop is _M; stop-stop = 0; J (joker) match = 0
 */
#define _M      -8      /* value of a match with a stop */

int      _day[26][26] = {
/* A */      { 2, 0, -2, 0, 0, -4, 1, -1, -1, 0, -1, -2, -1, 0, _M, 1, 0, -2, 1, 1, 0, 0, -6, 0, -3, 0},
/* B */      { 0, 3, -4, 3, 2, -5, 0, 1, -2, 0, 0, -3, -2, 2, _M, -1, 1, 0, 0, 0, 0, -2, -5, 0, -3, 1},
/* C */      {-2, -4, 15, -5, -5, -4, -3, -3, -2, 0, -5, -6, -5, -4, _M, -3, -5, -4, 0, -2, 0, -2, -8, 0, 0, -5},
/* D */      { 0, 3, -5, 4, 3, -6, 1, 1, -2, 0, 0, -4, -3, 2, _M, -1, 2, -1, 0, 0, 0, -2, -7, 0, -4, 2},
/* E */      { 0, 2, -5, 3, 4, -5, 0, 1, -2, 0, 0, -3, -2, 1, _M, -1, 2, -1, 0, 0, 0, -2, -7, 0, -4, 3},
/* F */      {-4, -5, -4, -6, -5, 9, -5, -2, 1, 0, -5, 2, 0, -4, _M, -5, -5, -4, -3, -3, 0, -1, 0, 0, 7, -5},
/* G */      { 1, 0, -3, 1, 0, -5, 5, -2, -3, 0, -2, -4, -3, 0, _M, -1, -1, -3, 1, 0, 0, -1, -7, 0, -5, 0},
/* H */      {-1, 1, -3, 1, 1, -2, -2, 6, -2, 0, 0, -2, -2, 2, _M, 0, 3, 2, -1, -1, 0, -2, -3, 0, 0, 2},
/* I */      {-1, -2, -2, -2, -2, 1, -3, -2, 5, 0, -2, 2, 2, -2, _M, -2, -2, -2, -1, 0, 0, 4, -5, 0, -1, -2},
/* J */      { 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0},
/* K */      {-1, 0, -5, 0, 0, -5, -2, 0, -2, 0, 5, -3, 0, 1, _M, -1, 1, 3, 0, 0, 0, -2, -3, 0, -4, 0},
/* L */      {-2, -3, -6, -4, -3, 2, -4, -2, 2, 0, -3, 6, 4, -3, _M, -3, -2, -3, -3, -1, 0, 2, -2, 0, -1, -2},
/* M */      {-1, -2, -5, -3, -2, 0, -3, -2, 2, 0, 0, 4, 6, -2, _M, -2, -1, 0, -2, -1, 0, 2, -4, 0, -2, -1},
/* N */      { 0, 2, -4, 2, 1, -4, 0, 2, -2, 0, 1, -3, -2, 2, _M, -1, 1, 0, 1, 0, 0, -2, -4, 0, -2, 1},
/* O */      { _M, _M, _M, _M, _M, _M, _M, _M, _M, _M, _M, _M, _M, _M, 0, _M, _M, _M, _M, _M, _M, _M, _M, _M, _M},
/* P */      {-1, -1, -3, -1, -1, -5, -1, 0, -2, 0, -1, -3, -2, -1, _M, 6, 0, 0, 1, 0, 0, -1, -6, 0, -5, 0},
/* Q */      { 0, 1, -5, 2, 2, -5, -1, 3, -2, 0, 1, -2, -1, 1, _M, 0, 4, 1, -1, -1, 0, -2, -5, 0, -4, 3},
/* R */      {-2, 0, -4, -1, -1, -4, -3, 2, -2, 0, 3, -3, 0, 0, _M, 0, 1, 6, 0, -1, 0, -2, 2, 0, -4, 0},
/* S */      { 1, 0, 0, 0, 0, -3, 1, -1, -1, 0, 0, -3, -2, 1, _M, 1, -1, 0, 2, 1, 0, -1, -2, 0, -3, 0},
/* T */      { 1, 0, -2, 0, 0, -3, 0, -1, 0, 0, 0, -1, -1, 0, _M, 0, -1, -1, 1, 3, 0, 0, -5, 0, -3, 0},
/* U */      { 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0},
/* V */      { 0, -2, -2, -2, -2, -1, -1, -2, 4, 0, -2, 2, 2, -2, _M, -1, -2, -2, -1, 0, 0, 4, -6, 0, -2, -2},
/* W */      {-6, -5, -8, -7, -7, 0, -7, -3, -5, 0, -3, -2, -4, -4, _M, -6, -5, 2, -2, -5, 0, -6, 17, 0, 0, -6},
/* X */      { 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0},
/* Y */      {-3, -3, 0, -4, -4, 7, -5, 0, -1, 0, -4, -1, -2, -2, _M, -5, -4, -4, -3, -3, 0, -2, 0, 0, 10, -4},
/* Z */      { 0, 1, -5, 2, 3, -5, 0, 2, -2, 0, 0, -2, -1, 1, _M, 0, 3, 0, 0, 0, 0, -2, -6, 0, -4, 4}
};

```

FIGURE 62B

nw.h

nw.h

```
#include <stdio.h>
#include <ctype.h>

#define MAXJMP 16      /* max jumps in a diag */
#define MAXGAP 24      /* don't continue to penalize gaps larger than this */
#define JMPS 1024      /* max jmps in an path */
#define MX 4           /* save if there's at least MX-1 bases since last jmp */

#define DMAT 3         /* value of matching bases */
#define DMIS 0         /* penalty for mismatched bases */
#define DINSO 8        /* penalty for a gap */
#define DINS1 1        /* penalty per base */
#define PINSO 8        /* penalty for a gap */
#define PINS1 4        /* penalty per residue */

struct jmp {
    short          n[MAXJMP];      /* size of jmp (neg for dely) */
    unsigned short x[MAXJMP];      /* base no. of jmp in seq x */
};                                /* limits seq to 216 -1 */

struct diag {
    int            score;          /* score at last jmp */
    long           offset;         /* offset of prev block */
    short          ijmp;          /* current jmp index */
    struct jmp     jp;            /* list of jmps */
};

struct path {
    int            spc;            /* number of leading spaces */
    short          n[JMPS];        /* size of jmp (gap) */
    int            x[JMPS];        /* loc of jmp (last elem before gap) */
};

char              *ofile;          /* output file name */
char              *namex[2];       /* seq names: getseqs0 */
char              *prog;           /* prog name for err msgs */
char              *seqx[2];        /* seqs: getseqs0 */
int               dmax;            /* best diag: nw0 */
int               dmax0;           /* final diag */
int               dna;             /* set if dna: main0 */
int               endgaps;         /* set if penalizing end gaps */
int               gapx, gapy;       /* total gaps in seqs */
int               len0, len1;       /* seq lens */
int               ngapx, ngapy;     /* total size of gaps */
int               smax;            /* max score: nw0 */
int               *xbm;            /* bitmap for matching */
long              offset;          /* current offset in jmp file */
struct diag       *dx;            /* holds diagonals */
struct path       pp[2];          /* holds path for seqs */

char              *calloc(), *malloc(), *index(), *strcpy();
char              *getseq(), *g_calloc();
```

FIGURE 62C

nw.c

nw.c

```

/* Needleman-Wunsch alignment program
 *
 * usage: prog file1 file2
 *   where file1 and file2 are two dna or two protein sequences.
 *   The sequences can be in upper- or lower-case and may contain ambiguity
 *   Any lines beginning with '!', '>' or '<' are ignored
 *   Max file length is 65535 (limited by unsigned short x in the jmp struct)
 *   A sequence with 1/3 or more of its elements ACGTU is assumed to be DNA
 *   Output is in the file "align.out"
 *
 * The program may create a tmp file in /tmp to hold info about traceback.
 * Original version developed under BSD 4.3 on a vax 8650
 */
#include "nw.h"
#include "day.h"

static dbval[26] = {
1,14,2,13,0,0,4,11,0,0,12,0,3,15,0,0,0,5,6,8,8,7,9,0,10,0
};

static pbval[26] = {
1, 2*(1<<('D'-'A'))*(1<<('N'-'A')), 4, 8, 16, 32, 64,
128, 256, 0xFFFFFFFF, 1<<10, 1<<11, 1<<12, 1<<13, 1<<14,
1<<15, 1<<16, 1<<17, 1<<18, 1<<19, 1<<20, 1<<21, 1<<22,
1<<23, 1<<24, 1<<25*(1<<('E'-'A'))*(1<<('Q'-'A'))
};

main(ac, av)
int ac;
char *av[];
{
    prog = av[0];
    if (ac != 3) {
        fprintf(stderr, "usage: %s file1 file2\n", prog);
        fprintf(stderr, "where file1 and file2 are two dna or two protein sequences.\n");
        fprintf(stderr, "The sequences can be in upper- or lower-case\n");
        fprintf(stderr, "Any lines beginning with '!' or '<' are ignored\n");
        fprintf(stderr, "Output is in the file \"align.out\"\n");
        exit(1);
    }
    namex[0] = av[1];
    namex[1] = av[2];
    seqx[0] = getseq(namex[0], &len0);
    seqx[1] = getseq(namex[1], &len1);
    xbm = (dna)? _dbval : _pbval;

    endgaps = 0; /* 1 to penalize endgaps */
    ofile = "align.out"; /* output file */

    nw(); /* fill in the matrix, get the possible jmps */
    readjmps(); /* get the actual jmps */
    print(); /* print stats, alignment */

    cleanup(0); /* unlink any tmp files */
}

```

main

NW.C

NW.C

```

/* do the alignment, return best score: main0
 * dna: values in Fitch and Smith, PNAS, 80, 1382-1386, 1983
 * pro: PAM 250 values
 * When scores are equal, we prefer mismatches to any gap, prefer
 * a new gap to extending an ongoing gap, and prefer a gap in seqx
 * to a gap in seq y.
 */
nw()
{
    char      *px, *py;          /* seqs and ptrs */
    int       *ndely, *dely;     /* keep track of dely */
    int       ndelx, delx;       /* keep track of delx */
    int       *tmp;              /* for swapping row0, row1 */
    int       mis;               /* score for each type */
    int       ins0, ins1;        /* insertion penalties */
    register  id;                /* diagonal index */
    register  ij;                /* jmp index */
    register  *col0, *coll;      /* score for curr, last row */
    register  xx, yy;            /* index into seqs */

    dx = (struct diag *)g_calloc("to get diags", len0+len1+1, sizeof(struct diag));

    ndely = (int *)g_calloc("to get ndely", len1+1, sizeof(int));
    dely = (int *)g_calloc("to get dely", len1+1, sizeof(int));
    col0 = (int *)g_calloc("to get col0", len1+1, sizeof(int));
    coll = (int *)g_calloc("to get coll", len1+1, sizeof(int));
    ins0 = (dna)? DINS0 : PINS0;
    ins1 = (dna)? DINS1 : PINS1;

    smax = -10000;
    if (endgaps) {
        for (col0[0] = dely[0] = -ins0, yy = 1; yy <= len1; yy++) {
            col0[yy] = dely[yy] = col0[yy-1] - ins1;
            ndely[yy] = yy;
        }
        col0[0] = 0;          /* Waterman Bull Math Biol 84 */
    }
    else
        for (yy = 1; yy <= len1; yy++)
            dely[yy] = -ins0;

    /* fill in match matrix
     */
    for (px = seqx[0], xx = 1; xx <= len0; px++, xx++) {
        /* initialize first entry in col
         */
        if (endgaps) {
            if (xx == 1)
                coll[0] = delx = -(ins0+ins1);
            else
                coll[0] = delx = col0[0] - ins1;
            ndelx = xx;
        }
        else {
            coll[0] = 0;
            delx = -ins0;
            ndelx = 0;
        }
        for (py = seqx[1], yy = 1; yy <= len1; py++, yy++) {
            mis = col0[yy-1];
            if (dna)
                mis += (xbm[px-'A']&xbm[py-'A'])? DMAT : DMIS;
            else
                mis += _day[px-'A'][py-'A'];
        }
    }
}

```

NW

nw.c

nw.c

...nw

```

/* update penalty for del in x seq;
 * favor new del over ongoing del
 * ignore MAXGAP if weighting endgaps
 */
if (endgaps || ndely[yy] < MAXGAP) {
    if (col0[yy] - ins0 >= dely[yy]) {
        dely[yy] = col0[yy] - (ins0+ins1);
        ndely[yy] = 1;
    } else {
        dely[yy] -= ins1;
        ndely[yy]++;
    }
} else {
    if (col0[yy] - (ins0+ins1) >= dely[yy]) {
        dely[yy] = col0[yy] - (ins0+ins1);
        ndely[yy] = 1;
    } else
        ndely[yy]++;
}

/* update penalty for del in y seq;
 * favor new del over ongoing del
 */
if (endgaps || ndelx < MAXGAP) {
    if (col1[yy-1] - ins0 >= delx) {
        delx = col1[yy-1] - (ins0+ins1);
        ndelx = 1;
    } else {
        delx -= ins1;
        ndelx++;
    }
} else {
    if (col1[yy-1] - (ins0+ins1) >= delx) {
        delx = col1[yy-1] - (ins0+ins1);
        ndelx = 1;
    } else
        ndelx++;
}

/* pick the maximum score; we're favoring
 * mis over any del and delx over dely
 */
id = xx - yy + len1 - 1;
if (mis >= delx && mis >= dely[yy])
    col1[yy] = mis;
else if (delx >= dely[yy]) {
    col1[yy] = delx;
    ij = dx[id].ljump;
    if (dx[id].jp.n[0] && (idna || (ndelx >= MAXJMP
    && xx > dx[id].jp.x[ij]+MX) || mis > dx[id].score+DINSO)) {
        dx[id].ljump++;
        if (++ij >= MAXJMP) {
            writejumps(id);
            ij = dx[id].ljump - 0;
            dx[id].offset = offset;
            offset += sizeof(struct jmp) + sizeof(offset);
        }
    }
    dx[id].jp.n[ij] = ndelx;
    dx[id].jp.x[ij] = xx;
    dx[id].score = delx;
} else {
    col1[yy] = dely[yy];
    ij = dx[id].ljump;

```

FIGURE 62F**nw.c****nw.c**

```

...nw
if (dx[id].jp.n[0] && (idna || (ndely[yy] >= MAXJMP
&& xx > dx[id].jp.x[ij]+MX) || mis > dx[id].score+DINS0)) {
    dx[id].ljmp++;
    if (++ij >= MAXJMP) {
        writejmps(id);
        ij = dx[id].ljmp = 0;
        dx[id].offset = offset;
        offset += sizeof(struct jmp) + sizeof(offset);
    }
    dx[id].jp.n[ij] = -ndely[yy];
    dx[id].jp.x[ij] = xx;
    dx[id].score = dely[yy];
}
if (xx == len0 && yy < len1) {
    /* last col
    */
    if (endgaps)
        coll[yy] -= ins0+ins1*(len1-yy);
    if (coll[yy] > smax) {
        smax = coll[yy];
        dmax = id;
    }
}
if (endgaps && xx < len0)
    coll[yy-1] -= ins0+ins1*(len0-xx);
if (coll[yy-1] > smax) {
    smax = coll[yy-1];
    dmax = id;
}
tmp = col0; col0 = coll; coll = tmp;
}
(void) free((char *)ndely);
(void) free((char *)dely);
(void) free((char *)col0);
(void) free((char *)coll);
}

```

FIGURE 62G

nwprint.c

nwprint.c

```
/* print() — only routine visible outside this module
 *
 * static:
 * getmat() — trace back best path, count matches: print()
 * pr_align() — print alignment of described in array p[]: print()
 * dumpblock() — dump a block of lines with numbers, stars: pr_align()
 * nums() — put out a number line: dumpblock()
 * putline() — put out a line (name, [num], seq, [num]): dumpblock()
 * stars() — put a line of stars: dumpblock()
 * stripname() — strip any path and prefix from a seqname
 */

#include "nw.h"

#define SPC 3
#define P_LINE 256 /* maximum output line */
#define P_SPC 3 /* space between name or num and seq */

extern day[26][26];
int olen; /* set output line length */
FILE *fx; /* output file */

print()
{
    int lx, ly, firstgap, lastgap; /* overlap */

    if ((fx = fopen(ofile, "w")) == 0) {
        fprintf(stderr, "%s: can't write %s\n", prog, ofile);
        cleanup(1);
    }
    fprintf(fx, "<first sequence: %s (length = %d)\n", namex[0], len0);
    fprintf(fx, "<second sequence: %s (length = %d)\n", namex[1], len1);
    olen = 60;
    lx = len0;
    ly = len1;
    firstgap = lastgap = 0;
    if (dmax < len1 - 1) { /* leading gap in x */
        pp[0].spc = firstgap = len1 - dmax - 1;
        ly -= pp[0].spc;
    }
    else if (dmax > len1 - 1) { /* leading gap in y */
        pp[1].spc = firstgap = dmax - (len1 - 1);
        lx -= pp[1].spc;
    }
    if (dmax0 < len0 - 1) { /* trailing gap in x */
        lastgap = len0 - dmax0 - 1;
        lx -= lastgap;
    }
    else if (dmax0 > len0 - 1) { /* trailing gap in y */
        lastgap = dmax0 - (len0 - 1);
        ly -= lastgap;
    }
    getmat(lx, ly, firstgap, lastgap);
    pr_align();
}
```

print

nwprint.c

nwprint.c

```
/*
 * trace back the best path, count matches
 */
```

```
static
```

```
getmat(lx, ly, firstgap, lastgap)
```

```
int lx, ly;
```

```
int firstgap, lastgap;
```

```
/* "core" (minus endgaps) */
```

```
/* leading trailing overlap */
```

getmat

```
{
```

```
int nm, i0, i1, siz0, siz1;
```

```
char outx[32];
```

```
double pct;
```

```
register n0, n1;
```

```
register char *p0, *p1;
```

```
/* get total matches, score
```

```
*/
```

```
i0 = i1 = siz0 = siz1 = 0;
```

```
p0 = seqx[0] + pp[1].spc;
```

```
p1 = seqx[1] + pp[0].spc;
```

```
n0 = pp[1].spc + 1;
```

```
n1 = pp[0].spc + 1;
```

```
nm = 0;
```

```
while ( *p0 && *p1 ) {
```

```
if (siz0) {
```

```
p1++;
```

```
n1++;
```

```
siz0--;
```

```
}
```

```
else if (siz1) {
```

```
p0++;
```

```
n0++;
```

```
siz1--;
```

```
}
```

```
else {
```

```
if (xbm[*p0-'A']&xbm[*p1-'A'])
```

```
nm++;
```

```
if (n0++ == pp[0].x[i0])
```

```
siz0 = pp[0].n[i0++];
```

```
if (n1++ == pp[1].x[i1])
```

```
siz1 = pp[1].n[i1++];
```

```
p0++;
```

```
p1++;
```

```
}
```

```
}
```

```
/* pct homology:
```

```
* if penalizing endgaps, base is the shorter seq
```

```
* else, knock off overhangs and take shorter core
```

```
*/
```

```
if (endgaps)
```

```
lx = (len0 < len1)? len0 : len1;
```

```
else
```

```
lx = (lx < ly)? lx : ly;
```

```
pct = 100.*(double)nm/(double)lx;
```

```
fprintf(fx, "\n");
```

```
fprintf(fx, "< %d match%s in an overlap of %d: %.2f percent similarity\n",
```

```
nm, (nm == 1)? "" : "es", lx, pct);
```

```
fprintf(fx, "< gaps in first sequence: %d", gapx);
```

```
if (gapx) {
```

```
(void) sprintf(outx, " (%d %s%s)",
```

```
ngapx, (dna)? "base":"residue", (ngapx == 1)? "" : "s");
```

```
fprintf(fx, "%s", outx);
```

nwprint.c

nwprint.c

...getmat

```

fprintf(fx, ". gaps in second sequence: %d", gapy);
if (gapy) {
    (void) sprintf(outx, " (%d %s%s)",
        ngapy, (dna)? "base":"residue", (ngapy -- 1)? "" : "s");
    fprintf(fx, "%s", outx);
}
if (dna)
    fprintf(fx,
        "\n<score: %d (match = %d, mismatch = %d, gap penalty = %d + %d per base)\n",
        smax, DMAT, DMIS, DINSO, DINS1);
else
    fprintf(fx,
        "\n<score: %d (Dayhoff PAM 250 matrix, gap penalty = %d + %d per residue)\n",
        smax, PINSO, PINS1);
if (endgaps)
    fprintf(fx,
        "<endgaps penalized. left endgap: %d %s%s, right endgap: %d %s%s\n",
        firstgap, (dna)? "base" : "residue", (firstgap -- 1)? "" : "s",
        lastgap, (dna)? "base" : "residue", (lastgap -- 1)? "" : "s");
else
    fprintf(fx, "<endgaps not penalized\n");
}

static      nm;           /* matches in core --- for checking */
static      lmax;         /* lengths of stripped file names */
static      ij[2];        /* jmp index for a path */
static      nc[2];        /* number at start of current line */
static      ni[2];        /* current elem number --- for gapping */
static      siz[2];
static char *ps[2];       /* ptr to current element */
static char *po[2];       /* ptr to next output char slot */
static char out[2][P_LINE]; /* output line */
static char star[P_LINE]; /* set by stars() */

/*
 * print alignment of described in struct path pp[]
 */
static
pr_align()
{
    int      nn;           /* char count */
    int      more;
    register i;

    for (i = 0, lmax = 0; i < 2; i++) {
        nn = stripname(nameex[i]);
        if (nn > lmax)
            lmax = nn;

        nc[i] = 1;
        ni[i] = 1;
        siz[i] = ij[i] = 0;
        ps[i] = seqx[i];
        po[i] = out[i];
    }

    for (nn = nm = 0, more = 1; more; ) {
        for (i = more = 0; i < 2; i++) {
            /*
             * do we have more of this sequence?
             */
            if (!*ps[i])
                continue;

            more++;
        }
    }
}

```

pr_align

nwprint.c

nwprint.c

...pr_align

```

if (pp[i].spc) { /* leading space */
    *po[i]++ = ' ';
    pp[i].spc--;
}
else if (siz[i]) { /* in a gap */
    *po[i]++ = '-';
    siz[i]--;
}
else { /* we're putting a seq element */
    *po[i] = *ps[i];
    if (islower(*ps[i]))
        *ps[i] = toupper(*ps[i]);
    po[i]++;
    ps[i]++;

    /*
     * are we at next gap for this seq?
     */
    if (nl[i] == pp[i].x[i]) {
        /*
         * we need to merge all gaps
         * at this location
         */
        siz[i] = pp[i].n[i]++;
        while (nl[i] == pp[i].x[i])
            siz[i] += pp[i].n[i]++;
    }
    nl[i]++;
}

if (++nn == olen || !more && nn) {
    dumpblock();
    for (i = 0; i < 2; i++)
        po[i] = out[i];
    nn = 0;
}

}

/*
 * dump a block of lines, including numbers, stars: pr_align0
 */
static
dumpblock()
{
    register i;

    for (i = 0; i < 2; i++)
        *po[i]-- = '\0';

    (void) putc('\n', fx);
    for (i = 0; i < 2; i++) {
        if (*out[i] && (*out[i] != ' ' || *(po[i]) != ' ')) {
            if (i == 0)
                nums(i);
            if (i == 0 && *out[1])
                stars();
            putline(i);
            if (i == 0 && *out[1])
                fprintf(fx, star);
            if (i == 1)
                nums(i);
        }
    }
}

```

dumpblock

nwprint.c

nwprint.c

...dumpblock

```

1
/*
 * put out a number line: dumpblock0
 */
static
nums(ix)
{
    int      ix;          /* Index in out[] holding seq line */

    char      nline[P_LINE];
    register  i, j;
    register  char *pn, *px, *py;

    for (pn = nline, i = 0; i < lmax+P_SPC; i++, pn++)
        *pn = ' ';
    for (i = nc[ix], py = out[ix]; *py; py++, pn++) {
        if (*py == '-' || *py == '+')
            *pn = *py;
        else {
            if (i%10 == 0 || (i == 1 && nc[ix] != 1)) {
                j = (i < 0)? -1 : 1;
                for (px = pn; j /= 10, px--)
                    *px = j%10 + '0';
                if (i < 0)
                    *px = '-';
            }
            else
                *pn = ' ';
            i++;
        }
    }
    *pn = '\0';
    nc[ix] = i;
    for (pn = nline; *pn; pn++)
        (void) putc(*pn, fx);
    (void) putc('\n', fx);
}

/*
 * put out a line (name, [num], seq, [num]): dumpblock0
 */
static
putline(ix)
{
    int      ix;

    int      i;
    register  char *px;

    for (px = name[ix], i = 0; *px && *px != '\0'; px++, i++)
        (void) putc(*px, fx);
    for (; i < lmax+P_SPC; i++)
        (void) putc(' ', fx);

    /* these count from 1:
     * nl[] is current element (from 1)
     * nc[] is number at start of current line
     */
    for (px = out[ix]; *px; px++)
        (void) putc(*px&0x7F, fx);
    (void) putc('\n', fx);
}

```

nums

putline

nwprint.c

nwprint.c

```

/*
 * put a line of stars (seqs always in out[0], out[1]): dumpblock0
 */
static
stars()
{
    int i;
    register char *p0, *p1, cx, *px;

    if (!(*out[0] || (*out[0] == ' ' && (*p0[0] == ' ')) ||
        !(*out[1] || (*out[1] == ' ' && (*p0[1] == ' '))))
        return;
    px = star;
    for (i = lmax+P_SPC; i; i--)
        *px++ = ' ';

    for (p0 = out[0], p1 = out[1]; *p0 && *p1; p0++, p1++) {
        if (isalpha(*p0) && isalpha(*p1)) {
            if (xbm[*p0-'A'] & xbm[*p1-'A']) {
                cx = '*';
                nm++;
            }
            else if (ldna && _day[*p0-'A'][*p1-'A'] > 0)
                cx = '.';
            else
                cx = ' ';
        }
        else
            cx = ' ';
        *px++ = cx;
    }
    *px++ = '\n';
    *px = '\0';
}

/*
 * strip path or prefix from pn, return len: pr_align0
 */
static
stripname(pn)
char *pn; /* file name (may be path) */
{
    register char *px, *py;

    py = 0;
    for (px = pn; *px; px++)
        if (*px == '/')
            py = px + 1;

    if (py)
        (void) strcpy(pn, py);
    return(strlen(pn));
}

```

stars

stripname

nwsubr.c

nwsubr.c

```

/* cleanup0 — cleanup any tmp file
 * getseq0 — read in seq, set dna, len, maxlen
 * g_calloc0 — calloc0 with error checkin
 * readjumps0 — get the good jumps, from tmp file if necessary
 * writejumps0 — write a filled array of jumps to a tmp file: nw0
 */
#include "nw.h"
#include <sys/file.h>

char    *jname = "/tmp/homgXXXXXX";    /* tmp file for jumps */
FILE    *fj;

int      cleanup();                    /* cleanup tmp file */
long     lseek();

/*
 * remove any tmp file if we blow
 */
cleanup(i)
int      i;
{
    if (fj)
        (void) unlink(jname);
    exit(i);
}

/*
 * read, return ptr to seq, set dna, len, maxlen
 * skip lines starting with ':', '<', or '>'
 * seq in upper or lower case
 */
char *
getseq(file, len)
char    *file;    /* file name */
int     *len;     /* seq len */
{
    char    line[1024], *pseq;
    register char    *px, *py;
    int     natgc, tlen;
    FILE    *fp;

    if ((fp = fopen(file, "r")) == 0) {
        fprintf(stderr, "%s: can't read %s\n", prog, file);
        exit(1);
    }
    tlen = natgc = 0;
    while (fgets(line, 1024, fp)) {
        if (*line == ':' || *line == '<' || *line == '>')
            continue;
        for (px = line; *px != '\n'; px++)
            if (isupper(*px) || islower(*px))
                tlen++;
    }
    if ((pseq = malloc((unsigned)(tlen+6))) == 0) {
        fprintf(stderr, "%s: malloc() failed to get %d bytes for %s\n", prog, tlen+6, file);
        exit(1);
    }
    pseq[0] = pseq[1] = pseq[2] = pseq[3] = '\0';

```

cleanup

getseq

nwsubr.c

nwsubr.c

...getseq

```

py = pseq + 4;
tlen = tlen;
rewind(fp);

while (fgets(line, 1024, fp)) {
    if (*line == ':' || *line == '<' || *line == '>')
        continue;
    for (px = line; *px != '\n'; px++) {
        if (isupper(*px))
            *py++ = *px;
        else if (islower(*px))
            *py++ = toupper(*px);
        if (index("ATGCU", *(py-1)))
            natgc++;
    }
    *py++ = '\0';
    *py = '\0';
    (void) fclose(fp);
    dna = natgc > (tlen/3);
    return(pseq+4);
}

```

```

char *
g_alloc(msg, nx, sz)
char *msg;          /* program, calling routine */
int nx, sz;         /* number and size of elements */
{
    char *px, *calloc();

    if ((px = calloc((unsigned)nx, (unsigned)sz)) == 0) {
        if (*msg) {
            fprintf(stderr, "%s: g_alloc() failed %s (n=%d, sz=%d)\n", prog, msg, nx, sz);
            exit(1);
        }
    }
    return(px);
}

```

g_alloc

/ get final jmps from dx[] or tmp file, set pp[], reset dmax: main0*

```

/*
readjmps()
{
    int fd = -1;
    int siz, io, il;
    register i, j, xx;

    if (fj) {
        (void) fclose(fj);
        if ((fd = open(jname, O_RDONLY, 0)) < 0) {
            fprintf(stderr, "%s: can't open() %s\n", prog, jname);
            cleanup(1);
        }
    }
    for (i = io = il = 0, dmax0 = dmax, xx = len0; ; i++) {
        while (1) {
            for (j = dx[dmax].jmp; j >= 0 && dx[dmax].jp.x[j] >= xx; j--)
                ;
            if (j < 0 && dx[dmax].offset && fj) {
                (void) lseek(fd, dx[dmax].offset, 0);
                (void) read(fd, (char *)&dx[dmax].jp, sizeof(struct jmp));
                (void) read(fd, (char *)&dx[dmax].offset, sizeof(dx[dmax].offset));
                dx[dmax].jmp = MAXJMP-1;
            }
        }
    }
}

```

readjmps

nwsubr.c

nwsubr.c

...readjumps

```

        else
            break;
    }
    if (i >= JMPS) {
        fprintf(stderr, "%s: too many gaps in alignment\n", prog);
        cleanup(1);
    }
    if (j >= 0) {
        siz = dx[dmax].jp.n[j];
        xx = dx[dmax].jp.x[j];
        dmax += siz;
        if (siz < 0) {
            /* gap in second seq */
            pp[1].n[i1] = -siz;
            xx += siz;

            /* ld = xx - yy + len1 - 1
            */
            pp[1].x[i1] = xx - dmax + len1 - 1;
            gapy++;
            ngapy -= siz;
        /* ignore MAXGAP when doing endgaps */
        siz = (-siz < MAXGAP || endgaps)? -siz : MAXGAP;
        i1++;

        }
        else if (siz > 0) {
            /* gap in first seq */
            pp[0].n[i0] = siz;
            pp[0].x[i0] = xx;
            gapx++;
            ngapx += siz;
        /* ignore MAXGAP when doing endgaps */
        siz = (siz < MAXGAP || endgaps)? siz : MAXGAP;
        i0++;

        }
    }
    else
        break;
}

/* reverse the order of jumps
*/
for (j = 0, i0--; j < i0; j++, i0--) {
    i = pp[0].n[j]; pp[0].n[j] = pp[0].n[i0]; pp[0].n[i0] = i;
    i = pp[0].x[j]; pp[0].x[j] = pp[0].x[i0]; pp[0].x[i0] = i;
}
for (j = 0, i1--; j < i1; j++, i1--) {
    i = pp[1].n[j]; pp[1].n[j] = pp[1].n[i1]; pp[1].n[i1] = i;
    i = pp[1].x[j]; pp[1].x[j] = pp[1].x[i1]; pp[1].x[i1] = i;
}
if (fd >= 0)
    (void) close(fd);
if (fj) {
    (void) unlink(jname);
    fj = 0;
    offset = 0;
}

}

/*
 * write a filled jmp struct offset of the prev one (if any): nw0
 */
writejumps(ix)
    int      ix;
{
    char      *mktemp();

```

writejumps

nwsubr.c

nwsubr.c

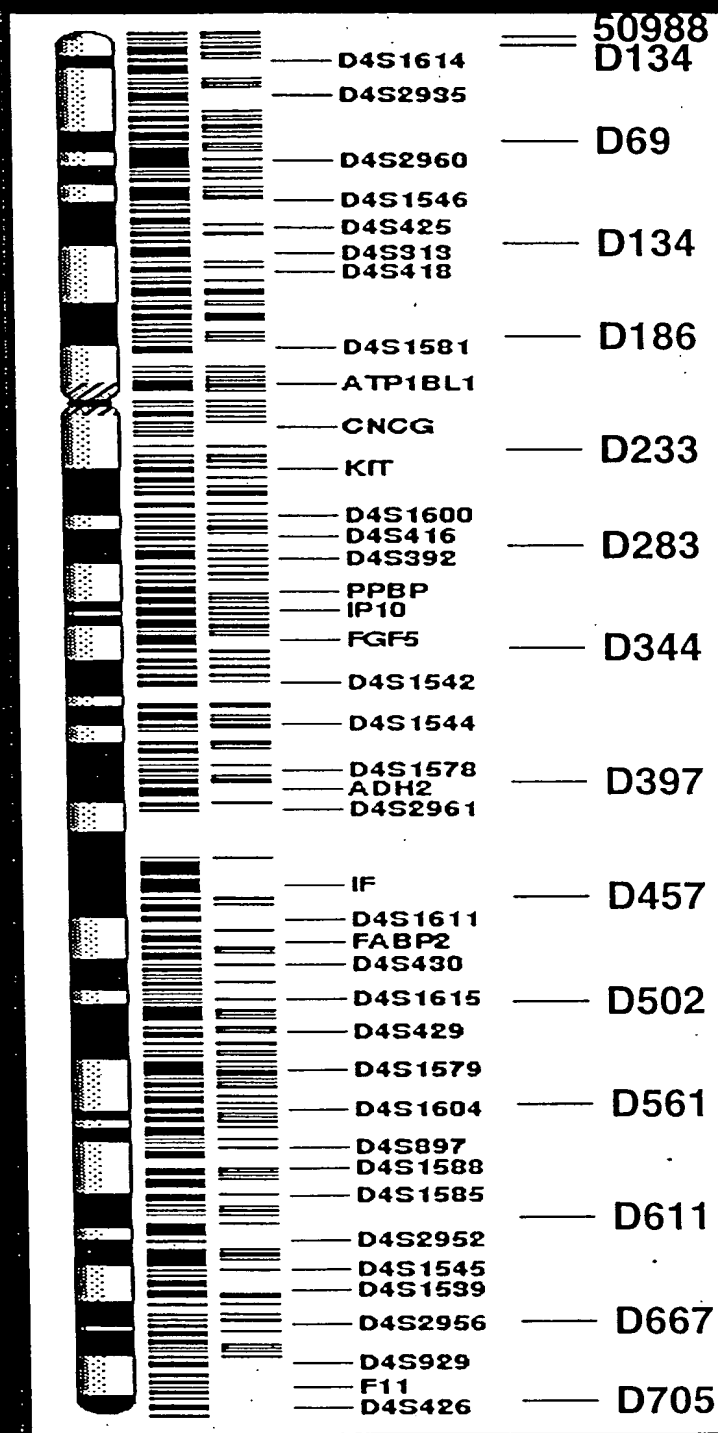
...writejmps

```

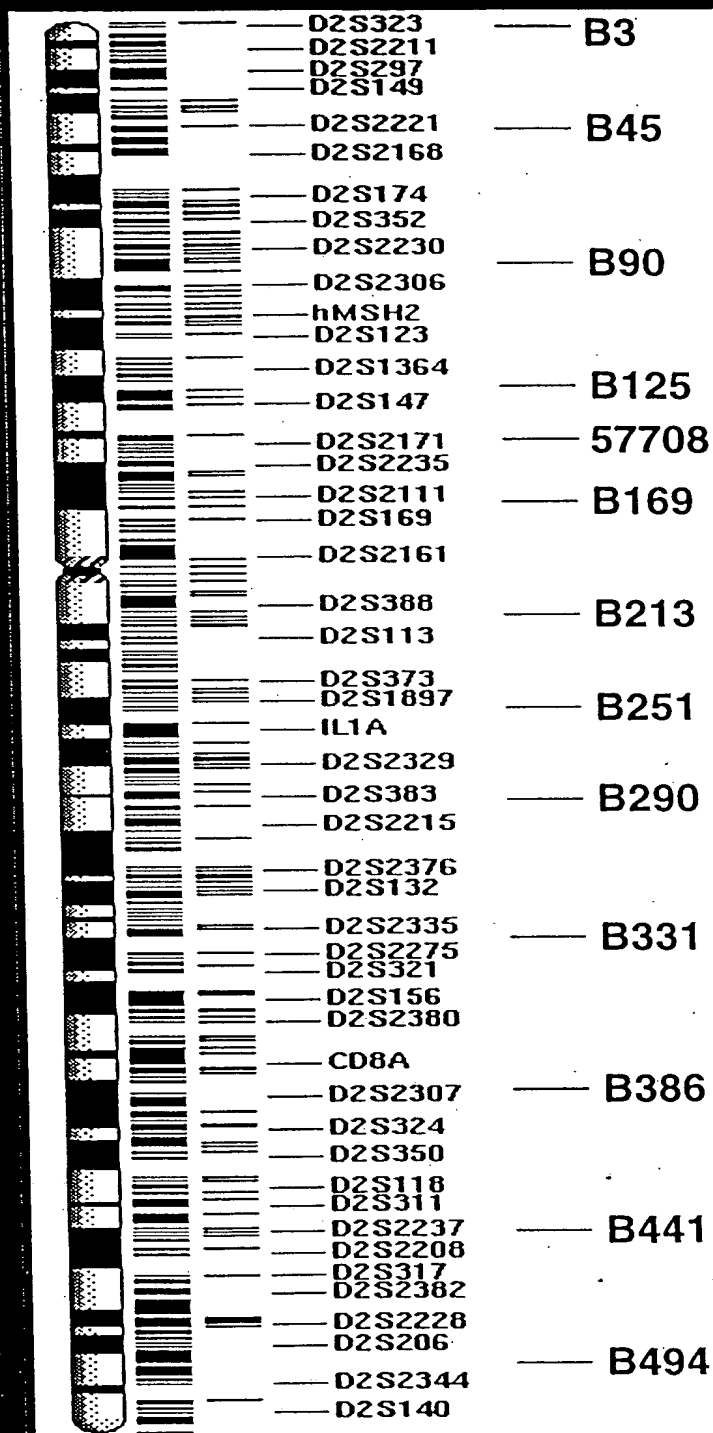
if (!fj) {
    if (mktemp(jname) < 0) {
        fprintf(stderr, "%s: can't mktemp() %s\n", prog, jname);
        cleanup(1);
    }
    if ((fj = fopen(jname, "w")) == 0) {
        fprintf(stderr, "%s: can't write %s\n", prog, jname);
        exit(1);
    }
    (void) fwrite((char *)&dx[ix].jp, sizeof(struct jmp), 1, fj);
    (void) fwrite((char *)&dx[ix].offset, sizeof(dx[ix].offset), 1, fj);
}

```

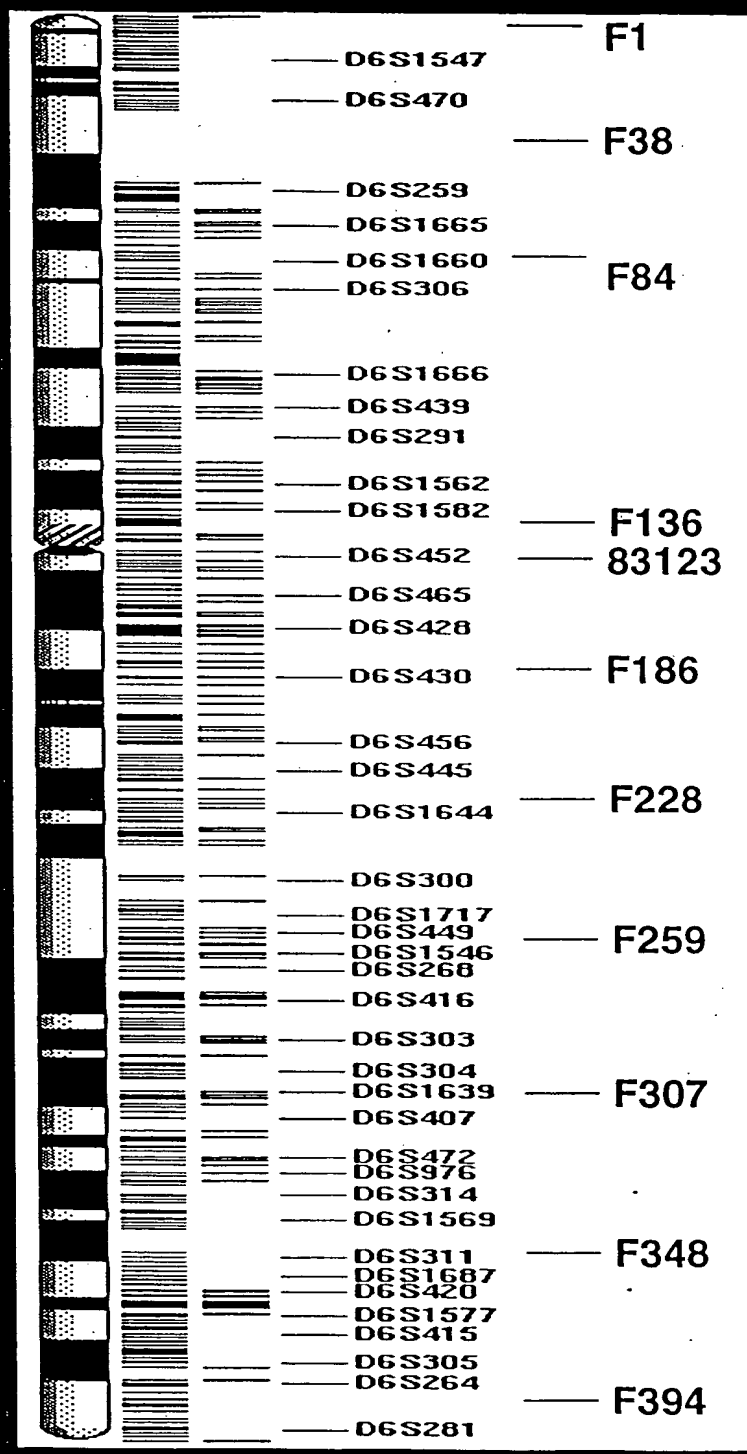
Chromosome 4



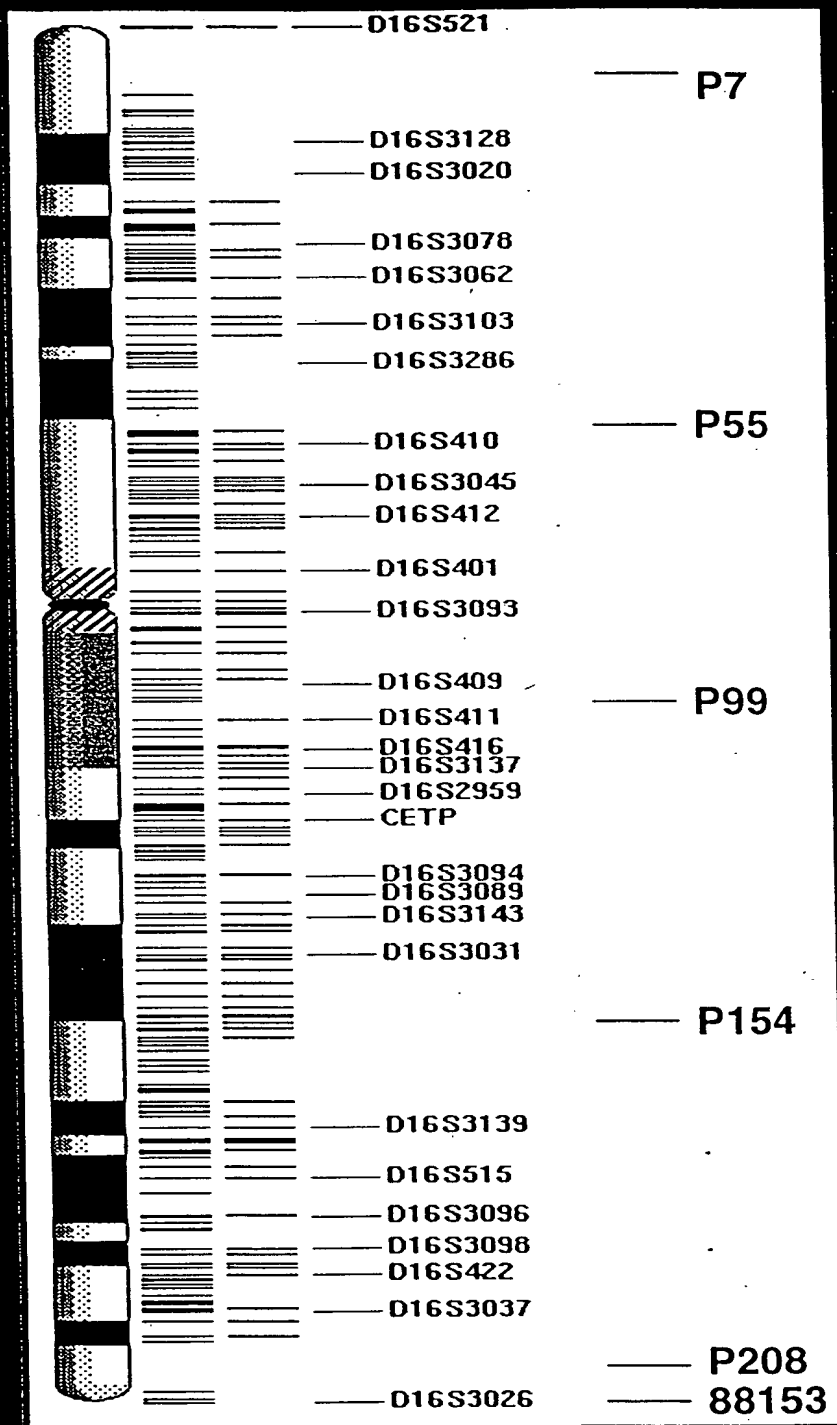
Chromosome 2



Chromosome 6



Chromosome 16



Inventor Information**GENE AMPLIFICATION BIN NO. 4**

- | | | |
|------|---------------|---|
| (1) | DNA26288-1239 | Kevin Baker; Audrey Goddard |
| (2) | DNA30942-1134 | Avi Ashkenazi; Austin Gurney; William Wood |
| (3) | DNA35680-1212 | Audrey Goddard; Austin Gurney |
| (4) | DNA49143-1429 | Colin Watanabe; William Wood |
| (5) | DNA49821-1562 | Colin Watanabe; William Wood; Audrey Goddard; Austin Gurney |
| (6) | DNA50988-1326 | William Wood |
| (7) | DNA52192-1369 | William Wood; Audrey Goddard; Austin Gurney |
| (8) | DNA56866-1342 | Colin Watanabe; William Wood |
| (9) | DNA57129-1413 | William Wood |
| (10) | DNA57708-1411 | Colin Watanabe; William Wood; Austin Gurney |
| (11) | DNA57836-1338 | William Wood; Audrey Goddard |
| (12) | DNA58721-1475 | William Wood; Austin Gurney |
| (13) | DNA59220-1514 | William Wood |
| (14) | DNA59485-1336 | William Wood; Colin Watanabe; Audrey Goddard |
| (15) | DNA59606-1471 | William Wood |
| (16) | DNA59622-1334 | William Wood |
| (17) | DNA59820-1549 | William Wood; Audrey Goddard |
| (18) | DNA59839-1461 | William Wood |
| (19) | DNA59841-1460 | Colin Watanabe; William Wood |
| (20) | DNA59842-1502 | William Wood |
| (21) | DNA59848-1512 | Colin Watanabe; William Wood |
| (22) | DNA62876-1517 | Colin Watanabe; William Wood |

Margaret Roy

Gene Amplification

Kenneth Hillan

in situ hybridization

alpha order:

Ashkenazi

Baker

Goddard

Gurney

Hillan

Roy

Watanabe

Wood